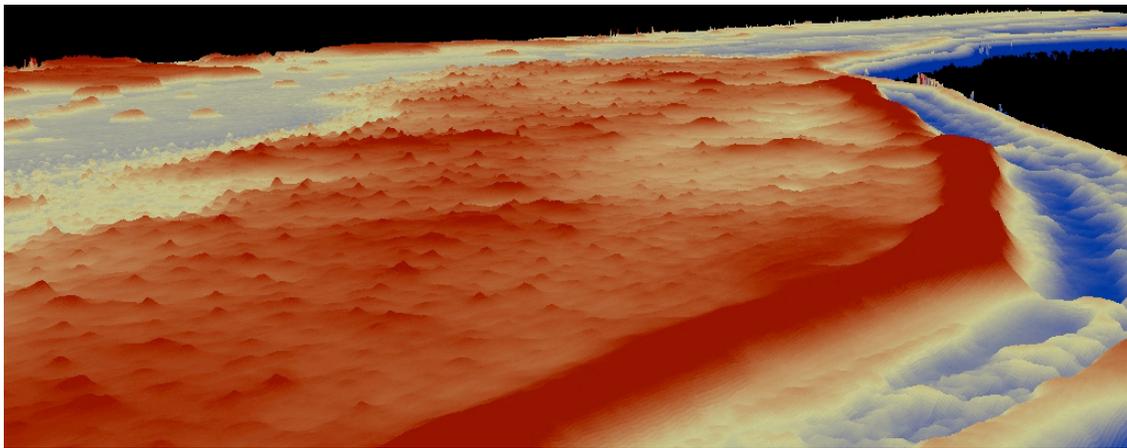


**CHARACTERIZATION OF DEEP WATER REEF COMMUNITIES WITHIN THE
MARINE CONSERVATION DISTRICT, ST. THOMAS, U.S. VIRGIN ISLANDS**

FINAL REPORT



Submitted to the Caribbean Fisheries Management Council

April 30st, 2008

Prepared by
Richard S. Nemeth, Tyler B. Smith, Jeremiah Blondeau, Elizabeth Kadison,
Jacquelyn M. Calnan, and Jordan Gass

Center for Marine and Environmental Studies
University of the Virgin Islands



<http://marsci.uvi.edu/>

TABLE OF CONTENTS

Table of Contents	I
List of Tables	III
List of Figures	IV
List of Appendices	VII
Executive Summary	1
Introduction	3
Background.....	3
Methods	8
Habitat Stratification and Sampling Design	8
In Situ Sampling Protocols	9
Benthic Composition	9
Coral Health	10
Fish and Other Commercially Important Motile Resources.....	11
Results and recommendations	12
Benthic Composition	12
Scleractinian Corals and Hydrocorals.....	15
Gorgonians and Sponges	18
Algae	19
Non-living Benthic Cover	21
Benthic Composition by Sampling Strata	23
Benthic Epifauna	23
Algae	24
Non-living Benthic Composition	24
Predicted Versus Sampled Benthic Strata	27
Algae.....	27
Coral	27
Pavement	28
Sand	28
Coral Health.....	31
Disease and Bleaching	31
Partial Mortality	33
Unknown Necrosis.....	34
Assemblage Structure of Fish and Motile Mega-Invertebrates.....	39
Diversity, Abundance and Biomass of Reef Fishes	39
Trophic composition among habitat strata.....	49

Size structure of Reef Fishes	51
Conclusions and Recommendations	59
Improved Evaluation of EFH within the MCD	59
Expansion of the Evaluation of EFH Outside the MCD	60
ACKNOWLEDGEMENTS	61
LITERATURE CITED	62
APPENDICES	64

LIST OF TABLES

Table 1. Species known to spawn or form aggregations around the red hind spawning aggregation site (18.202 N, 65.002 W).....	5
Table 2. The ranked percent coverage and percent of total coral cover for scleractinian coral and hydrocoral species recorded in video surveys.	15
Table 3. A list of known scleractinian coral and hydrocoral species found within the Marine Conservation District. “X” indicates that the coral was found in this study and/or in the Territorial Coral Reef Monitoring Program. “o” indicates that these species were likely encountered in this study, but could not be distinguished in the field.	17
Table 4. Species of scleractinian coral affected by unknown necrosis, and the prevalence, severity, and sample size among affected species in the Marine Conservation District.	34
Table 5. Abundance, size and biomass of commercially important species of grouper and snappers in 80 fish transects from all habitat strata combined in the Marine Conservation District.	53
Table 6. Species observed during roving dives in the MCD at fixed sites (sampled 2005-2007) and on four designated habitat types sampled during the MCD survey. Fixed sites, RHB and CSE, are both coral reef habitat. Designated habitat types on the MCD survey include algae (A), coral (C), pavement (P) and sand (S). Abundance categories are 0 (no fish), 1(one fish), 2 (2-10 fish), 3 (11-100 fish) and 4 (101-1000 fish).....	54

LIST OF FIGURES

Figure 1. Map of the wider Caribbean (top panel), the shelf area of the Puerto Rican Shelf, U.S. Caribbean, with the Red Hind Marine Conservation District (MCD) indicated (yellow polygon) (middle panel), bathymetry of the MCD (lower panel). Permanent coral reef monitoring locations are College Shoal East (CSE) and Hind Bank (HB)..... 6

Figure 2. A) MCD multibeam bathymetry data at 1m resolution, inset shows bathymetry detail. B) MCD habitat map indicating the four strata (algae, coral, pavement and sand) used in study design, inset shows habitat detail. Maps created with GIS using data provided by Moody 2003 and Prada 2003. . 7

Figure 3. Representative photos of distinct habitat types sampled within the Marine Conservation District and Corresponding to A) Algae, B) Coral, (following page) C) Pavement, and D) Sand..... 13

Figure 4. Percent coverage of A) all coral species and B) *Montastraea annularis* species complex in the Marine Conservation District. 16

Figure 5. Percent coverage of A) gorgonians and B) sponges in the Marine Conservation District. 18

Figure 6. Percent coverage of A) macroalgae, B) *Schizothrix* spp., (following pages) C) *Lobophora variegata*, D) *Dictyota* spp., E) dead coral covered with turf algae, and F) crustose coralline algae in the Marine Conservation District. 19

Figure 7. Percent coverage of A) all non-living substrata, (following page) B) sand/sediment, C) pavement, and D) rubble in the Marine Conservation District. 21

Figure 8. Depths of sampling locations among four predicted strata of the Marine Conservation District. Figure components are mean (thick black line), median (thin black line), 25th and 75th percentiles (bottom and top of box, respectively), 10th and 90th percentiles (bottom and top whiskers, respectively) and values outside 10th and 90th percentile (dots). Homogeneous subsets of means are indicated with letters..... 23

Figure 9. The percent coverage (\pm SE) of the benthic fauna categories coral, gorgonians, and sponges among the predicted benthic strata (N = 80). Homogeneous subsets of means are indicated with letters (Tukey HSD Post Hoc comparison)..... 25

Figure 10. . The percent coverage (\pm SE) of the benthic algae categories total algae, total macroalgae, the filamentous cyanobacteria *Schizothrix* spp., and the phaeophyte *Lobophora variegata* among the predicted benthic strata (N = 80). Homogeneous subsets of means are indicated with letters (Tukey HSD Post Hoc comparison)..... 25

Figure 11. The percent coverage (\pm SE) of the benthic algae categories the phaeophyte *Dictyota* spp., dead coral covered with turf algae, and crustose coralline algae among the predicted benthic strata (N = 80). Homogeneous subsets of means are indicated with letters (Tukey HSD Post Hoc comparison).26

Figure 12. The percent coverage (\pm SE) of the abiotic benthic categories total non-living substrata, sand/sediment, pavement, and rubble among the predicted benthic strata (N = 80). Homogeneous subsets of means are indicated with letters (Tukey HSD Post Hoc comparison). 26

Figure 13. Proportionate benthic composition determined from in situ surveys across four general benthic strata determined from side-scan sonar processing: algae, coral, pavement, sand. In situ data within each benthic strata were apportioned into hard substrate, soft substrate, and variable substrate categories with proportion of total (%) in brackets. 30

Figure 14. Prevalence and severity of A) total disease, (following page) B) unknown necrosis, C) white plague, and D) bleaching on hard corals of the Marine Conservation District. 31

Figure 15. Prevalence and Severity of A) recent mortality, and B) old mortality on hard corals in the Marine Conservation District. 33

Figure 16. Photographic examples of coral disease in the Marine Conservation District: A) a coral colony affected by white plague (site: coral 58), B) corals affected by unknown necrosis. (site: Pavement 119, October 9, 2007). 36

Figure 17. Photographic examples of coral disease in the Marine Conservation District: A) intercostal necrosis B) close-up of intercostal necrosis (site: pavement 111, October 29, 2007). 37

Figure 18. Photographic examples of coral disease in the Marine Conservation District: A) intercostals necrosis grading to general necrosis B) close-up of intercostal necrosis grading to general necrosis (site: pavement 111, October 9, 2007) 38

Figure 19. Mean species richness (\pm SE) for each habitat strata in the Marine Conservation District. 39

Figure 20. Family richness of A) Scaridae, B) Serranidae, C) Lutjanidae, and D) Haemulidae at each sampling location in the Marine Conservation District. 40

Figure 21. Total number of the most abundant species observed in belt transects in each habitat strata of the Marine Conservation District. Species were included if cumulative number in all habitats was ≥ 5 and then ordered from the most to least abundant from the coral habitat. 42

Figure 22. Abundance (ind. /100m²) of commercially important species (*E. guttatus*, *M. venenosa*, *E. striatus*, *L. analis*, *O. chrysurus* and *B. vetula*) at each sampling location in the Marine Conservation District. 43

Figure 23. Family abundance (ind. /100m²) of A) Acanthuridae, B) Scaridae, C) Haemulidae, D) Serranidae, E) Lutjanidae, and F) Balistidae at each sampling location in the Marine Conservation District. 44

Figure 24. *E. guttatus* A) abundance (ind. /100m²) and B) biomass (g/100m²) at each sampling location in the Marine Conservation District. 46

Figure 25. Mean biomass (SE bars) for all fish at each sampling location in the Marine Conservation District. 46

Figure 26. Family biomass (g/100m²) of A) Acanthuridae, (following pages) B) Scaridae, C) Haemulidae, D) Serranidae, E) Lutjanidae, and F) Balistidae at each sampling location in the Marine Conservation District. 47

Figure 27. . Trophic level abundance (ind. /100m²) of A) herbivores, B) invertivores, C) spongivores, D) piscivores, E) planktivores, and F) omnivores at each sampling location in the Marine Conservation District. 50

Figure 28. Length frequency histograms for selected commercially important families (Acanthuridae and Scaridae) and species (*O. chrysurus*, *E. guttatus* and *B. vetula*) in all habitat strata combined in the Marine Conservation District.. 52

LIST OF APPENDICES

Appendix I	Sampling locations in the Marine Conservation District and their general characteristics....	64
Appendix II	Electronic supplement (DVD). Doted images used in benthic composition assessments..	66
Appendix III	Electronic supplement (DVD). Video captures of each sampling location.....	66
Appendix IV	Video mosaic images of representative coral reef locations from the strata coral (Coral 52) and sand (Sand 166). Scaling quadrat is 25 X 25 cm. Video mosaic analysis courtesy of A. Gleason and P. Reid (RSMAS-University of Miami).....	67
Appendix V	Electronic supplement (DVD). Benthic cover data for each location sampled in the Marine Conservation District.	69
Appendix VI	Electronic supplement (DVD). Coral health data for coral harboring locations sampled in the Marine Conservation District.	69
Appendix VII	Electronic supplement (DVD). Fish data for each location sampled in the Marine Conservation District.	69
Appendix VIII	Literature Review: literature pertinent to the Marine Conservation District	69

EXECUTIVE SUMMARY

The University of the Virgin Islands, in collaboration with the Caribbean Fisheries Management Council, completed a survey of the habitat and fisheries resources of the Red Hind Marine Conservation District (MCD), St Thomas, United States Virgin Islands. The purpose of this research was to validate habitat classifications developed for the CFMC and to assess fisheries and non-fisheries resources within this marine protected area. This research provides information that is applicable for the classification and ranking of essential fish habitat (EFH) within the MCD and similar mesophotic reef habitat (30 – 50 m) along the Puerto Rican Shelf.

Benthic habitat assessments revealed extensive and well developed mesophotic coral reefs at depths of 34 – 47 m. Coral reefs were determined to occupy 65% of sites sampled in the MCD (coral cover > 4%, N = 80), with an average coral coverage of $25.3\% \pm 2.1$ SE and maximum coral coverage of 50.1%. Coral species richness was high, with 37 species or genera recorded from the MCD, including the threatened elkhorn coral (*Acropora cervicornis*). Coral coverage was dominated (91.8%) by members of the *Montastraea annularis* species complex.

Benthic habitats were predicted with variable accuracy using classified sonar imagery. Coral reefs were found to occupy almost all sampling strata predicted to contain pavement and sand habitat types. However, algae and coral strata were well predicted.

Coral health assessments revealed an extensive and severe cryptic coral mortality event caused by an unknown disease referred to as unknown necrosis. Disease signs and mortality covered a coherent region comprising over one fifth of the area of the MCD. The mean prevalence ($42.4\% \pm 6.3$ SE, N = 27) and severity ($32.8\% \pm 4.6$ SE) of unknown necrosis at affected sites suggested that the effects of the disease were intense in these areas, and may have contributed to a loss of over half the coral coverage.

Motile resource surveys of fish and commercially important invertebrates showed a total of 112 fish species. No motile macro invertebrates were seen during our surveys, but previous studies within the MCD have documented spiny lobster (*Panulirus argus*) and channel crabs (*Mithrax spinosissimus*). Species richness and biomass was highest in coral habitats followed by sand and pavement. Species richness was significantly lower in algal habitats than in the three other habitat types. Fish assemblage structure was dominated numerically by herbivorous and planktivorous species. Greatest fish biomass was found among invertivores, herbivores, and piscivores, respectively. Commercially important species were found primarily in coral and hard-bottom habitats. The fact that fish communities in the sand stratum were similar to coral and pavement strata indicates the discrepancy in habitat classification. True sand habitats contained a fish assemblage structure more similar to algal plains. Abundance and distribution of reef

fishes varied throughout the MCD depending upon taxa and presence of habitat types within the MCD. For example, the queen triggerfish (*Balistes vetula*) showed higher biomass near the northern boundary and in the eastern end of the MCD. These portions of the MCD contained large, sandy areas that are the preferred habitat of *B. vetula*.

INTRODUCTION

The Sustainable Fisheries Act of 1996 provided for significant changes in the management of fishery resources. In particular, it created the concept of an essential fish habitat (EFH) and required that scientific research be undertaken to determine habitats that were critically important to maintain fish stocks. Defined as “those waters and substrate necessary to fish for spawning, breeding, feeding or growth to maturity” this amendment to the Magnuson-Stevens Act requires a broader assessment of the habitats and locations that should be afforded protection.

To address threats of over fishing and provide additional protective measures to essential fish habitat, the Caribbean Fishery Management Council (CFMC) initiated a study of one of the most significant marine protected areas within the US Virgin Islands, the Red Hind Bank Marine Conservation District (MCD; Fig. 1, Fig. 2). Established in 1999, regulations within the MCD prohibit anchoring and fishing of any kind, except trolling for pelagic species (Federal Register 64:213). The MCD encompasses 44.5 square kilometers (39.5 square kilometer < 50 m depth) of federal waters at the edge of the Puerto Rican Shelf south of St. Thomas.

Although a number of studies have been conducted within the MCD (Beets and Friedlander 1999, Nemeth 2004, Whiteman et al 2005, Nemeth and Quandt 2005, Herzlieb et al. 2006) there remain significant gaps in knowledge about the fish and benthic resources within its boundaries. For example, little is known about deepwater shelf slope and shelf edge benthic communities nor the status of the fish stocks that utilize these habitats.

Characterization and assessment of benthic habitats and associated communities will provide valuable data to inform the development and/or revision of fishery management plans based on the principles expressed by the EFH concept. This information can be used to understand the necessity of future marine protected areas and guide their designation. The specific objectives of the research presented in this report were to characterize benthic habitat composition across the MCD, provide ground validation to support existing GIS habitat maps of the MCD, assess the health of coral resources, and quantitatively describe the associated fish and fisheries resources. This report fills in gaps of previous research within the MCD and provides baseline characterization data that will allow greater assessment and management of EFH within the MCD and across similar mesophotic reefs within the U.S. Caribbean.

Background

Studies conducted in the MCD have indicated that the area includes numerous deep habitats (30-100+ m) that contain resources important to fisheries dependent economies and regional biodiversity. A detailed

bathymetric and habitat characterization of the MCD was conducted in 2003 (Moody 2003; Prada 2003, Rivera) using multibeam sonar and Side Scan Sonar (SSS) imagery. These studies delineated three Meta-communities including corals and gorgonians, submerged aquatic vegetation, and sand. Within these Meta-communities 23 habitat types were classified.

Direct visual surveys of sessile benthic communities within the MCD have suggested the presence of exceptionally rich and extensive mesophotic coral reef communities. Autonomous underwater vehicle (AUV) benthic surveys (Armstrong et al. 2006) and in situ assessments and monitoring (Nemeth et al. 2005, Herzlieb et al. 2006, Smith in review) have revealed coral reef banks and patch reefs dominated by reef forming corals of the *Montastraea annularis* species complex. These surveys also have shown that coral cover is typically higher (10-50%) and coral health greater than on shallow and midshelf reefs in the USVI and wider Caribbean (Gardner et al. 2003). Coral cover and health at two locations within the MCD (Hind Bank and College Shoal East) have been under semi-annual to annual benthic monitoring since 2003 by the USVI Territorial Monitoring Program (Figure 1, lower panel; Nemeth et al. 2006, Smith et al. 2007). These detailed studies of benthic composition and trajectory, and coral health, have shown that reefs within the MCD may be partially buffered from the effects of climate change through mechanisms that include reduced light intensity and moderation of temperature as the result of upwelling along the shelf edge (Smith et al., in prep). Such studies suggest that the MCD may be a regionally important refuge for coral reef biodiversity and fish habitat under scenarios of future seawater warming in the Western Atlantic (Donner et al. 2007).

The MCD is likely to protect a large biomass of resident and transient commercially important fishes. Fish and fisheries resources in the MCD are broadly similar to shallow waters of the Puerto Rican Shelf, but differ from shallow and midshelf coral reefs in relative species composition and the occurrence of more rare deep water associated fishes (Nemeth et al. 2006). An important red hind spawning aggregation site (SPAG) within the MCD is well characterized (Olsen and Laplace 1979, Beets and Friedlander 1999, Nemeth 2005) and was the initial stimulus for establishment of the management area along the shelf edge. Historical SPAGs of the federally protected Nassau grouper (*Epinephelus striatus*) and existing or extant spawning aggregations of other commercially important species are also known from the MCD (Olsen and LaPlace 1979, Nemeth unpub data, Table 1). In addition, the Grammanik Bank, a 1.4 km reef less than 5 km east of the MCD boundary, hosts spawning aggregations of Nassau, yellowfin (*Mycteroperca venenosa*), yellowmouth (*M. interstitialis*) and tiger (*M. tigris*) grouper (Nemeth et al. 2006b). Recent hydro-acoustic data from Nassau and yellowfin grouper tagged on the Grammanik Bank indicate that many move west into the MCD between monthly spawning events in February, March and April, and that the MCD affords protection to those species from fishing during a very critical period of the year (Nemeth, unpub. data).

Table 1. Species known to spawn or form aggregations around the red hind spawning aggregation site (18.202 N, 65.002 W).

Common Name	Scientific Name	Number observed	Timing
Red hind	<i>Epinephelus guttatus</i>	80,000	Dec-Feb
Nassau grouper	<i>E. striatus</i>	Historic	Dec-Feb
Tiger grouper	<i>Mycteroperca tigris</i>	100	Jan-Mar
Mutton snapper	<i>Lutjanus analis</i>	200	Mar-May
Schoolmaster snapper	<i>L. apodus</i>	100	Mar-May
Horse-eye jack	<i>Caranx latus</i>	300	Feb-Apr
Black jack	<i>Caranx lugubris</i>	500	April

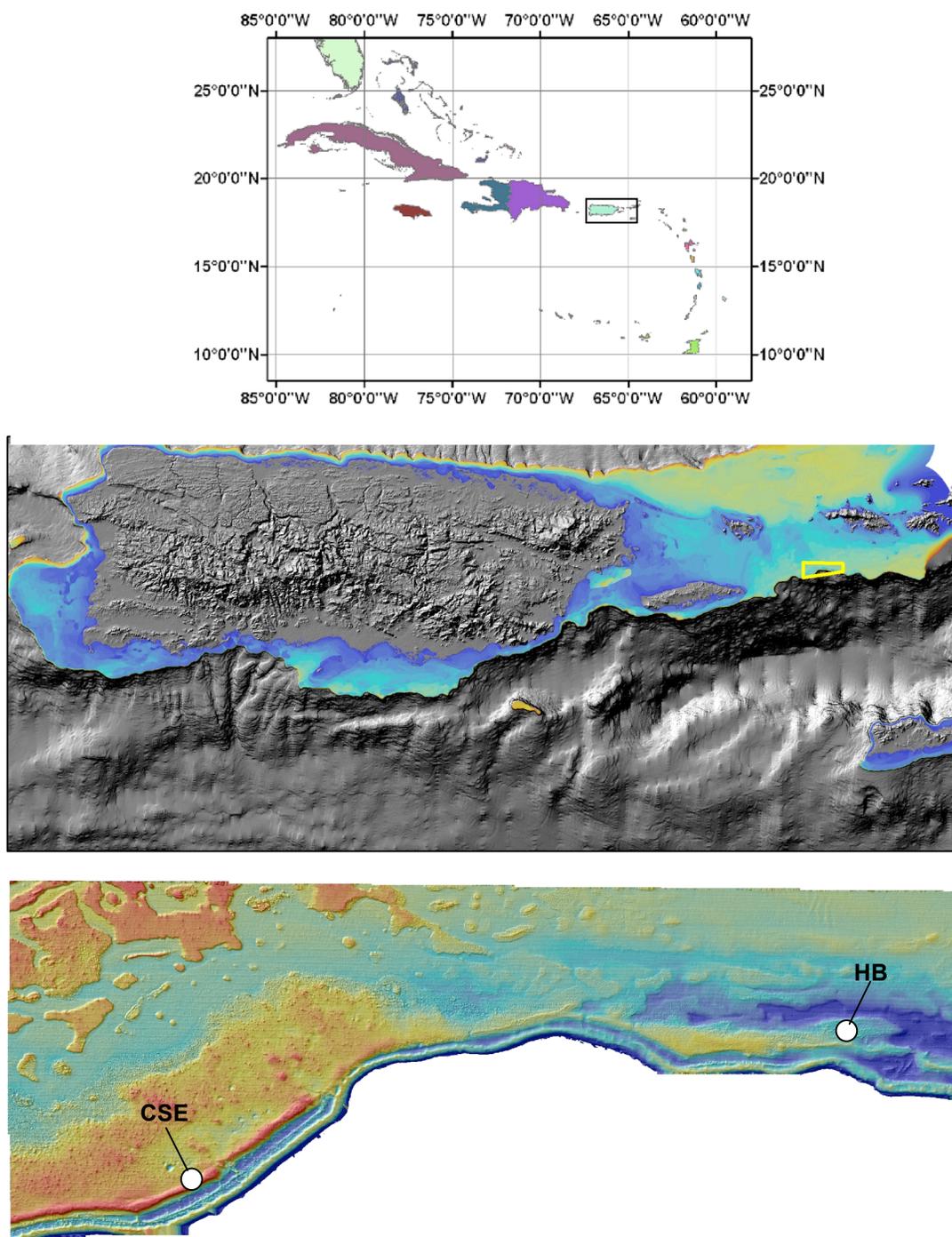


Figure 1. Map of the wider Caribbean (top panel), the shelf area of the Puerto Rican Shelf, U.S. Caribbean, with the Red Hind Marine Conservation District (MCD) indicated (yellow polygon) (middle panel), bathymetry of the MCD (lower panel). Permanent coral reef monitoring locations are College Shoal East (CSE) and Hind Bank (HB).

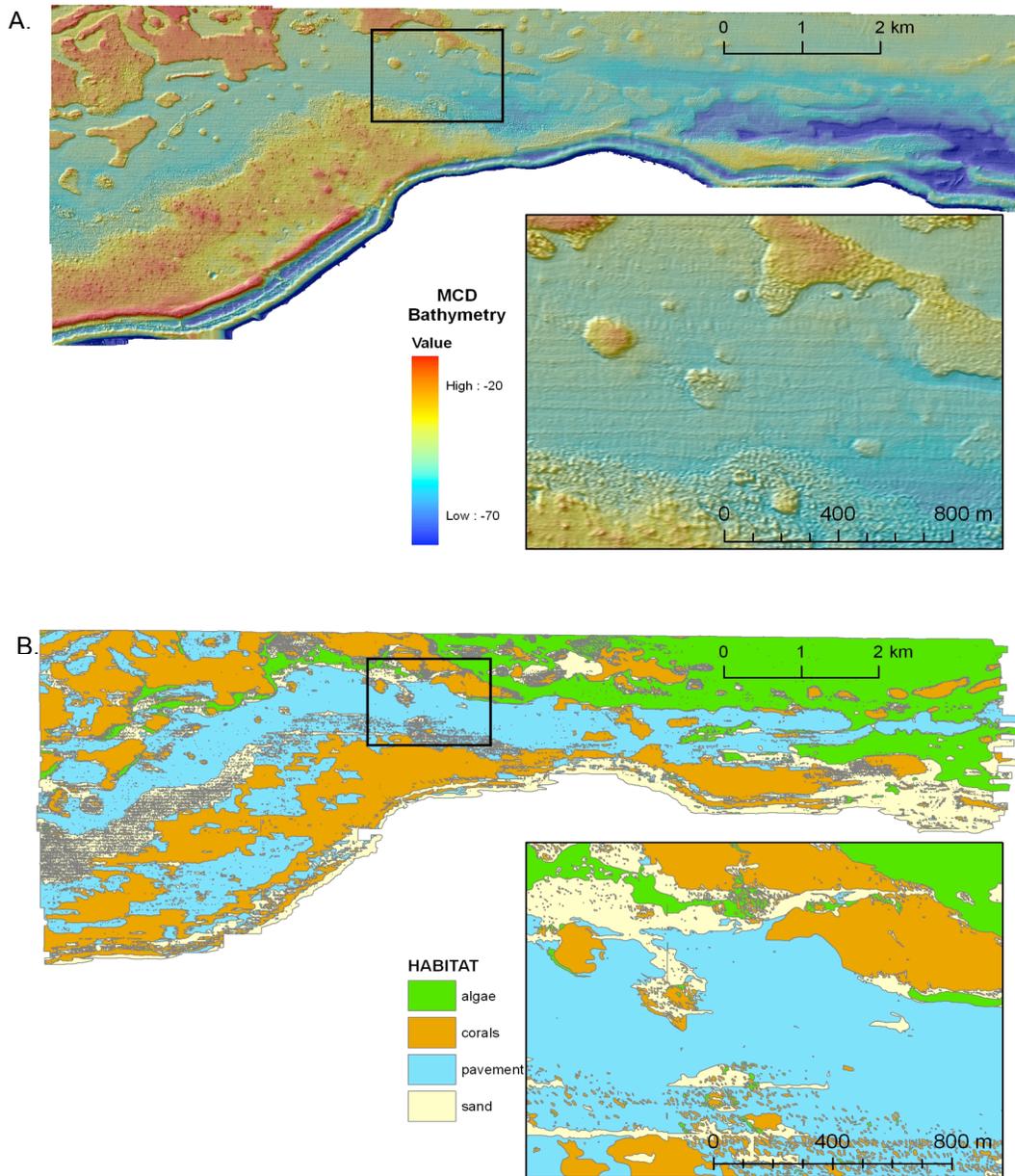


Figure 2. A) MCD multibeam bathymetry data at 1m resolution, inset shows bathymetry detail. B) MCD habitat map indicating the four strata (algae, coral, pavement and sand) used in study design, inset shows habitat detail. Maps created with GIS using data provided by Moody 2003 and Prada 2003.

METHODS

Habitat Stratification and Sampling Design

The ecosystem-based assessment of the Red Hind Marine Conservation District (MCD) presented in this report was based on a stratified random sampling strategy. Stratified random assessments offer a robust appraisal of natural resources with minimized bias (Menza et al. 2006). They allow for statistical assessment of sampling sufficiency for variables of interest because they are minimally biased and are amenable to predictive scaling of data, such as fish biomass and benthic composition, to larger geographic regions. Assignment of sampling sites within the MCD was based on predicted habitat structure defined in GIS products produced by Prada 2003 (Figure 1B). Habitat designations by Prada (2003) were determined using modifications of the scheme produced by NOAA for shallow waters of Puerto Rico and U.S. Virgin Islands (NOAA-NOS 2001). Four predicted habitat designations were chosen for sampling and included: coral, pavement, sand, and algae. These designations encompassed all benthic areas of the MCD less than 50 m. The choice of these four general habitat strata, as opposed to more specific habitat designations that were available from GIS products, was two-fold. First, because of uncertainties in true habitat composition from the assignment algorithms developed for shallow-water Caribbean benthos, it was decided that broad categories offered the greatest possibility of locating in situ assessments within accurately predicted habitats. Second, because of limitations in diver-based sampling effort in deep (30-50 m) benthic areas (i.e., 10 – 25 min. safe repetitive bottom times and the allocated study budget), it was predetermined that diver pairs could accomplish a robust number of surveys per stratum by using only four strata (80 total surveys). With these constraints, and with the lack of a priori information on habitat structure and motile resources, it was predicted that 20 surveys per predicted habitat strata would be required to make reliable assessments.

Allocation of sampling effort within each predicted benthic strata was accomplished by randomized assignment of sampling locations using the Geographic Information System (GIS). Using the four predicted benthic habitat strata supplied in GIS format, gridlines were removed and all adjacent polygons were aggregated using the Dissolve tool in ArcGIS 9.2 (ESRI software) and clipped to remove habitat defined outside the MCD boundary. To exclude benthic sampling units smaller than the size of sampling surveys (25 m linear distance, see below), any polygons in the resulting file that had areas less than 625 m² were removed from the shapefile. Hawth's Tools extension for ArcGIS was then used to generate 200 random points, with 50 points allocated for each habitat type. Generated sampling points (locations) were constrained so that points could not lie within 25 meters of any other sampling point. This was done to avoid the possibility of re-sampling. Sampling units were also allocated to a maximum depth of 50 m, which was considered the maximum safe depth for repetitive dives following the sampling protocol. Sampling sites and their general characteristics are presented in Appendix I.

In Situ Sampling Protocols

Randomly determined points were used as survey locations for in situ sampling protocols. Within each of the 50 random sampling points, the first 20 points were used as the drop point for diver pairs. Subsequent sampling points (randomly generated points 21-50) in each habitat strata were sequentially used if ship-board depth sounder measurements or initial diver reconnaissance of the sampling point determined that benthic areas were unsafe for completion of sampling protocols. This resulted in four randomly replaced surveys. Diver pairs utilizing technical NITROX or closed circuit rebreather were launched on the water surface within ten meters of the designated sampling point using ship-board GPS. Divers descended directly downwards on the sampling point. It was estimated that actual benthic sampling areas deviated no further than 25 meters from the predetermined sampling point of each survey.

The sampling protocol was designed to assess benthic composition, coral health, and the abundance of fish and other motile resources. Upon reaching the seafloor diver pairs deployed 30 m transect line along a compass direction randomly determined a priori using the function $RAND()*360$ in Microsoft Excel. Each diver pair had divided responsibility for resource assessments. Diver 1 deployed the transect tape while assessing fish and other motile resources. Following Diver 1, Diver 2 assessed the health of coral resources along the transect line. After Diver 1 had deployed the transect tape, Diver 1 returned along the transect line and recorded the benthos using digital video. Detailed methods for assessment of benthic composition, coral health, fish, and other motile resources are described below.

Benthic Composition

Benthic composition was recorded along a transect using standardized video monitoring protocols (Aronson et al. 1994, Carleton and Done 1995, Rogers and Miller 2001, Rogers et al. 2001). The video monitoring protocol is used in numerous coral reef benthic sampling programs, including the USVI Territorial Coral Reef Monitoring Program (Nemeth et al. 2006a, Smith et al. 2007). This protocol maximizes assessment times underwater and is particularly useful in time-limited assessments, such as in deep diving conditions. During video sampling, a diver swam at a uniform speed (~5 min. per transect) recording the benthic cover using a Sony TRV-950 digital camcorder in a Light and Motion Stingray II underwater housing. The diver pointed the camera down and perpendicular to the substrata remaining approximately 0.4 m about the substrate at all times. A guide wand attached to the camera housing was used to help the diver maintain the camera at a constant distance as they followed the vertical contour of the substratum. The total length of the transect taped by the diver was variable but ranged between a minimum of 10m to a max of 30m. In habitats that the diver determined were homogenous (e.g., unbroken coral cover, algal plane), the diver recorded the terminal 10 m of the transect. In habitats that were more variable (e.g., reef edge habitats and other mixed bottom habitats), the diver recorded the full 30 m of the

transect. This strategy maximized limited bottom time when the habitat could be sufficiently sampled in a short transect, but captured greater variability in heterogeneous sampling locations.

After taping, approximately 40 - 80 non-overlapping images per transect were captured and saved as JPEG files on a computer using a Sony video capture card. Captured images represented a planar area of reef that varied around a true planar area of 0.31 m² (0.64 m x 0.48 m), and ultimately depended on the rugosity of the substratum. Microsoft Excel and Adobe Photoshop were used to superimpose ten randomly located dots on each captured image. The benthic cover under each of the points was then identified by experienced observers (TCRMP) to the lowest identifiable taxonomic level or abiotic group. For each transect, the percent cover of benthic categories was calculated by dividing the number of random points falling on the substrata type by the total number of points for the transect. The benthic categories that were assessed included: coral, dead coral with turf algae, macroalgae, sponges, gorgonians, and non-living substrata (sand, sediment, rubble, pavement). A total of 4,403 images were analyzed across the study.

Mean values for percent cover were calculated for each site and values were arcsine transformed prior to analysis. Data were tested for normality and homogeneity of variance. Data that met assumptions were analyzed with one-way ANOVA between benthic strata (algae, coral, pavement, non-living). Data that did not meet assumptions were tested with a non-parametric Wilcoxon test on rank sums. Significantly different means were analyzed post hoc with Tukey's HSD tests.

Coral Health

Scleractinian coral and hydrocoral colonies (all sizes) located directly under the transect lines were assessed in situ for signs of mortality and disease following a modified Atlantic and Gulf Rapid Reef Assessment protocol (AGRRA; Kramer et al. 2005). The line intercept method of coral health assessment provided a total coral sample size of 1,233 colonies for the study. Partial mortality of coral colonies was broken into the categories 'old partial mortality', skeleton eroded and covered with turf or macroalgae, and 'recent partial mortality', skeleton not eroded (fine corallite structure still intact) and bare or with a thin veneer of sheeting or filamentous algae. In the USVI, the transition between recent and old mortality categories usually occurs within three months following tissue death (Smith pers. obs.). In addition, old mortality becomes unrecognizable when the colony erodes into an amorphous form or a coral secretes new skeleton away from the dead surface. At this point it difficult to discern if a new coral has settled on a dead colony and sheeted, or if a surviving portion of a partially dead colony has resheeted. This transition takes place between 1 – 4 years after the initial mortality (Smith pers. obs.; also see <http://www.agrra.org/method/methodcor.html>). The surface area (%) of the colony that was dead was also estimated for each partial mortality category. Disease lesions and signs were categorized into recognized Caribbean scleractinian diseases and syndromes (e.g., white plague) following Bruckner 2007. In addition

to recognized coral diseases, a novel coral disease was encountered across numerous sites and is referred to in this study as Unknown Necrosis. The severity of disease on coral colonies was estimated as the area (percent of colony) of active disease lesion.

Bleaching¹ was assessed as abnormal paling of the colony, and, when present, the severity of the bleaching (paling or total whitening) and the area of the colony affected were assessed. This data was used to ordinate bleaching intensity into one of five categories: 0) unbleached, 1) any degree of paling less than completely white, or 1% - 10% bleached, 2) 10% – 50% bleached, and 4) 50% – 90% bleached, and 4) >90% bleached (after Gleason 1993). For each transect at each location, the prevalence of colonies with mortality, bleaching, and disease was calculated by dividing the number of affected colonies by the total number of colonies assessed.

Fish and Other Commercially Important Motile Resources

Characterization of the fish community and motile invertebrates (primarily lobster) was conducted along 25 x 4 m belt transects used in the benthic habitat surveys. Within this belt transect all fish species and motile invertebrates were identified to species level and total length estimated in 5 cm and 10 cm size categories (i.e. <5, 6-10, 11-20, 21-30, 31-40, >40). Data from fish and motile invertebrate density, diversity and size distribution were analyzed using non-parametric statistics (because of non-normal distribution patterns) to test for differences among habitat strata within the MCD.

Detection of the less abundant and often more commercially valuable species may be limited using standard belt transects. Therefore, a timed swim (i.e. roving diver technique) was conducted at a subset of randomly selected sites in which two divers actively searched for rare, cryptic and highly mobile species as well as invertebrate mega-fauna. Divers remained within a single habitat type but indicated on their data sheet if searches included habitat edges (abrupt change from one habitat type to another) or gradual transitions. The duration of the timed swim was constant throughout the study across all sites and habitats and was determined based on appropriate dive times and safety protocol. During the timed swim the observer recorded to species and enumerated all fish and invertebrate mega-fauna encountered on a logarithmic scale (i.e. 1, 2-10, 11-100, 101-1000). Data for total fish diversity was obtained from roving dives in the MCD at four designated habitat types (algae, coral, pavement, sand) sampled during the MCD survey and at two fixed sites sampled from 2005-2007 (see Table 6). These two sites included the Red Hind Bank (RHB), a coral reef site located in the eastern end of the MCD, and Collage Shoal East (CSE), another coral site located in the western part of the MCD (Figure 1, lower panel).

¹Bleaching is presented separately from other disease due to differing assessment of severity.

RESULTS AND RECOMMENDATIONS

Benthic Composition

Benthic sampling revealed a unique array of habitat types ranging from topographically simple algal and sand planes to highly complex coral reef banks. Examples of these habitats are presented as photographs in Fig. 3 (A-D), in captured stills from video transects (Appendix II), as videos (Appendix Video Captures III) and as video mosaics (Appendix III). The benthic composition of sampling locations is presented as percent cover from video assessments in Fig 4 - 7.

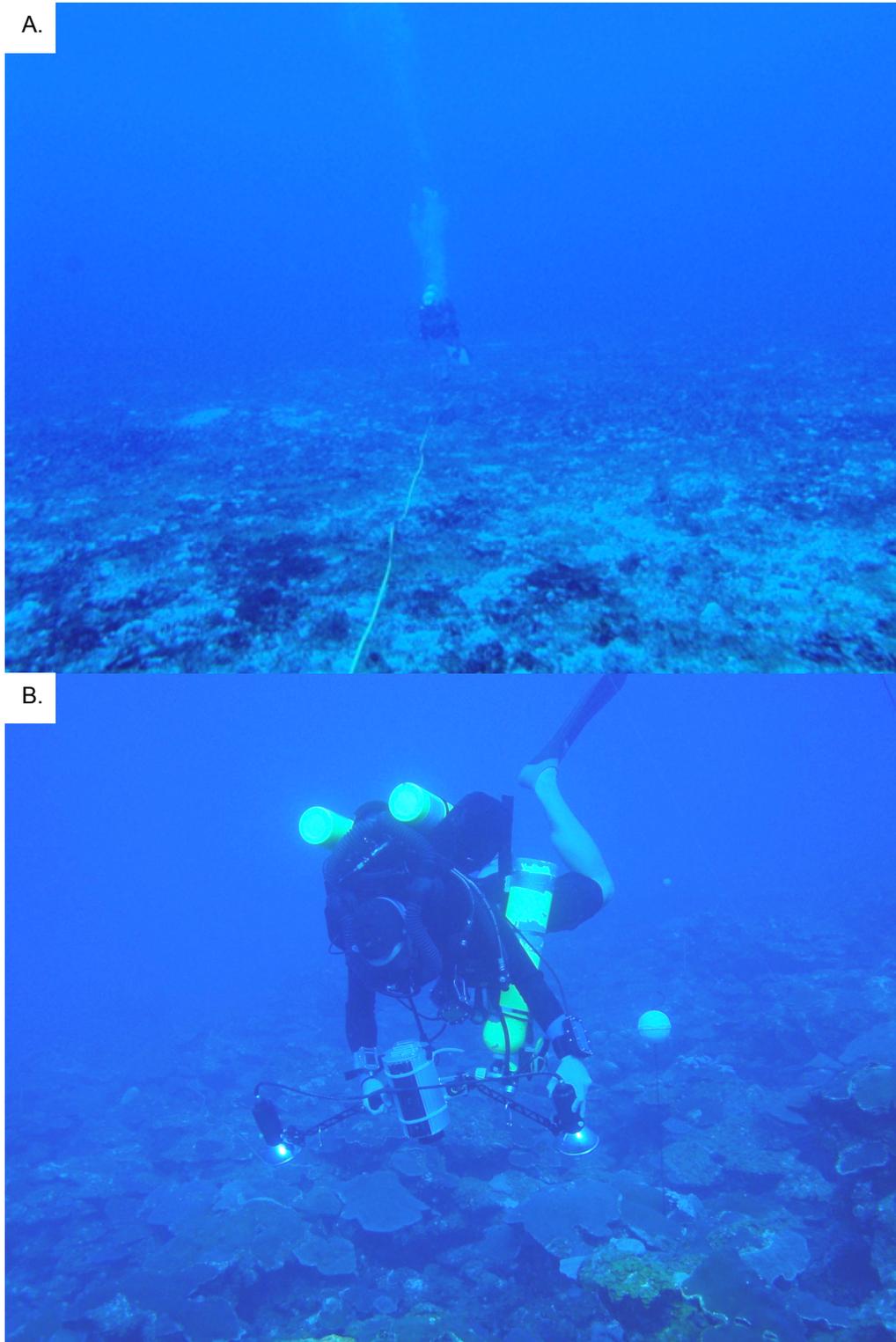


Figure 3. Representative photos of distinct habitat types sampled within the Marine Conservation District and Corresponding to A) Algae, B) Coral, (following page) C) Pavement, and D) Sand.



Figure 3. (continued)

Scleractinian Corals and Hydrocorals

Coral cover had a mean of 16.6% ± 1.9 SE and ranged from 0 to 50.1% across the MCD (Fig. 4A). For all locations that had coral cover greater than zero, coral cover had a mean of 20.7% ± 2.1 SE (N = 64). The dominant coral across the MCD were members of the *Montastraea annularis* species complex [*Montastraea annularis* (Ellis & Solander, 1786), *Montastraea faveolata* (Ellis and Solander, 1976), and *Montastraea franksi* Gregory, 1895)] and their coverage largely drove overall coral cover trends (Fig. 4B, Table 2). The cover of the *M. annularis* species complex across the MCD had a mean of 14.9% ± 1.8 SE and ranged from 0 to 48.1%.

In total, 20 scleractinian coral species or genera and one hydrocoral species were identified in video transects (Table 2). This estimate of scleractinian coral species richness was revised upwards to 25 from in situ coral health assessments that tended to include small coral species that were missed in video analysis and permitted a greater ability to separate rare genera into species (Table 3). At present, the number of known scleractinian coral and hydrocoral species for the MCD is 37 (Table 3), including one endangered or threatened species [*Acropora cervicornis* (Lamarck, 1816)].

Table 2. The ranked percent coverage and percent of total coral cover for scleractinian coral and hydrocoral species recorded in video surveys.

Species	Benthic Coverage	Percent of Coral Cover
<i>Montastraea annularis</i> species complex	14.81%	91.8%
<i>Agaricia</i> spp. (Lamarck, 1801)	0.47%	2.9%
<i>Porites astreoides</i> (Lamarck, 1816)	0.36%	2.3%
<i>Agaricia agaricites</i> (Linnaeus, 1758)	0.28%	1.7%
<i>Montastraea cavernosa</i> (Linnaeus, 1767)	0.21%	1.3%
<i>Siderastrea siderea</i> (Ellis and Solander, 1786)	0.08%	0.5%
<i>Porites porites</i> (Pallas, 1776)	0.05%	0.3%
<i>Stephanocoenia intersepta</i> (Lamarck, 1816)	0.04%	0.2%
<i>Diploria labyrinthiformis</i> (Linnaeus, 1758)	0.02%	0.1%
<i>Mycetophyllia</i> spp. (Milne Edwards and Haime, 1848)	0.02%	0.1%
<i>Madracis formosa</i> (Wells, 1973)	0.01%	0.1%
<i>Colpophyllia natans</i> (Houttyn, 1772)	0.01%	0.1%
<i>Siderastrea radians</i> (Pallas, 1766)	0.01%	0.1%
<i>Diploria strigosa</i> (Dana, 1848)	0.01%	0.1%
<i>Madracis decactis</i> (Lyman, 1859)	0.01%	0.1%
<i>Agaricia undata</i> (Ellis & Solander, 1786)	0.01%	0.04%
<i>Millepora alcicornis</i> (Linnaeus, 1758)	0.01%	0.03%
<i>Mycetophyllia aliciae</i> (Wells, 1973)	0.01%	0.03%
<i>Manicina areolata</i> (Linnaeus, 1758)	0.004%	0.02%
<i>Mycetophyllia ferox</i> (Wells, 1973)	0.003%	0.02%
<i>Eusmilia fastigiata</i> (Pallas, 1766)	0.002%	0.01%

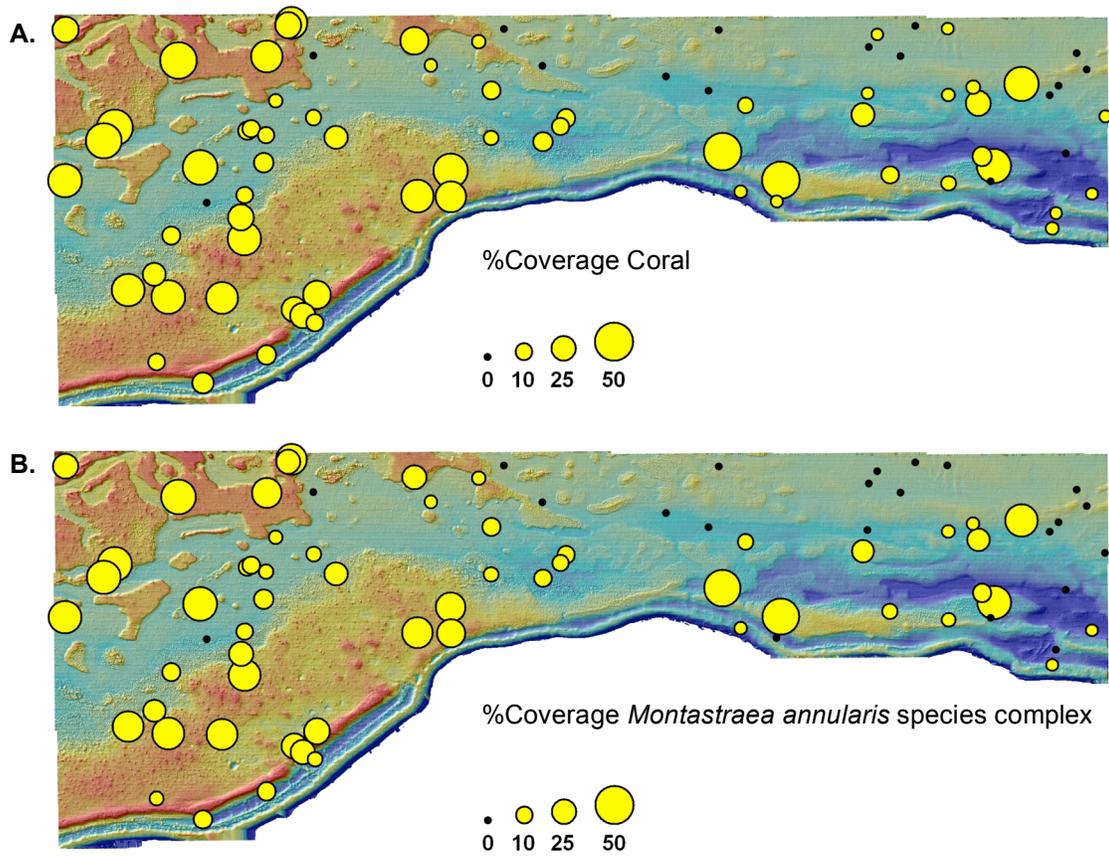


Figure 4. Percent coverage of A) all coral species and B) *Montastraea annularis* species complex in the Marine Conservation District.

Table 3. A list of known scleractinian coral and hydrocoral species found within the Marine Conservation District. “X” indicates that the coral was found in this study and/or in the Territorial Coral Reef Monitoring Program. “o” indicates that these species were likely encountered in this study, but could not be distinguished in the field.

Species	Found in Present Study	Found in TCRMP
<i>Acropora cervicornis</i>		X
<i>Agaricia agaricites</i>	X	X
<i>Agaricia fragilis</i> Dana, 1848		X
<i>Agaricia grahamae</i> Wells, 1973	o	X
<i>Agaricia humilis</i> Verrill, 1901		X
<i>Agaricia lamarcki</i> Milne Edwards and Haime, 1851	o	X
<i>Agaricia undata</i>	X	
<i>Colpophyllia natans</i>	X	X
<i>Dichocoenia stokesii</i> Milne Edwards and Haime, 1851		X
<i>Diploria labyrinthiformis</i>	X	X
<i>Diploria strigosa</i> (Dana, 1848)	X	X
<i>Eusmilia fastigiata</i>	X	X
<i>Isophyllia sinuosa</i> (Ellis & Solander, 1786)		X
<i>Helioseris cucullata</i> (Ellis & Solander, 1786)	X	
<i>Madracis decactis</i>	X	X
<i>Madracis formosa</i>	X	X
<i>Madracis mirabilis</i> (Duchassaing & Michelotti, 1860)		X
<i>Mancinia aerolata</i>	X	
<i>Meandrina meandrites</i> (Linnaeus, 1767)		X
<i>Montastraea annularis</i>	X	X
<i>Montastraea cavernosa</i>	X	X
<i>Montastraea faveolata</i>	X	X
<i>Montastraea franksi</i>	X	X
<i>Millepora alcicornis</i>	X	X
<i>Millepora complanata</i> Lamarck, 1816		X
<i>Mycetophyllia ferox</i>	X	
<i>Mycetophyllia aliciae</i>	X	
<i>Mycetophyllia danaana</i> Milne Edwards and Haime, 1848		X
<i>Mycetophyllia lamarckiana</i> Milne Edwards and Haime, 1851		X
<i>Oculina diffusa</i> Lamarck, 1816		X
<i>Porites astreoides</i>	X	X
<i>Porites divaricata</i> Lesueur, 1821	X	
<i>Porites porites</i>	X	X
<i>Scolymia</i> spp. Haime, 1852	X	
<i>Siderestrea siderea</i>	X	X
<i>Siderestrea radians</i>	X	
<i>Stephanocoenia intercepta</i>	X	

Gorgonians and Sponges

Gorgonians and sponges were minor constituents of the benthos of the MCD, and separately averaged less than 1% of benthic coverage (Fig. 5). The coverage of soft corals had a mean of $0.1\% \pm 0.02$ SE and ranged from 0 to 1.1%. The coverage of sponges had a mean of $0.7\% \pm 0.1$ SE and ranged from 0 to 3.9%.

Although species-level discrimination of soft corals and sponges was not attempted from video transects, observations suggested that diversity within these groups was high and included rare and commercially important shallow water types, such as antipatharians (black coral). Soft coral and sponge diversity can be more accurately assessed with surveys that directly target these groups, use methods to determine area-based density, and attempt in situ species level categorization.

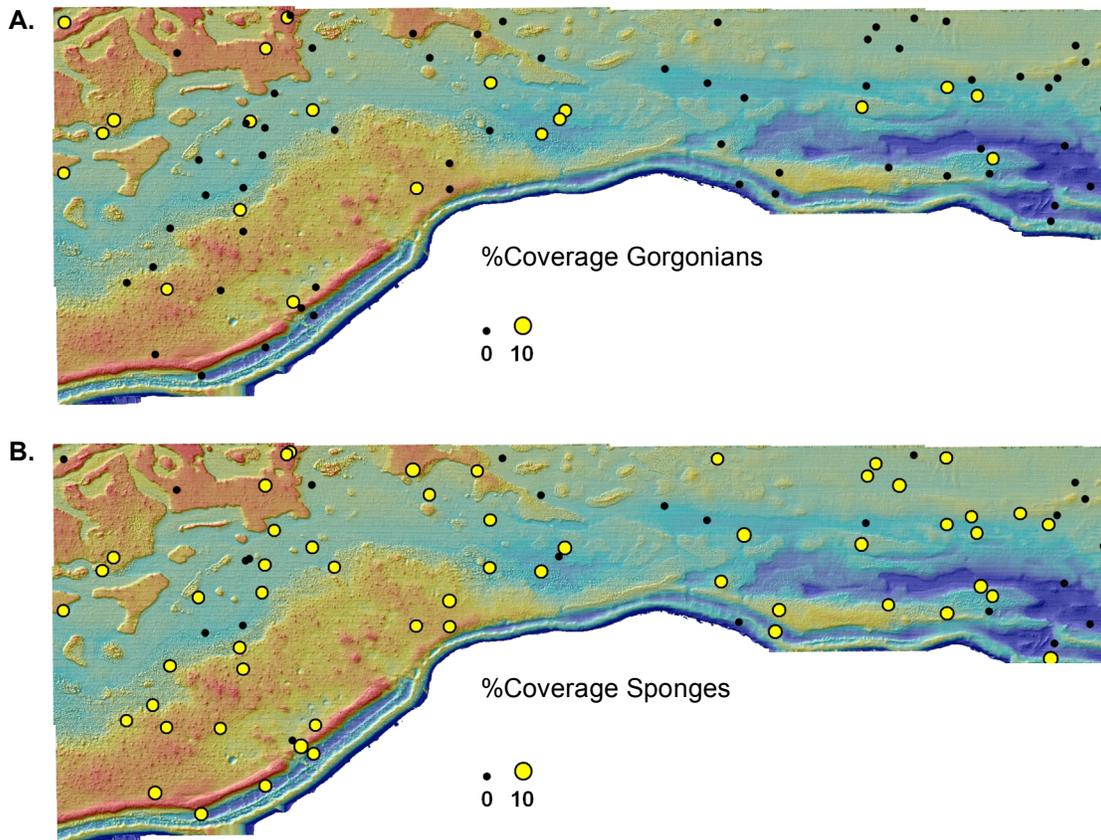


Figure 5. Percent coverage of A) gorgonians and B) sponges in the Marine Conservation District.

Algae

Algae covered the majority of the substrata for most sites in the MCD. Algal cover had a mean of $61.6\% \pm 1.7$ SE and ranged from 0 to 94.8% across the MCD. The majority of algal cover consisted of macroalgae and large filamentous cyanobacteria, and had a mean coverage of 38.6 ± 2.3 SE and ranged from 0 to 93.3% (Fig. 6A). Over half of macroalgae and filamentous cyanobacteria were composed of the filamentous cyanobacteria *Schizothrix* spp. (mean = $11.7\% \pm 1.9$ SE, range: 0 to 70.5%; Fig. 6B) and the phaeophyte *Lobophora variegata* (mean = $9.7\% \pm 1.0$ SE, range: 0 to 30.1%; Fig. 6C). A minor component of macroalgae was composed of *Dictyota* spp., (mean = $0.5\% \pm 0.1$ SE; Fig. 6D). Dead coral covered with turf algae comprised the second most important category of algae and had a mean of $18.9\% \pm 1.8$ SE and ranged from 0 to 64.7% (Fig. 6E). Crustose coralline algae formed a relatively minor component of the benthic algal cover and had a mean of $4.1\% \pm 0.5$ SE and ranged from 0 to 16.7% (Fig. 6F).

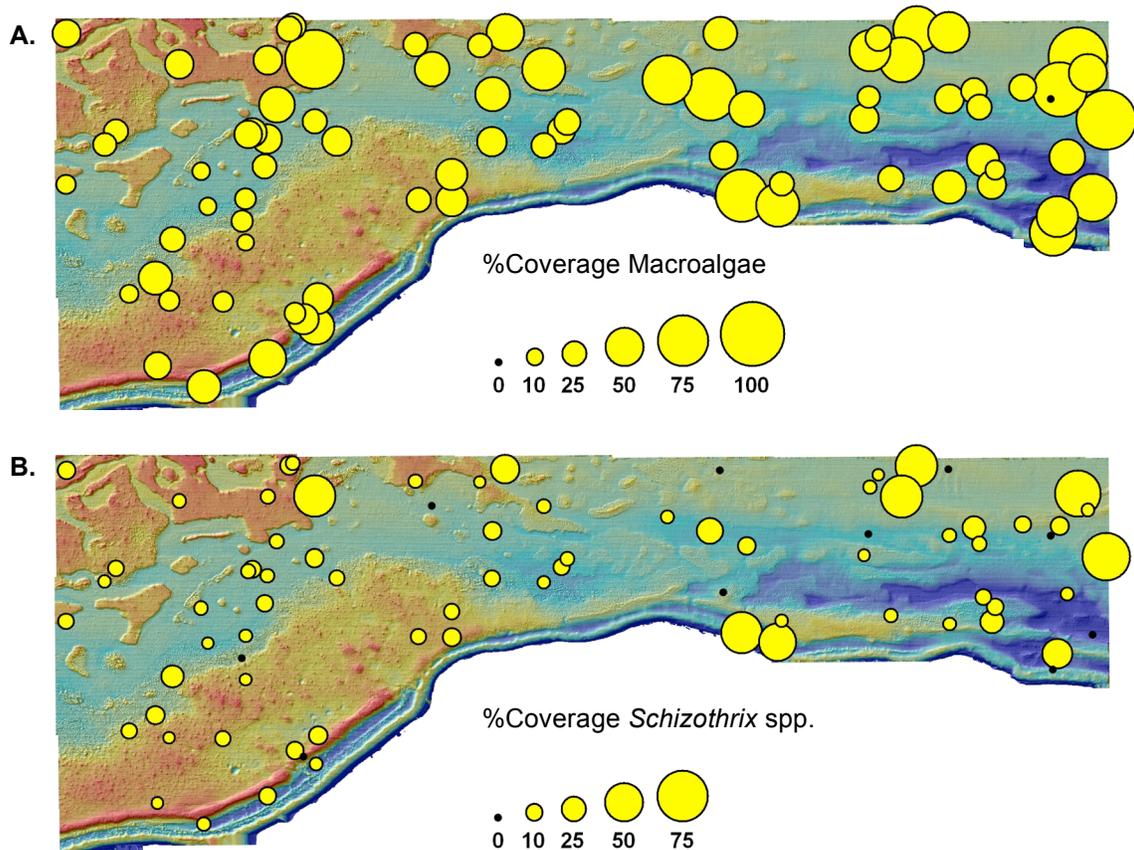


Figure 6. Percent coverage of A) macroalgae, B) *Schizothrix* spp., (following pages) C) *Lobophora variegata*, D) *Dictyota* spp., E) dead coral covered with turf algae, and F) crustose coralline algae in the Marine Conservation District.

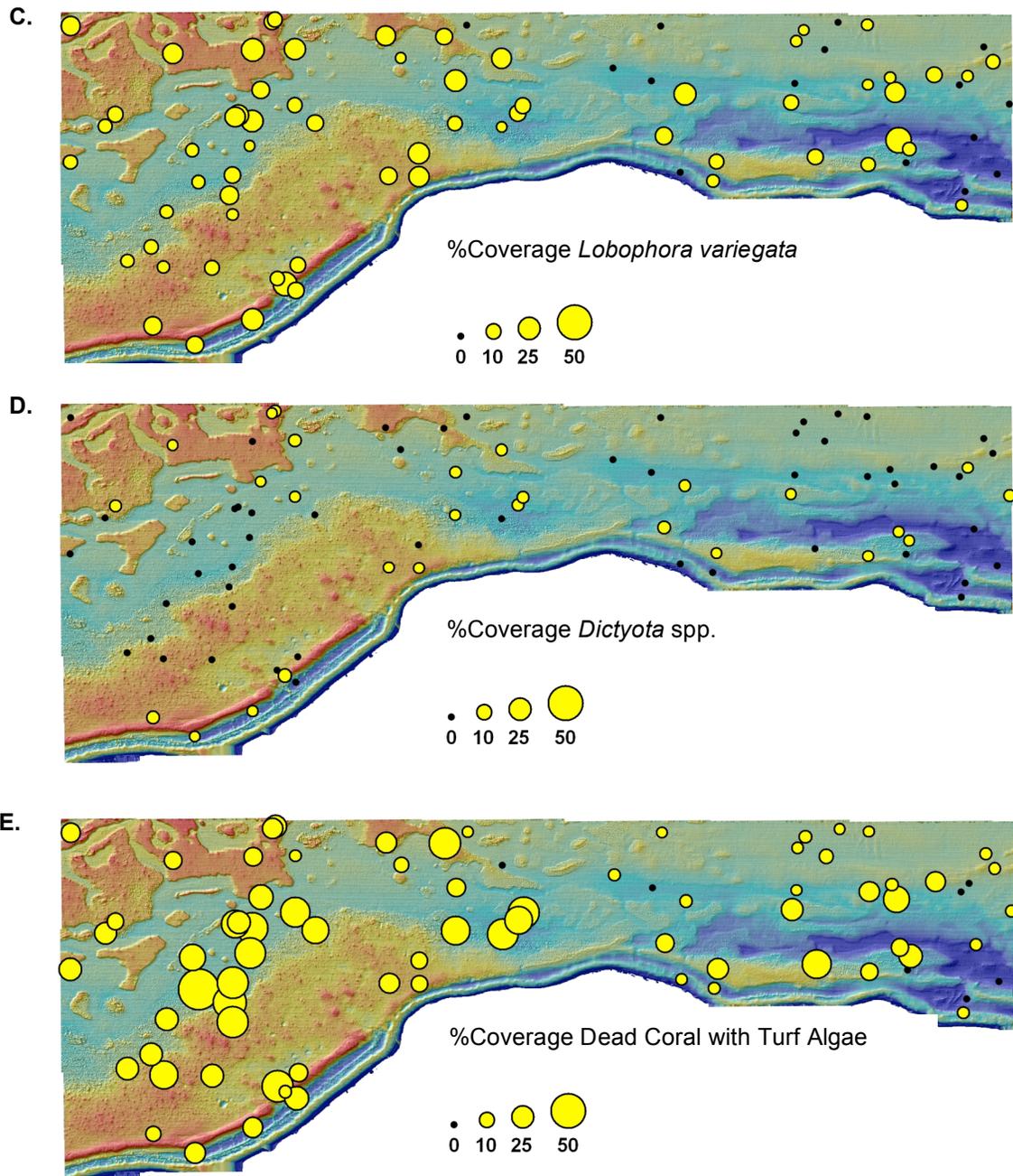


Figure 6. (continued)

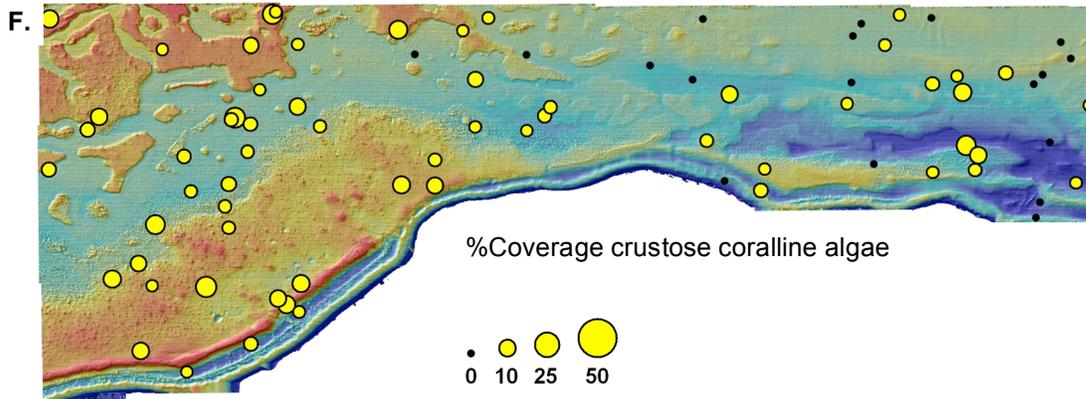


Figure 6. (continued)

Non-living Benthic Cover

Non-living benthic substrata formed the third highest benthic coverage category after algae and corals, and had a mean of $20.8\% \pm 2.3$ SE and ranged from 0 to 97.5% (Fig. 7A). The majority of non-living substrata was composed of sand/sediment, and had a mean coverage of $15.7\% \pm 2.2$ SE and ranged from 0 to 97.2% (Fig. 7B). Pavement formed the second highest portion of non-living substrata, and pavement had a mean coverage of $3.1\% \pm 0.6$ SE and ranged from 0 to 27.5% (Fig. 7C). Coral rubble formed the third highest portion of non-living substrata, and had a mean coverage of $2.1\% \pm 0.7$ SE and ranged from 0 to 38.5% (Fig 7D).

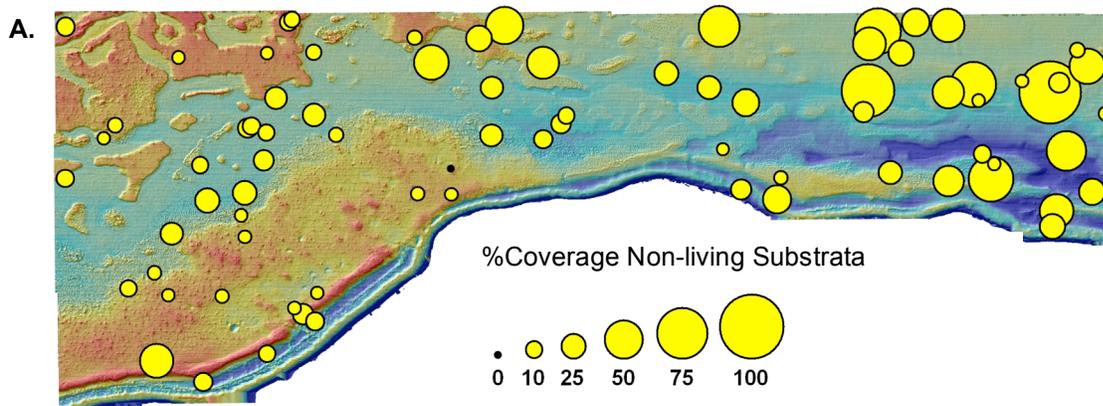


Figure 7. Percent coverage of A) all non-living substrata, (following page) B) sand/sediment, C) pavement, and D) rubble in the Marine Conservation District.

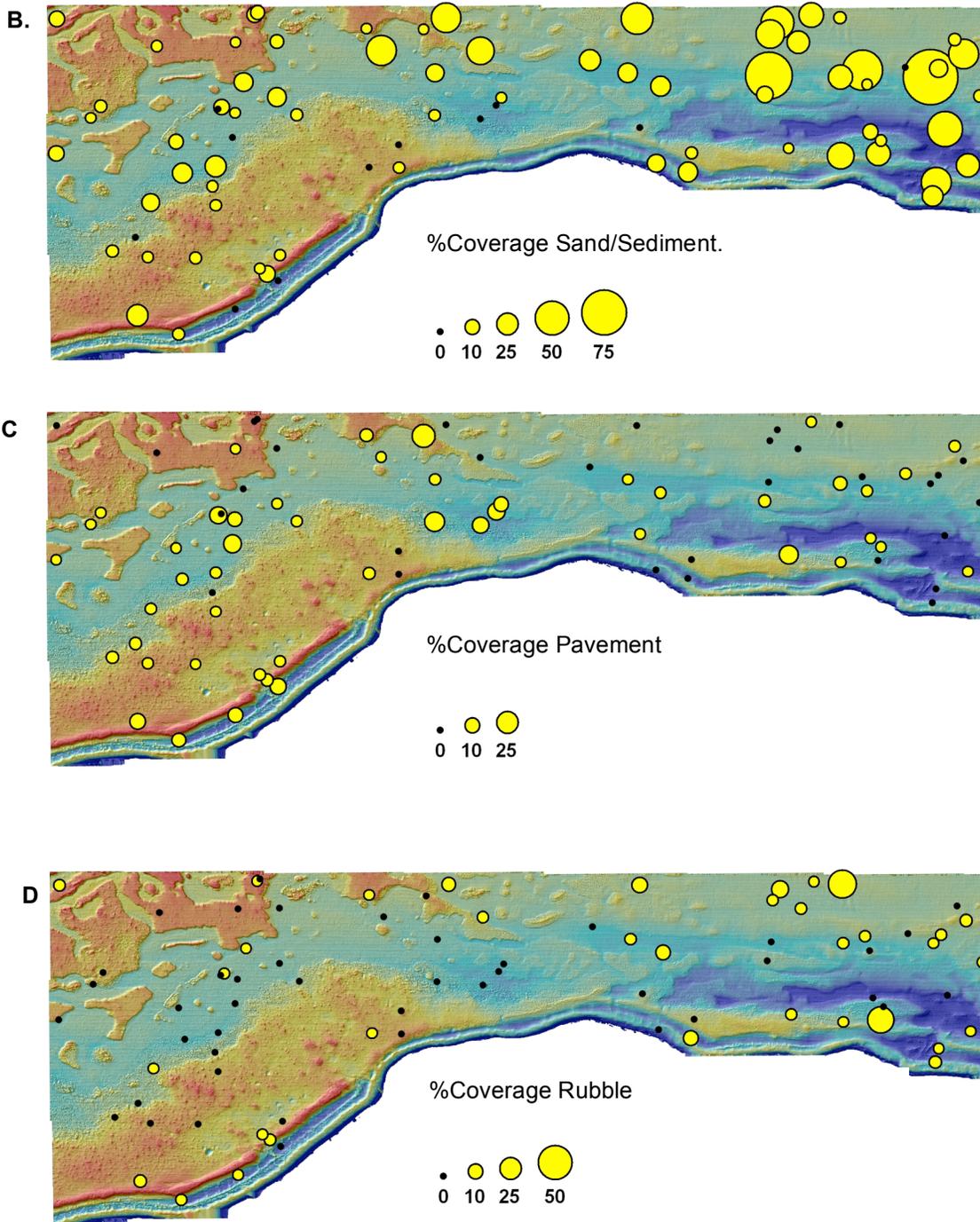


Figure 7. (continued)

Benthic Composition by Sampling Strata

The four predicted benthic strata used to stratify sampling effort had different characteristics for many of the examined variables. Depth was significantly different between strata ($F = 8.5, p < 0.0001, N = 80$), with the coral stratum significantly shallower than all other strata, which were not different from each other (Fig. 8).

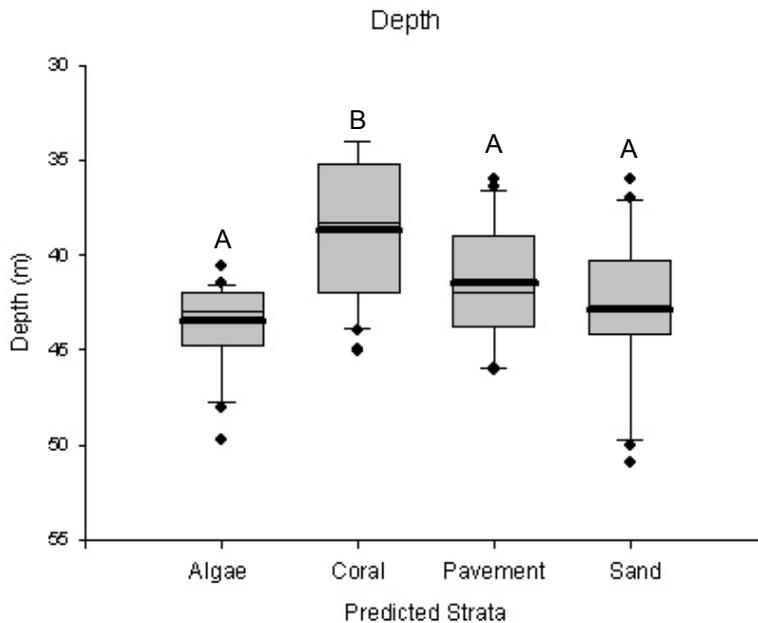


Figure 8. Depths of sampling locations among four predicted strata of the Marine Conservation District. Figure components are mean (thick black line), median (thin black line), 25th and 75th percentiles (bottom and top of box, respectively), 10th and 90th percentiles (bottom and top whiskers, respectively) and values outside 10th and 90th percentile (dots). Homogeneous subsets of means are indicated with letters.

Benthic Epifauna

Coral coverage was significantly different between strata ($\chi^2 = 55.1, p < 0.0001$). The coral stratum had significantly higher coral coverage than all other strata (nearly double), with the pavement and sand strata not significantly different from each other, but higher than the algae stratum, which was nearly zero (Fig. 9). Gorgonian coverage was significantly different between strata ($\chi^2 = 17.8, p < 0.0005$), and was highest in the coral and pavement strata, and over an order of magnitude less in the sand and algae strata (Fig. 9). However, pavement was not significantly different than the higher coverage coral stratum or the lower coverage sand and algae strata. Sponge coverage was significantly different between strata ($\chi^2 = 16.0, p$

< 0.0012), but gradually declined from the coral stratum, to the pavement and sand strata, and finally to the algae strata (Fig. 9). Only the coral and algae strata were significantly different from each other.

Algae

Algae coverage was significantly different between strata ($\chi^2 = 16.7$, $p < 0.0008$). The coral stratum had significantly less algal cover than all other strata, which were not significantly different from each other (Fig. 10). The cover of macroalgae was significantly different between strata ($\chi^2 = 22.3$, $p < 0.0001$). The coral stratum had the least cover of macroalgae and was significantly different from the algae stratum, which had the highest macroalgae cover (Fig. 10). Coverage of macroalgae in the sand and pavement strata was intermediate between the coral and algae strata. Sub-categories of macroalgae cover were only significantly different for *L. variegata* ($F = 9.4$, $p < 0.0001$). The coverage of *L. variegata* was highest in the coral and pavement strata, which were not significantly different from each other (Fig 10). Coverage of *L. variegata* was least in the algae stratum, and intermediate in the sand stratum. Other sub-categories of macroalgae cover were not significantly different between strata, and included *Schizothrix* spp. ($\chi^2 = 2.7$, $p = 0.438$) and *Dictyota* spp. ($F = 2.0$, $p = 0.122$). The remaining two categories of algal cover did show differences between strata. Dead coral covered with turf algae was significantly different between strata ($\chi^2 = 38.4$, $p < 0.0001$). The coverage of dead coral covered with turf algae was significantly lower in the algae stratum than all other strata, and was an order of magnitude less in the algae stratum than in the other strata (Fig. 11). The coverage of dead coral covered with turf algae was not significantly different between the coral, pavement, and sand strata. Crustose coralline algae coverage was significantly different between strata ($\chi^2 = 32.0$, $p < 0.0001$). The coral and the pavement strata had the highest coverage of crustose coralline algae and were not significantly different from each other (Fig. 11). The coral and pavement strata had coverage of crustose coralline algae that were significantly greater than the algae stratum, but not the sand stratum, which was intermediate between the coral and pavement strata, and the algae stratum.

Non-living Benthic Composition

Non-living benthic coverage was significantly different between strata ($\chi^2 = 35.5$, $p < 0.0001$). Non-living benthic coverage was significantly greater in the algae stratum than all other strata, which were not significantly different from each other (Fig. 12). Sand/sediment, the dominant component of non-living strata (see Benthic Composition Section), was significantly different between strata ($\chi^2 = 37.3$, $p < 0.0001$). Sand/sediment coverage was significantly greater in the algae stratum than all other strata, which were not significantly different from each other (Fig. 12). Pavement coverage was significantly different between strata ($\chi^2 = 22.8$, $p < 0.0001$). Pavement coverage in the pavement stratum was significantly greater than the algae and sand strata, which were not significantly different from each other (Fig. 12). Pavement coverage in the coral stratum was intermediate between the other strata. Rubble coverage was not significantly different between strata ($F = 1.9$, $p = 0.132$; Fig. 12).

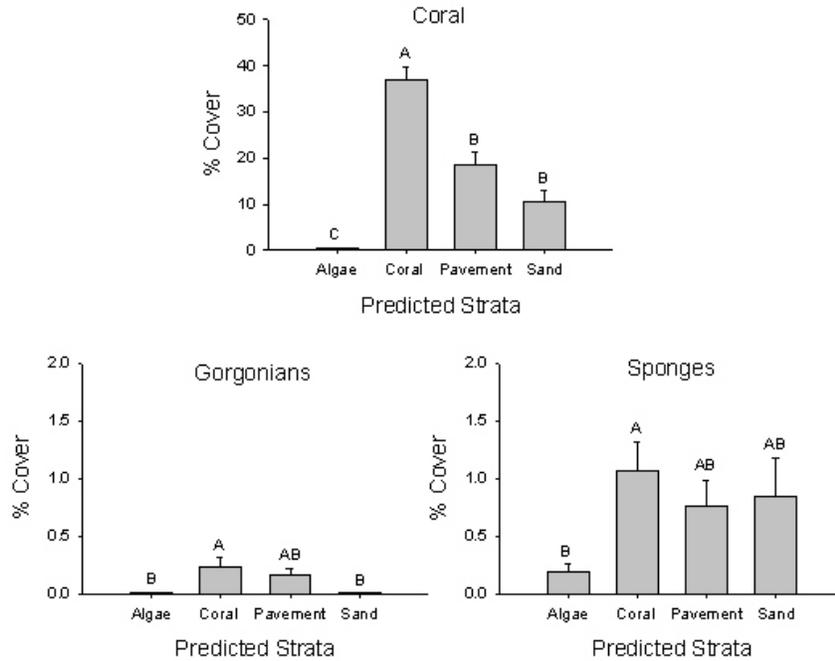


Figure 9. The percent coverage (\pm SE) of the benthic fauna categories coral, gorgonians, and sponges among the predicted benthic strata (N = 80). Homogeneous subsets of means are indicated with letters (Tukey HSD Post Hoc comparison).

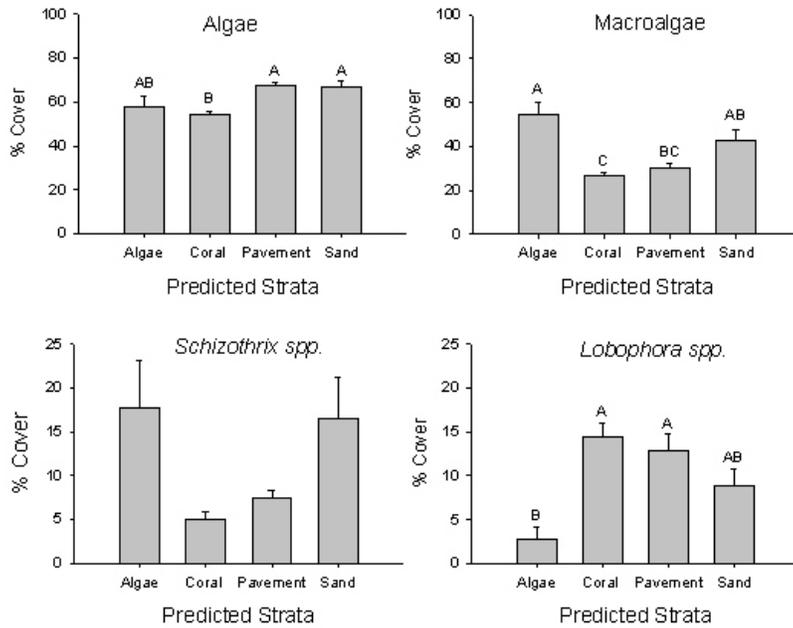


Figure 10. The percent coverage (\pm SE) of the benthic algae categories total algae, total macroalgae, the filamentous cyanobacteria *Schizothrix* spp., and the phaephyte *Lobophora variegata* among the predicted benthic strata (N = 80). Homogeneous subsets of means are indicated with letters (Tukey HSD Post Hoc comparison).

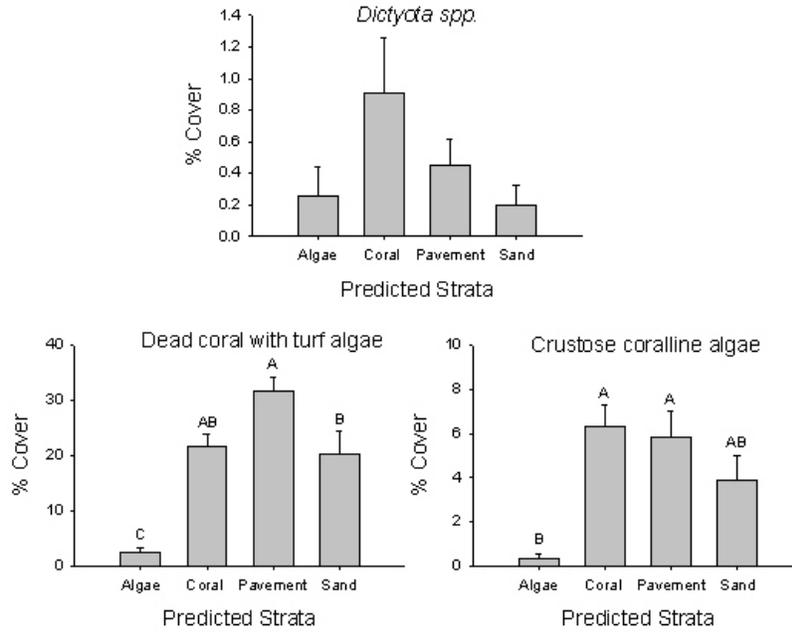


Figure 11. The percent coverage (\pm SE) of the benthic algae categories the phaephyte *Dictyota* spp., dead coral covered with turf algae, and crustose coralline algae among the predicted benthic strata (N = 80). Homogeneous subsets of means are indicated with letters (Tukey HSD Post Hoc comparison).

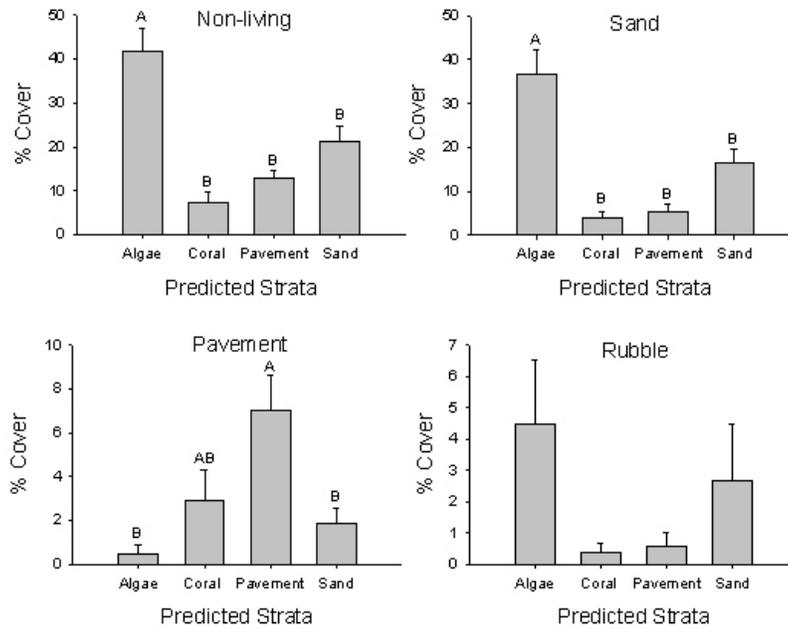


Figure 12. The percent coverage (\pm SE) of the abiotic benthic categories total non-living substrata, sand/sediment, pavement, and rubble among the predicted benthic strata (N = 80). Homogeneous subsets of means are indicated with letters (Tukey HSD Post Hoc comparison).

Predicted Versus Sampled Benthic Strata

The randomized stratified sampling design allows an assessment of the accuracy of multibeam and side-scan sonar benthic habitat characterization in the MCD. There was uncertainty in what proportionate coverage of various groups (e.g., macroalgae, pavement, coral) represented the four predicted benthic categories: algae, coral, pavement, sand. This makes it somewhat difficult to compare the results of this study with the predicted benthic habitat. However, it was assumed that large deviations from predicted habitat structure would indicate obvious areas requiring improvement in the benthic classification algorithms.

The percent composition of algae, coral, pavement, and sand strata are shown in Fig. 13. The predicted strata are broken into their constituent parts and represented as proportions of the total benthic coverage. In addition, the proportion of substrata that fell into hard substrata, soft substrata, or variable substrata is shown. Variable substrata are living benthic components (e.g., macroalgae, sponges) that are overlaying hard substrata (e.g., coral, dead coral with turf algae) or soft substrata (i.e., sand/sediment). Thus, their percent coverage could be representative of hard or soft substrata, but this was not determined from video records. Both in situ diver records and benthic cover reported in surveys deviate from predicted habitat types in many surveys. This problem was particularly evident in the pavement and sand strata. Each of the four predicted strata is assessed below:

Algae

The algae stratum was consistent and well predicted. Sites classified as algae were low relief algal communities atop unconsolidated sediment (Fig. 3A). Although 55% of the algae stratum had variable benthic composition, in situ observation strongly suggested that this largely overlaid unconsolidated sediment. Thus, about 92% of algae stratum was unconsolidated sediment, of which 55% was composed of macroalgae and filamentous cyanobacteria, and 37% was composed of sand/sediment. In these areas, expanses of unconsolidated sediment were broken by occasional coral colonies and dead coral rubble piles, and much less commonly by patch coral reefs and pavement. Site-attached fishes were often associated with these small habitat patches.

Coral

The coral stratum was consistent and well predicted. Sites classified as coral were medium to high relief coral reef. Although 28% of the coral stratum had variable benthic composition, in situ observation strongly suggested that this largely overlaid consolidated hardbottom. Thus, about 95% of the coral strata were composed of consolidated hardbottom, of which $38.7\% \pm 2.9$ SE was composed of scleractinian corals.

This was unusually high coral cover compared to modern shallow Caribbean coral reefs (~10% coral cover; Gardener et al. 2003), and was particularly striking following coral mortality in shallow and deep coral reefs of the U.S. Caribbean following bleaching and disease in 2005 and 2006 (Smith et al. in prep.).

In situ observations also suggested that the coral stratum could be further divided into sub-types associated with edges, shallow coral bank tops (30-40 m), and deeper coral plains (35 – 45 m). In all cases the dominant matrix was formed of low or plating morphologies of the *M. annularis* species complex (Fig 3B, Appendix IV). Living upper surfaces of corals often rested upon pillars of dead coral, creating a complex interstitial network of overhangs, channels, and tunnels. This under-explored network may form a large habitat area that is likely to be an important component of essential fish habitat in the MCD.

Pavement

The pavement stratum was not well predicted. Sites classified as pavement were dominantly composed of coral reef that fell into the coral reef sub-categories of shallow coral reef bank tops, and deeper coral plains (see 'coral' section above). The pavement stratum had 31% variable benthic composition, and in situ observation suggested that this was largely composed of consolidated hardbottom. However, in the deeper coral plains there were patches of sand interspersed between coral pillars. Thus, for the shallower (30 – 37 m) coral reef bank tops found in the southwestern corner of the MCD (Fig. 2B), about 95% of the benthos was composed of consolidated hardbottom. In the deeper coral plains that ran as a channel through the center of the western MCD (Fig. 2B), there was higher variability in the variable substratum category, and it was estimated that consolidated hardbottom comprised about 80% of the benthos.

As with the coral stratum, the matrix of the pavement stratum was dominantly composed of low or plating morphologies of the *M. annularis* species complex (Fig 3C). Living upper surfaces of corals often rested upon pillars of dead coral, creating a complex interstitial network of overhangs, channels, and tunnels; however, this network was more open than in the coral strata, and had fewer tunnels. Corals of this stratum may undergo periodic mortality events that generate a more open network. Sampling in the pavement strata revealed a widespread and severe coral mortality event, particularly evident and general to the deeper coral plain (see 'Coral Health' section below).

Sand

The sand stratum was predicted to varying accuracy that depended on location in the MCD. Sites classified as sand were either sand/algae habitats or were coral reef. Accurately classified sand stratum sites were largely confined to the southeast corner of the MCD (Fig. 2B). In situ observations suggest that sand habitat classified outside the shelf edge coral reef banks at the south drop-off were largely soft

bottom habitats to a depth of at least 65 m. The remainder of habitat classified as sand was formed in a complex area of coral pillars and hillocks interspersed with sand patches (Fig. 3D). This area comprised a large section of the western MCD, behind the primary and secondary shelf edge coral reef banks (Fig. 2B). The sand stratum had 44% variable benthic composition, and in situ observation suggested that in the majority of habitat in the western MCD this was composed of an equal mix of consolidated hardbottom and unconsolidated sediments. Thus, for the coral reef hillocks found in the southwestern corner of the MCD, approximately 60% of the benthos was composed of consolidated hardbottom, and the remaining 40% of unconsolidated sediments (i.e. sand/sediment)..

The majority of the sand stratum was comprised of coral hillocks that differed from the geomorphology of coral reefs in the coral and pavement strata. Instead of more horizontally uniform coral reef areas, the hillocks rose 2 to 7 m above the surrounding coral reef plain and were 5 to 15 in diameter (Fig. 3D). The hillocks and the surrounding reef was dominantly composed of low or plating morphologies of the *M. annularis* species complex. Living upper surfaces of corals often rested upon pillars of dead coral, creating a complex interstitial network of overhangs, channels, and tunnels; however, as in the pavement stratum, this network was more open than in the coral stratum, and had fewer tunnels.

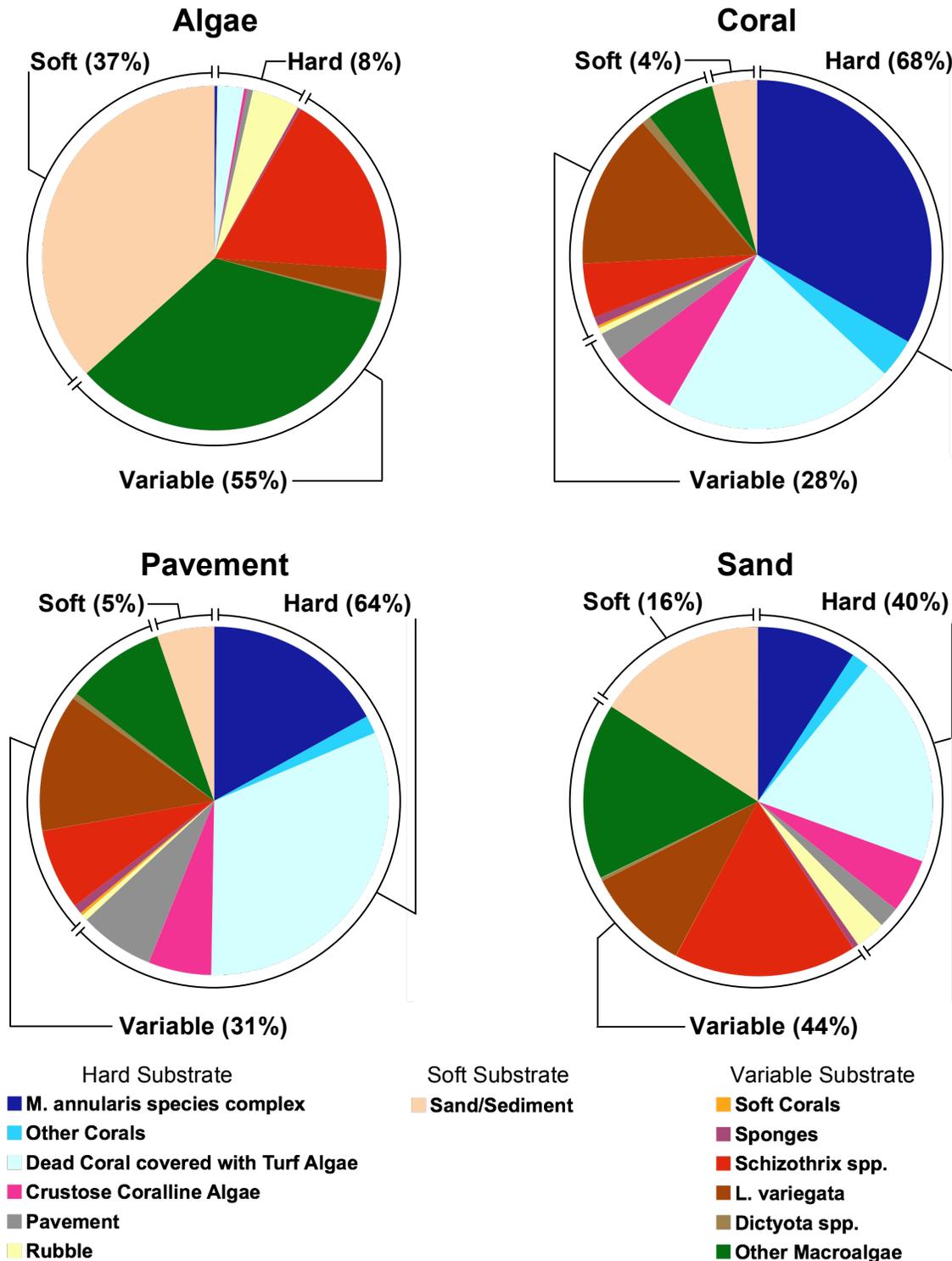


Figure 13. Proportionate benthic composition determined from in situ surveys across four general benthic strata determined from side-scan sonar processing: algae, coral, pavement, sand. In situ data within each benthic strata were apportioned into hard substrate, soft substrate, and variable substrate categories with proportion of total (%) in brackets.

Coral Health

Disease and Bleaching

Assessments of coral condition showed wide variations of coral health across sites and striking recent disease and degradation in a spatially coherent area of the MCD. Coral health was assessed on 1,233 colonies across 64 locations that contained coral within transects. Overall, coral disease (with the exception of bleaching) on coral colonies had a mean prevalence of $18.5\% \pm 0.03$ SE and a mean severity (percent of colony affected) of $26.9\% \pm 3.9$ SE (Fig. 14A). The vast majority of coral disease signs were caused by an unknown coral syndrome, named “unknown necrosis” (Fig. 14B). Unknown necrosis had a large mean prevalence of $17.4\% \pm 3.6$ SE and mean severity of $32.8\% \pm 4.7$ SE. This syndrome is described in further detail below. White Plague was the second most common disease, and had a mean prevalence of $0.8\% \pm 0.004$ SE and a mean severity of $11.6\% \pm 3.2$ SE (Fig. 14C). Bleaching had a mean prevalence of $12.4\% \pm 1.5$ SE and a mean severity (ordinated levels) of $1.9\% \pm 0.1$ SE (Fig. 14D). This severity most closely corresponds to level 2 bleaching: 10% – 50% coral colony bleached.

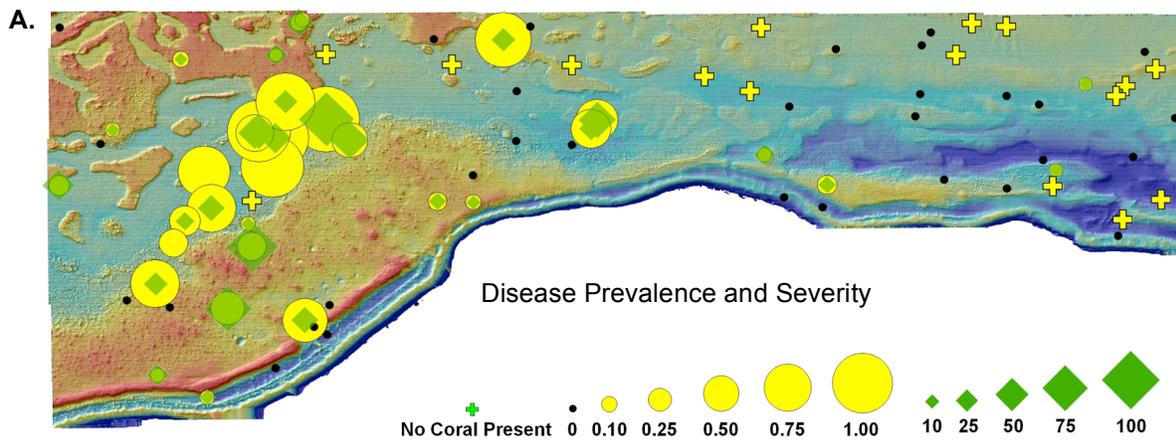


Figure 14. Prevalence and severity of A) total disease, (following page) B) unknown necrosis, C) white plague, and D) bleaching on hard corals of the Marine Conservation District.

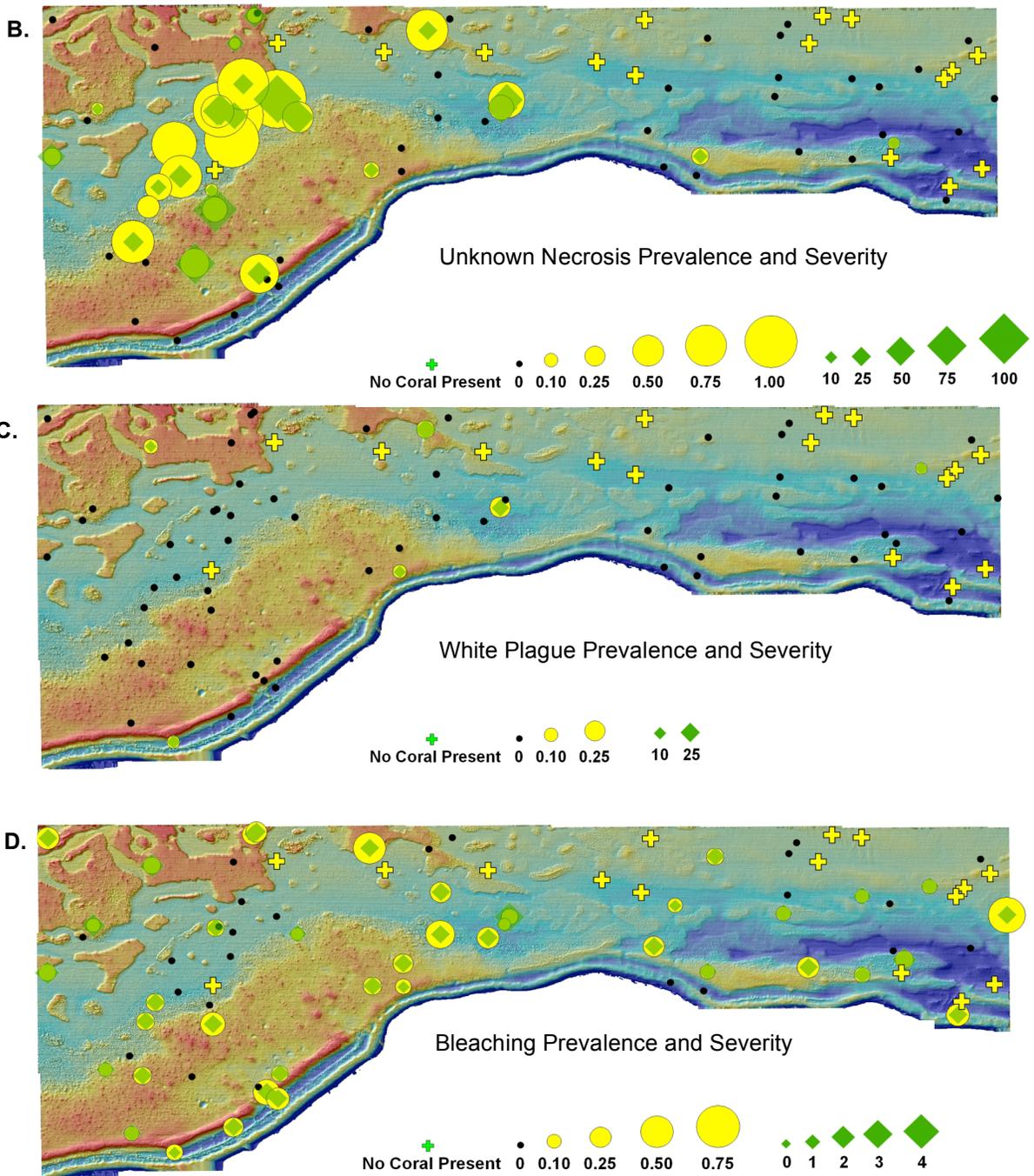


Figure 14. (continued)

Partial Mortality

Partial mortality of coral colonies was a common feature of coral reefs within the MCD. Recent partial mortality had mean prevalence of $30.6\% \pm 4.0$ SE and mean severity of $22.7\% \pm 2.9$ SE (Fig. 15A). The prevalence of recent partial mortality was strongly correlated with the prevalence of unknown necrosis ($R = 0.852$, $F = 171.1$, $p < 0.0001$). Old partial mortality had a mean prevalence of $65.0\% \pm 3.4$ SE and a mean severity of $26.2\% \pm 1.8$ SE (Fig. 15B). The prevalence of old partial mortality was a general characteristic of most coral harboring sites and was not significantly correlated with the prevalence of unknown necrosis ($R = 0.223$, $F = 3.4$, $p = 0.070$).

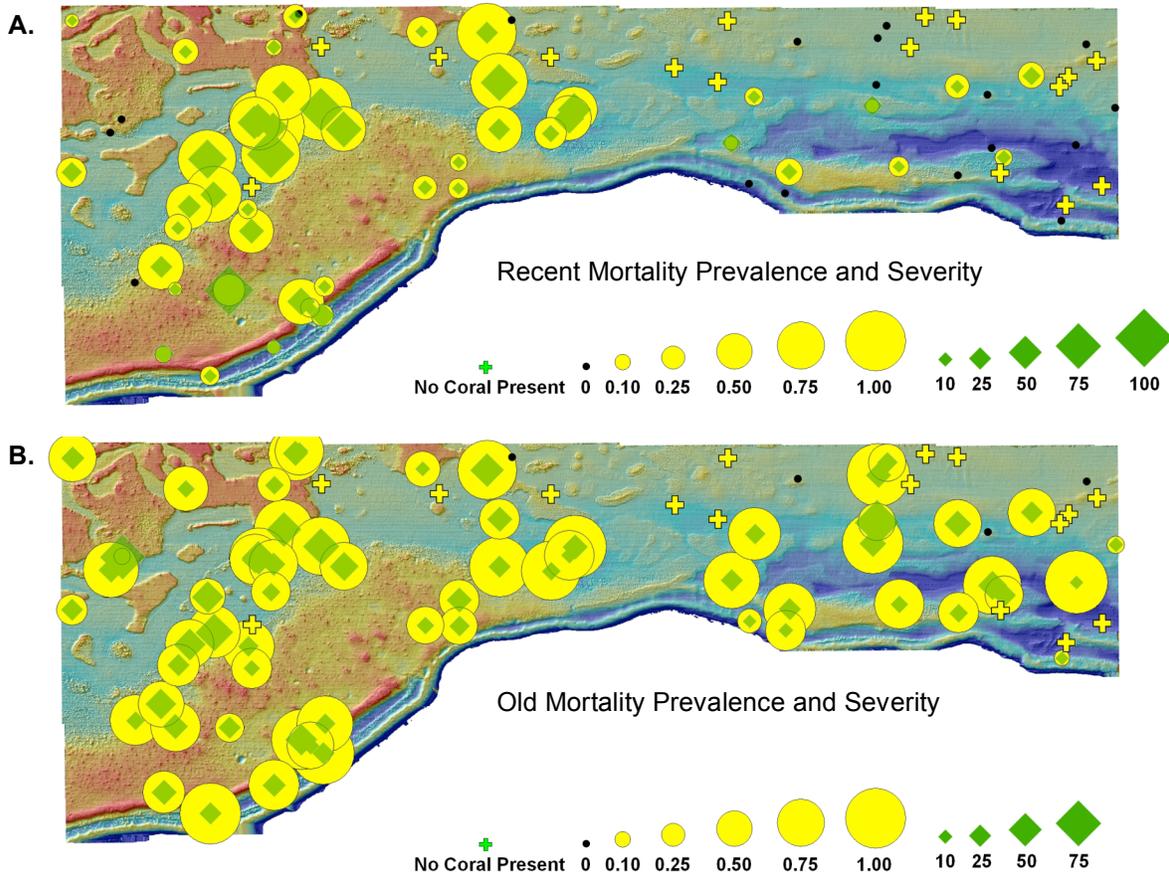


Figure 15. Prevalence and Severity of A) recent mortality, and B) old mortality on hard corals in the Marine Conservation District.

Unknown Necrosis

Coral disease sampling revealed an unusual, extensive, and dramatic syndrome affecting large numbers of coral. This unknown necrosis was manifested as white areas of tissue loss in a variable pattern. It was distinguished from white plague disease signs in that the areas of tissue loss were not present as a progressing linear lesion over colonies (Fig. 16). Instead, white areas of tissue necrosis appeared sporadically or generally over colonies. In members of the *M. annularis* species complex, presumably early stages of the disease primarily affected intercostal regions between the polyps (Fig. 17). These areas formed more general regions of necrosis in later stages of the disease (Fig. 18). Although the coral disease monitoring protocol was not longitudinal (individual colonies were not reassessed for pattern of disease progression), the progression of the disease was deduced from colonies displaying both sporadic intercostal necrosis and regional necrosis (Fig. 18). In coral species with meandering corallite structure (e.g., agariciids), signs of disease appeared as both tattered-appearing areas of necrosis and regional necrosis. Numerous colonies in affected areas were noticed that had recently suffered 100% mortality and 26 recently killed colonies were found in transects (2% prevalence). The disease was noted affecting three coral genera, however, prevalence was highest for the *M. annularis* species complex (Table 4).

Table 4. Species of scleractinian coral affected by unknown necrosis, and the prevalence, severity, and sample size among affected species in the Marine Conservation District.

Species	Prevalence	Severity	N
<i>Agaricia agaricites</i>	0.020	ND	49
<i>Agaricia</i> spp.	0.023	65%	86
<i>Montastraea annularis</i> species complex	0.203	40%	907
<i>Siderastrea radians</i>	0.167	50%	6
<i>Siderastrea siderea</i>	0.029	80%	35

Repeated observations and surveys of coral health in coral reefs affected by unknown necrosis suggested that the disease peaked during earlier stages of sampling (October 2007) and had largely abated within three months. There was a high prevalence and severity of unknown necrosis in October 2007 and repeated observation after the start of 2008 revealed few cases of unknown necrosis. Furthermore, two locations sampled in October 2007 had a mean prevalence of unknown necrosis of 35.8%, but had no cases of unknown necrosis in January 2008. The abatement of unknown necrosis is also illustrated in the video mosaic image of location S166 recorded in January 2007 (Appendix IV – Sand 166). No visible signs of unknown necrosis are visible on this image, although unknown necrosis was present in October 2007.

The coral disease signs consistent with unknown necrosis were present over a large swath of the MCD. The occurrence of unknown necrosis on coral reefs was largely confined to the western-central area of the MCD in a basin behind the thin primary and wide secondary outer reef banks forming the southwestern corner of the MCD (see Fig. 14B). Observations and surveys of coral reefs adjacent to affected areas, but on topographic highs, such as edges of reef banks or reef bank tops, showed that unknown necrosis was largely confined to the basin. The proximity of unaffected faunas and the spatial coherence of affected areas suggested that unknown necrosis was not being driven by a pathogen, but was a response to a common abiotic driver.

The effects and extent of unknown necrosis in the MCD suggest a strong structuring force of essential fish habitat (EFH) in this mesophotic reef system. The mean prevalence ($42.4\% \pm 6.3$ SE, $N = 27$) and severity ($32.8\% \pm 4.6$ SE) of unknown necrosis at affected sites suggested that the effects of the disease were intense in these areas. Extrapolation of the occurrence of unknown necrosis across the basin suggested that an area of approximately 9 km^2 was affected by the mortality event, over a fifth of the benthic habitat of the MCD shallower than 50m. While the driver of this disease and coral mortality were unknown, it is possible that cryptic mortality events are a recurrent and structuring force in these habitats. Importantly, the mass disease occurred outside of the two Territorial Coral Reef Monitoring Program sites maintained in the MCD. This mass disease and mortality event was captured opportunistically by coincidence with this research project to assess habitats and resources of the MCD. The occurrence, effects, and extent of large-scale mortality events in mesophotic reef systems may be an important consideration for EFH. The novel mass disease event presented in this research report should be an impetus for expanded and more intensive monitoring or coral health in mesophotic reefs.

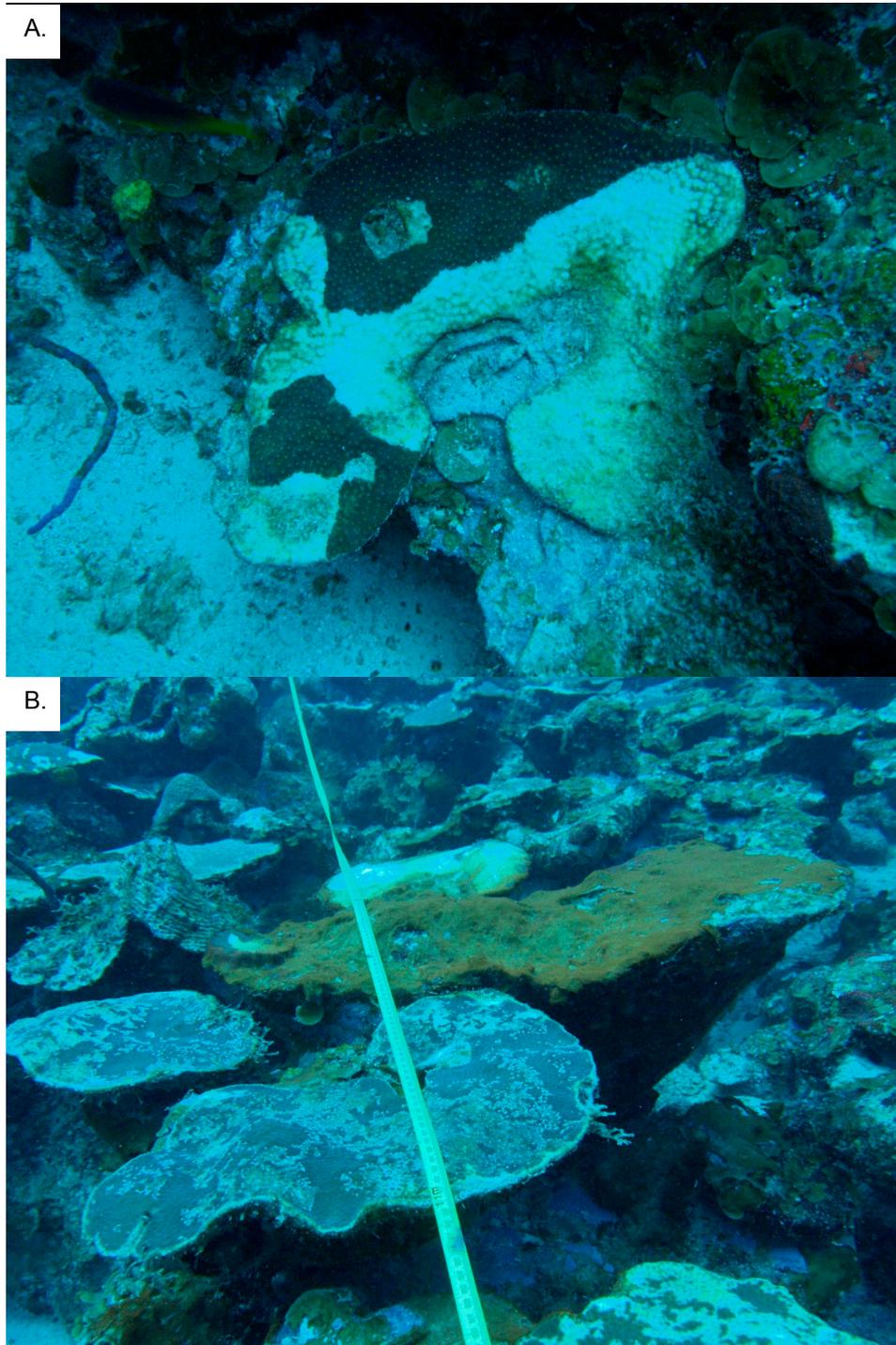


Figure 16. Photographic examples of coral disease in the Marine Conservation District: A) a coral colony affected by white plague (site: coral 58), B) corals affected by unknown necrosis. (site: Pavement 119, October 9, 2007).

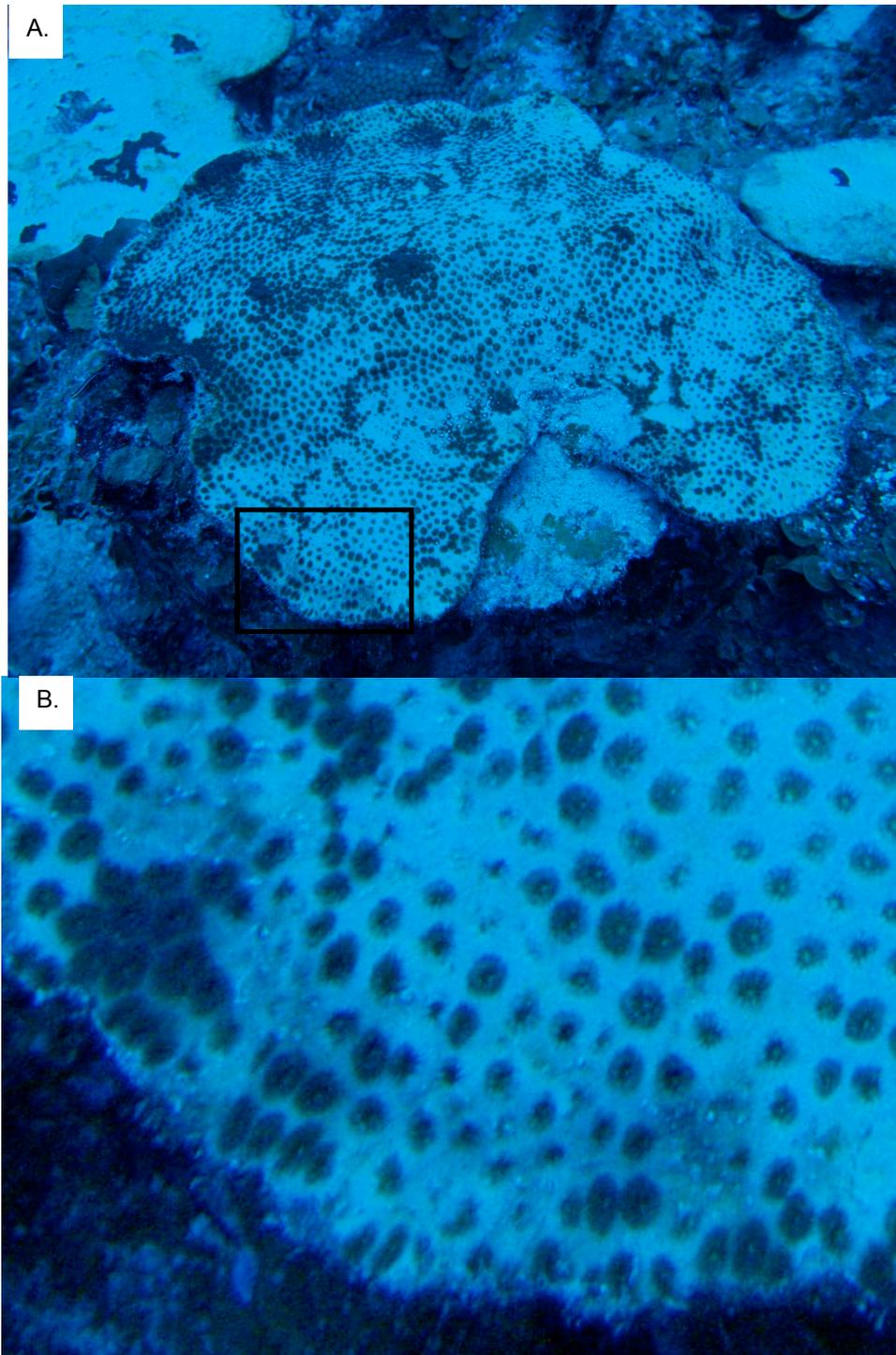


Figure 17. Photographic examples of coral disease in the Marine Conservation District: A) intercostal necrosis B) close-up of intercostal necrosis (site: pavement 111, October 29, 2007).

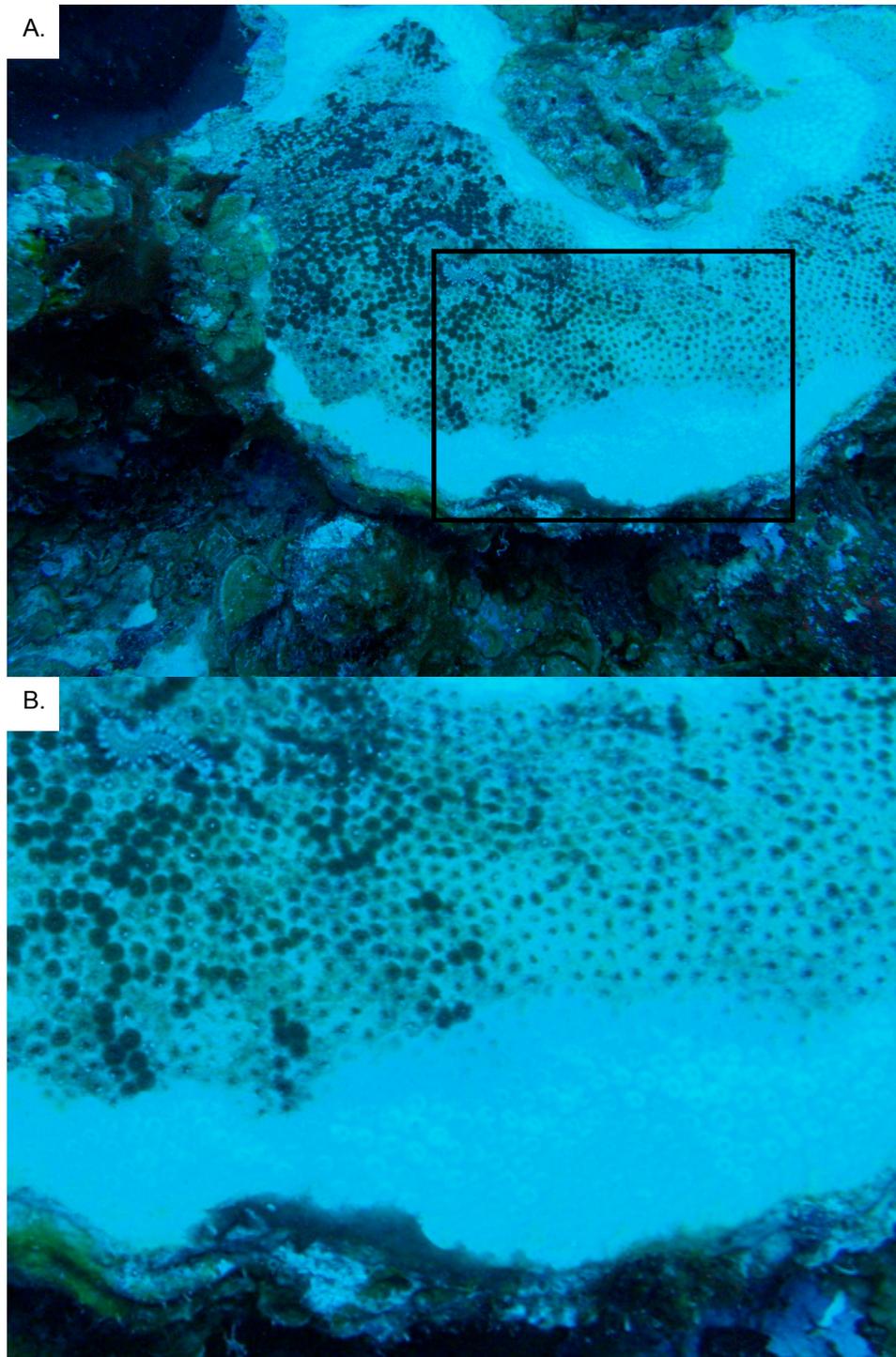


Figure 18. Photographic examples of coral disease in the Marine Conservation District: A) intercostals necrosis grading to general necrosis B) close-up of intercostal necrosis grading to general necrosis (site: pavement 111, October 9, 2007)

Assemblage Structure of Fish and Motile Mega-Invertebrates

The reef fish community structure at each sampling location in the Marine Conservation District is presented as density per 100 m² from belt transects. Motile mega-invertebrates were not observed in this study MCD.

Diversity, Abundance and Biomass of Reef Fishes

A total of 112 species were recorded throughout the MCD (see Table 6). Average fish species richness along belt transects was highest in coral habitats (mean = 14.4 100 m⁻², range = 6 to 25 spp), followed by sand habitats (mean = 12.6 spp 100 m⁻², range = 5 to 25 spp), pavement (mean = 11.5, spp 100 m⁻², range = 6 to 18 spp) and algal plains (mean = 6.0, spp 100 m⁻², range = 1 to 17 spp) (Fig. 19). Based on belt transects, Scarids showed the highest level of species richness followed by Serranids, Lutjanids and Haemulids (Fig. 20 A - D). Species richness was significantly lower in algal habitats ($p < 0.05$) than in the three other habitat types (i.e. coral, pavement and sand). The distribution of the most abundant species among the sampling strata is presented in Fig. 21.

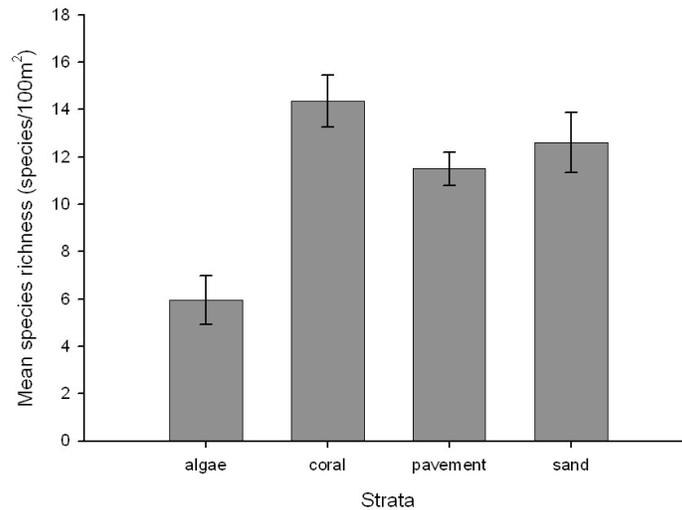


Figure 19. Mean species richness (\pm SE) for each habitat strata in the Marine Conservation District.

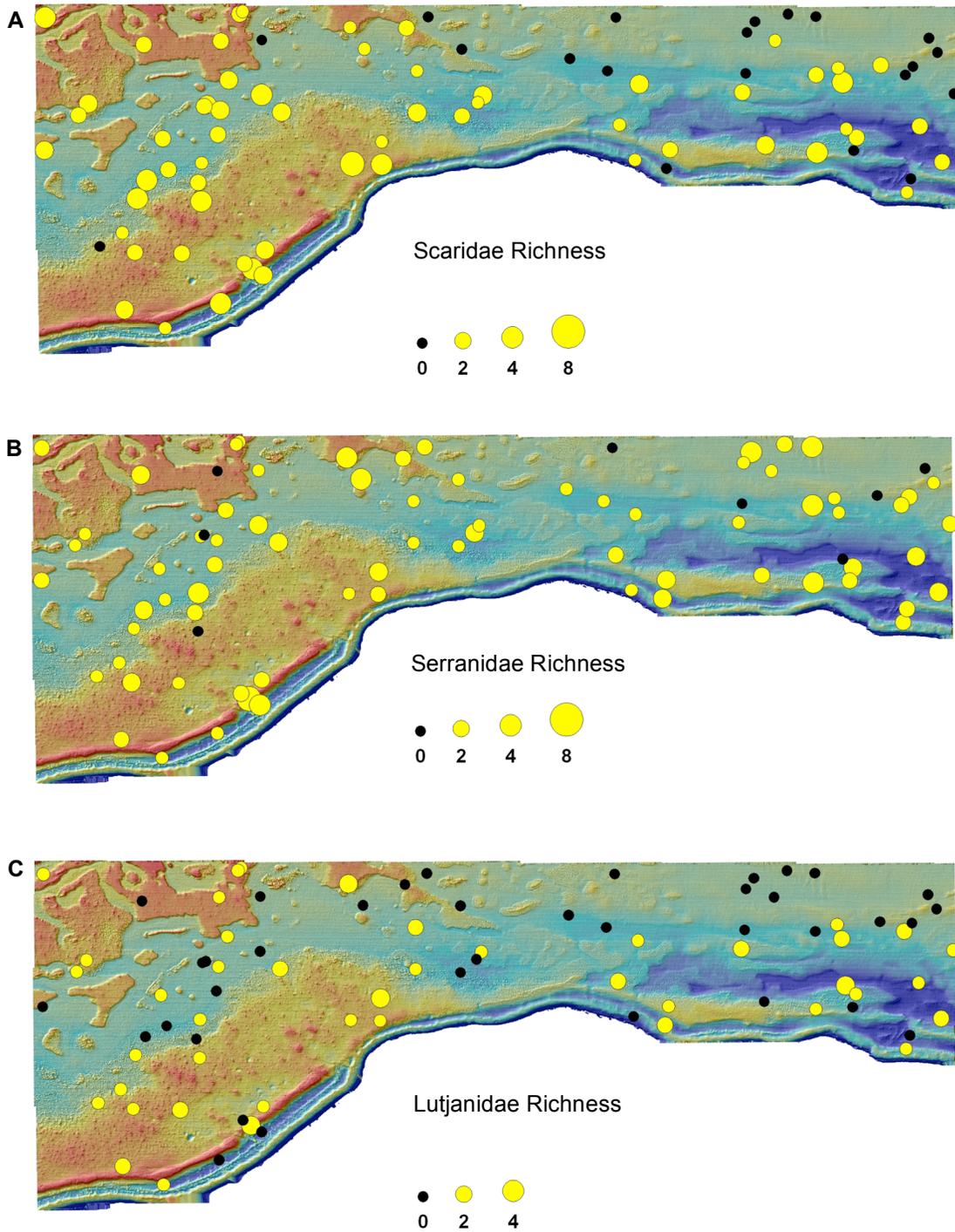


Figure 20. Family richness of A) Scaridae, B) Serranidae, C) Lutjanidae, and D) Haemulidae at each sampling location in the Marine Conservation District.

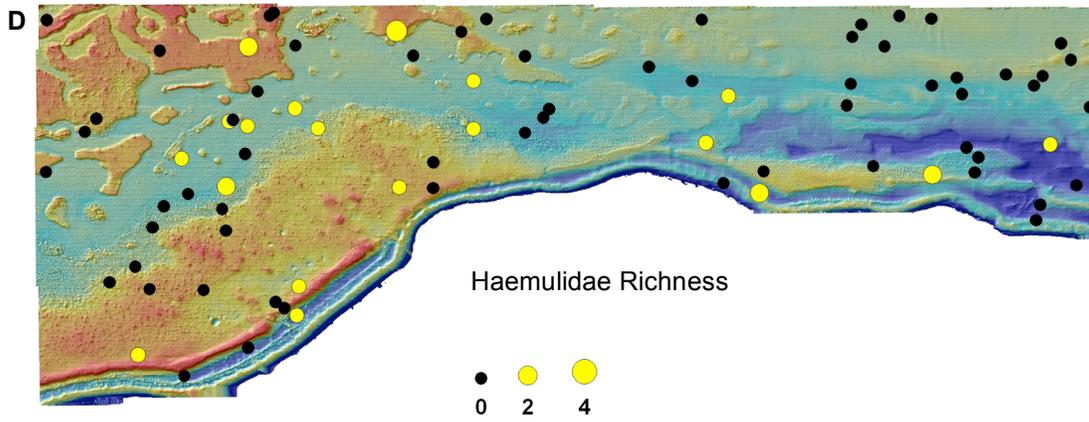


Figure 20. (continued)

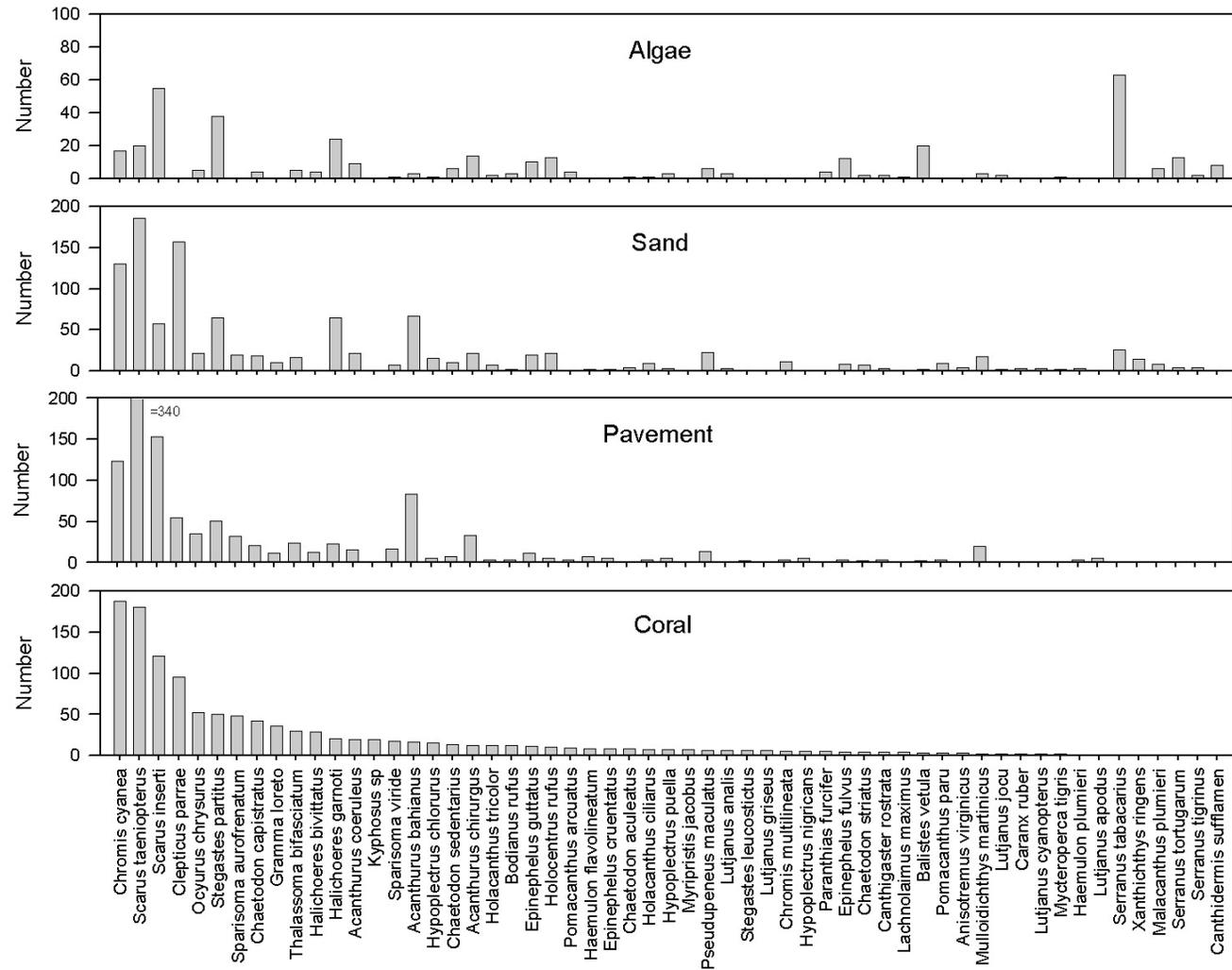


Figure 21. Total number of the most abundant species observed in belt transects in each habitat strata of the Marine Conservation District. Species were included if cumulative number in all habitats was ≥ 5 and then ordered from the most to least abundant from the coral habitat.

Abundance of reef fishes varied throughout the MCD depending upon taxa. Most commercially important species were commonly found in coral reefs and hard bottom habitats (Fig. 22), with higher concentrations near habitat edges and greater abundance near the shelf edge, but also along the northern and western MCD boundaries. Species with the highest abundances that were also of high economic and ecological importance were used to plot abundance distribution patterns. Acanthurids (*A. bahianus*, *A. coeruleus*, *A. chirurgus*) were fairly evenly distributed throughout the MCD with slightly lower abundances near the northern boundary of the closed area (Fig. 23A). The Scarids had similar distribution patterns as the Acanthurids (Fig. 23B) but were 4 times more abundant. Haemulids were uncommon within the MCD and were present at less than 25% of the 80 sampling locations (Fig. 23C). Serranid abundance was relatively evenly distributed across the MCD (Fig. 23D) and the group was largely composed of smaller species within the *Hypoplectrus* and *Serranus* genera (see Table 6). Lutjanids had a similar distribution as Haemulids, but were observed at just over 50% of the sampling sites (Fig. 23E). Balistids were more common on the eastern end of the MCD and were present at nearly 25% of the sites (Fig. 23F). The red hind (*Epinephelus guttatus*) was present at nearly 60% of all sites and had relatively high biomass levels (Fig. 24).

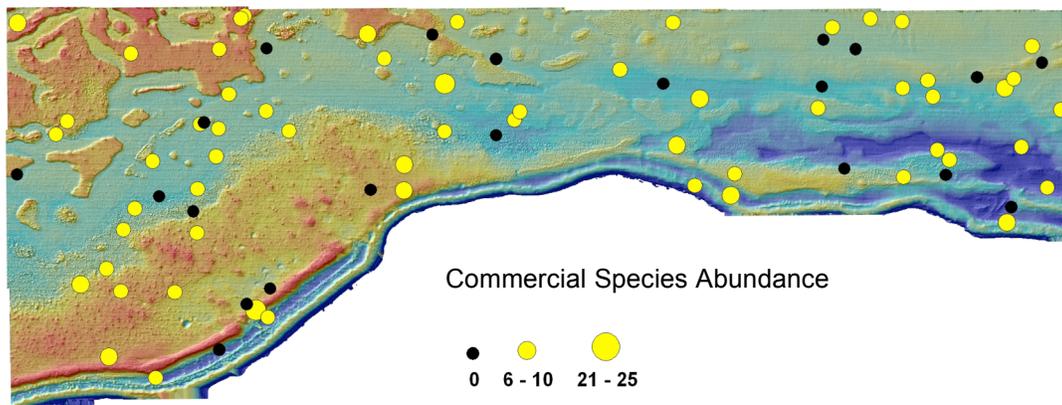


Figure 22. Abundance (ind. /100m²) of commercially important species (*E. guttatus*, *M. venenosus*, *E. striatus*, *L. analis*, *O. chrysurus* and *B. vetula*) at each sampling location in the Marine Conservation District.

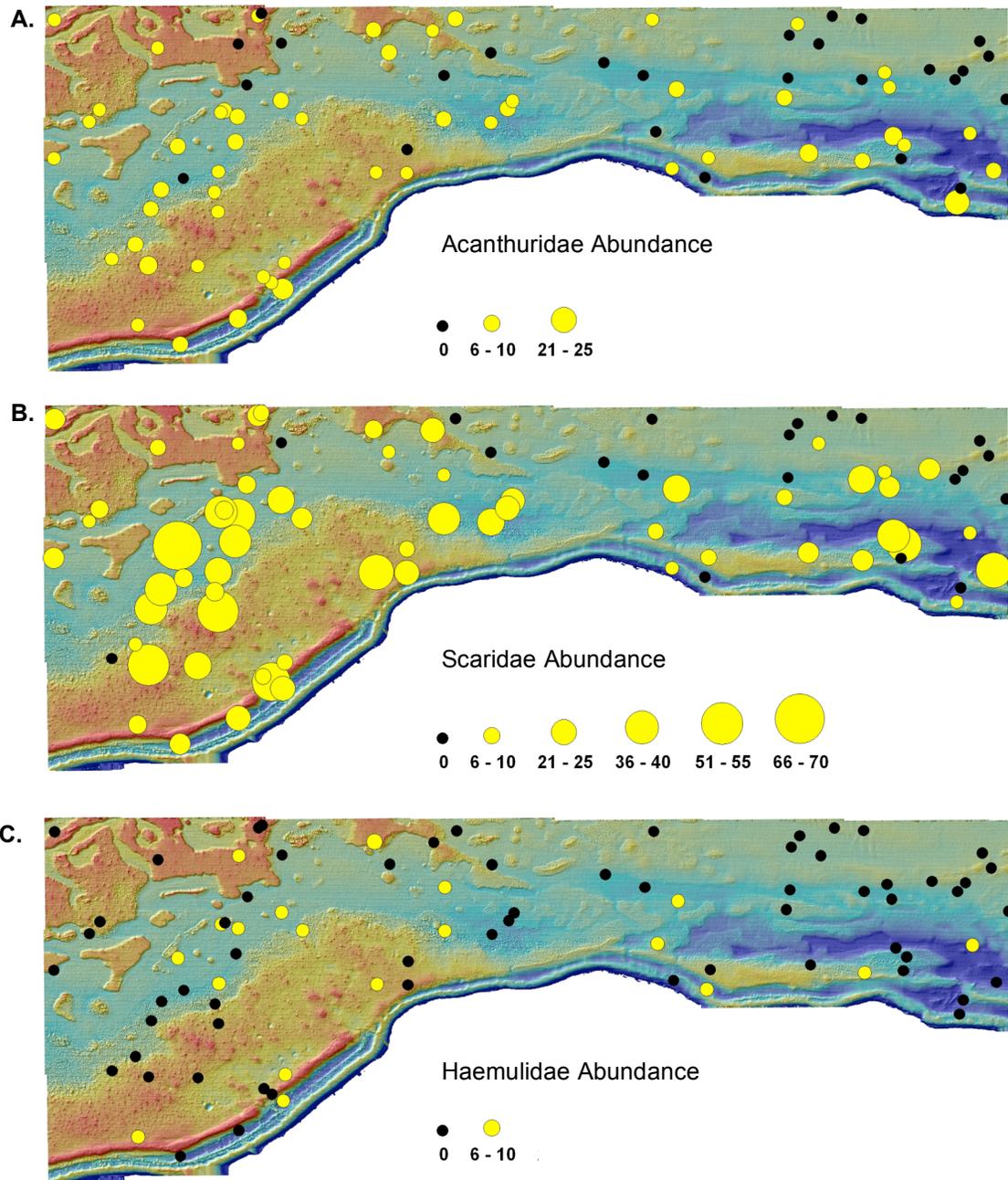


Figure 23. Family abundance (ind. /100m²) of A) Acanthuridae, B) Scaridae, C) Haemulidae, D) Serranidae, E) Lutjanidae, and F) Balistidae at each sampling location in the Marine Conservation District..

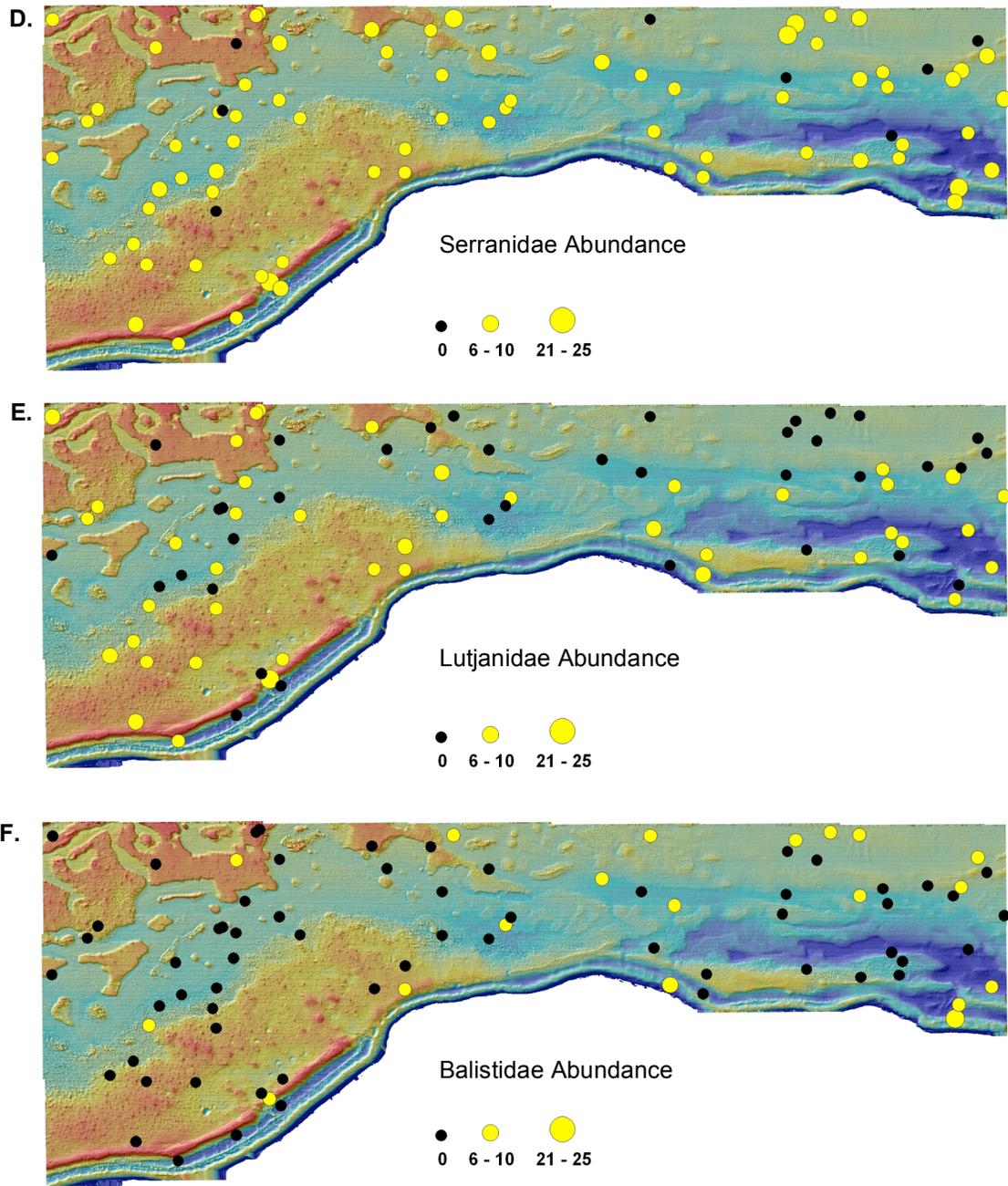


Figure 23. (continued)

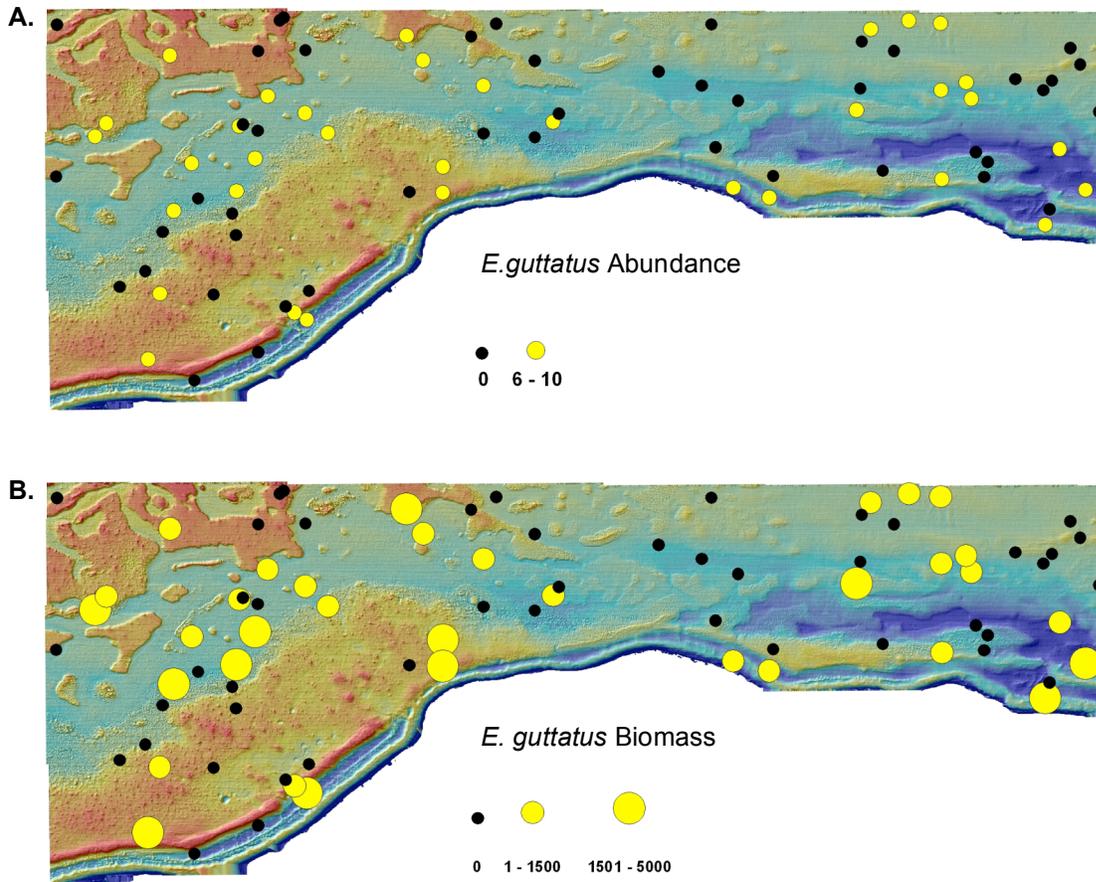


Figure 24. *E. guttatus* A) abundance (ind./100m²) and B) biomass (g/100m²) at each sampling location in the Marine Conservation District..

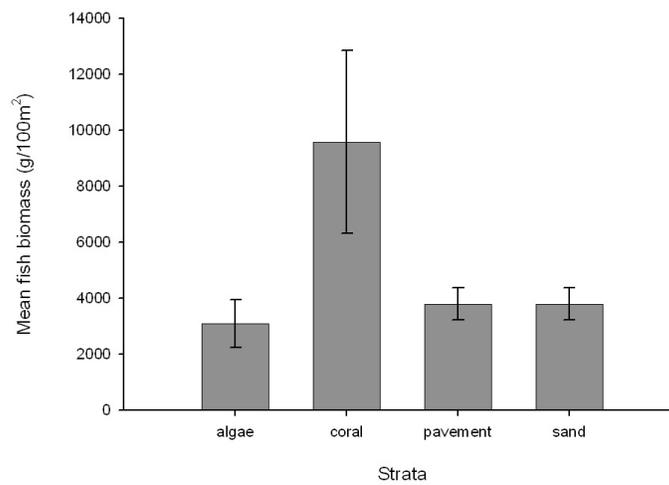


Figure 25. Mean biomass (SE bars) for all fish at each sampling location in the Marine Conservation District.

Fish biomass along belt transects, which ranged from <1 to $>60,000$ g 100m^{-2} , was highest in coral reef habitats, followed by sand, pavement and algal plains (Fig. 25). Due to high variability among transects no significant differences were found among habitat strata. The high biomass in sandy areas was partly due to the presence of significant coral cover that seemed to concentrate fish in these areas. At the family level, the biomass of Acanthurids was fairly uniform throughout sampling sites (Fig. 26A), whereas Scarid biomass was more variable with respect to location (Fig. 26B). Haemulid biomass was relatively high at one coral reef site near the northern boundary of the MCD (Fig. 26C) and Lutjanid biomass was high at several sites located on coral reef and hardbottom habitats (Fig. 26D). Alternatively Balistid biomass was high near the northern boundary and eastern end of the MCD (Fig. 26F) in sand habitat. Serranid biomass was fairly uniform throughout MCD with one high biomass area near the shelf edge (Fig. 26D). Most of the Serranid biomass resulted from red hind being relatively abundant throughout the MCD (Fig. 24).

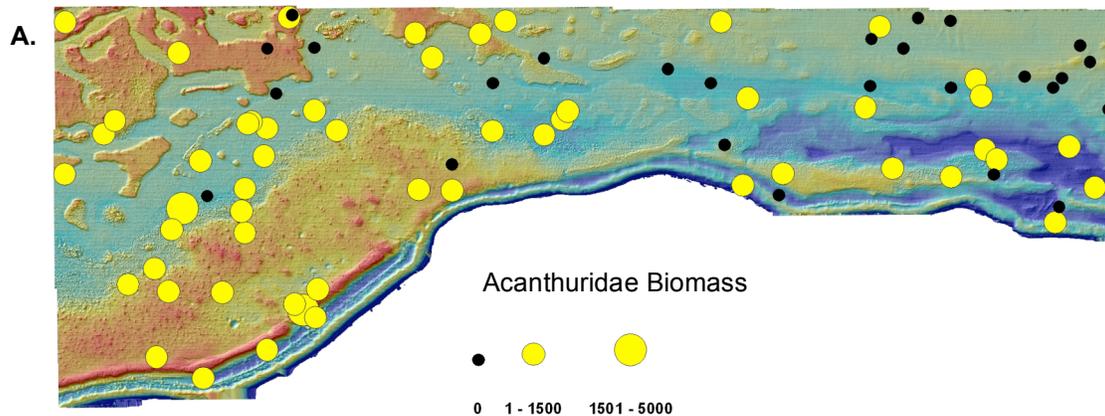


Figure 26. Family biomass (g/100m²) of A) Acanthuridae, (following pages) B) Scaridae, C) Haemulidae, D) Serranidae, E) Lutjanidae, and F) Balistidae at each sampling location in the Marine Conservation District.

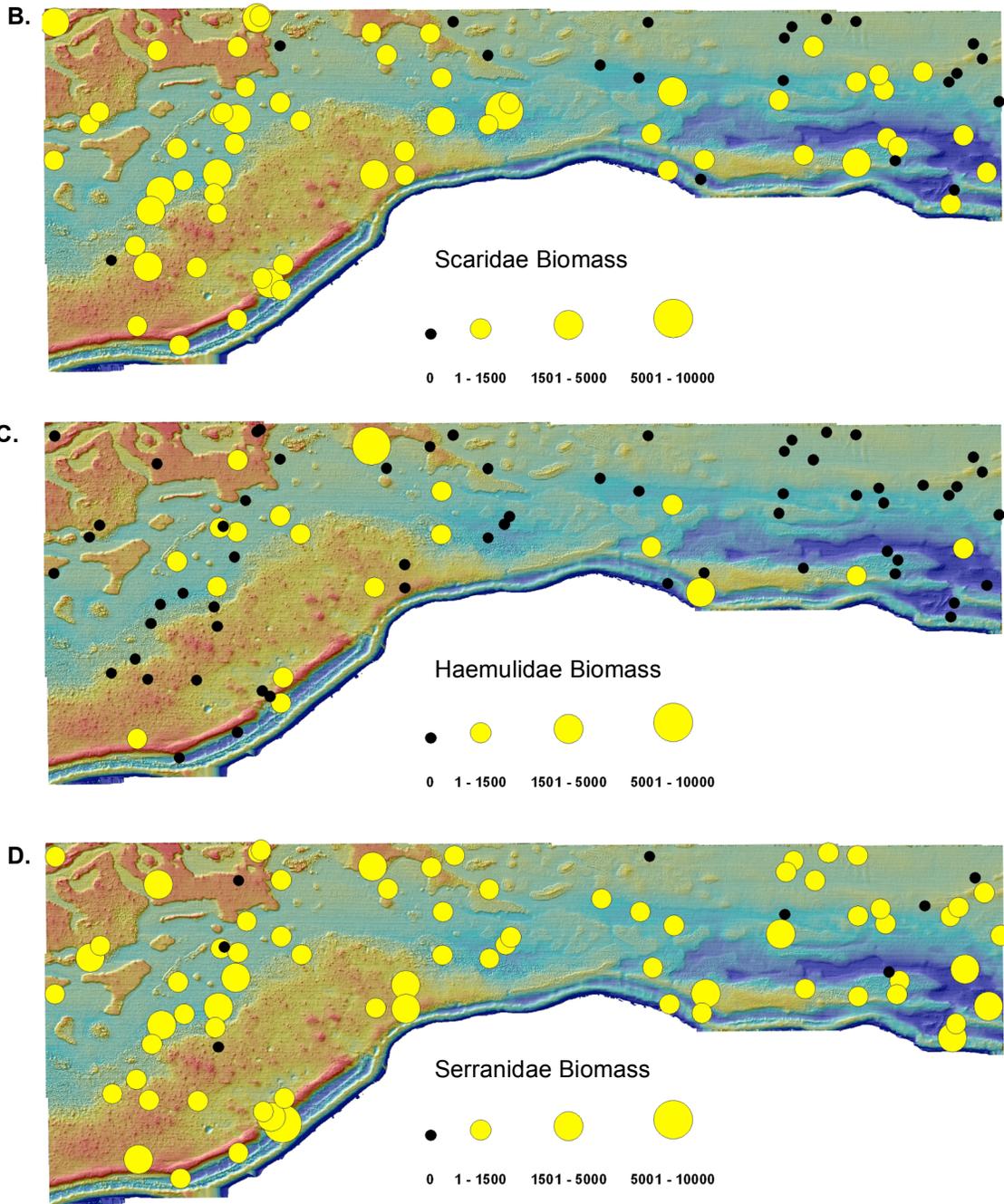


Figure 26. (continued)

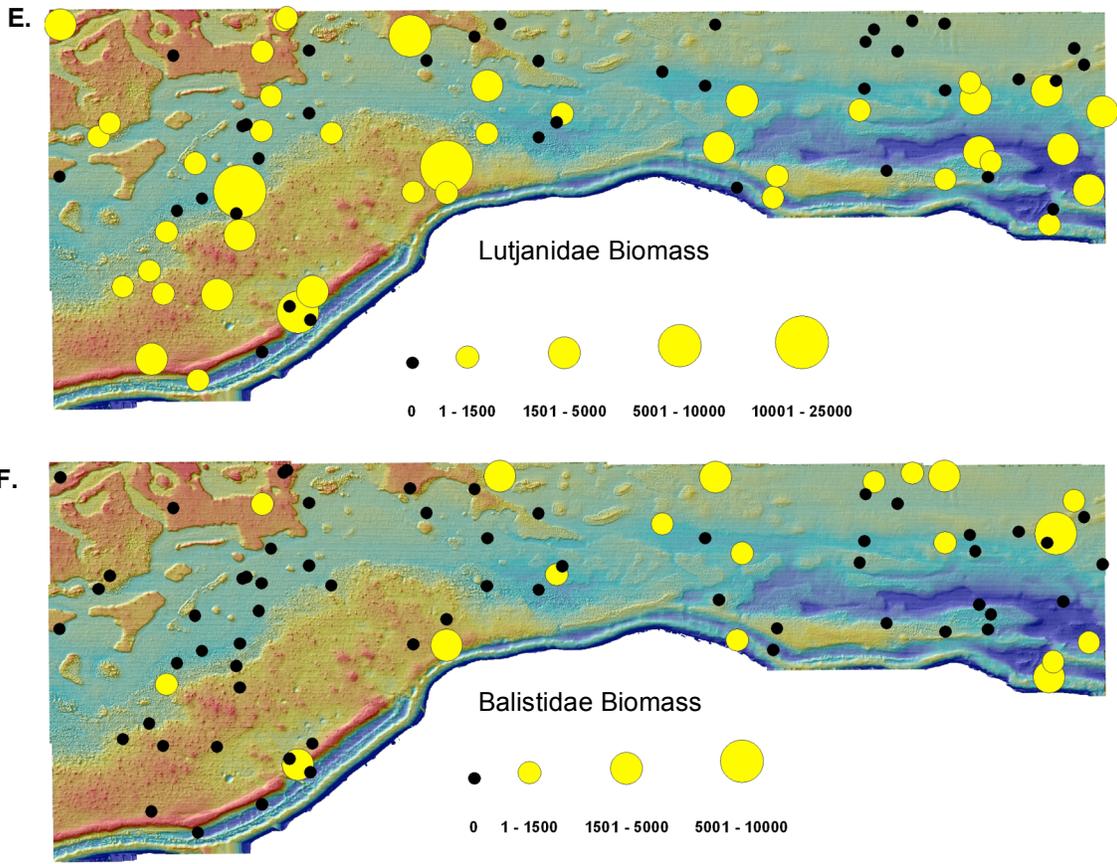


Figure 26. (continued)

Trophic composition among habitat strata

Invertivores were the most common trophic guild and were found at all but one site, with the largest abundances along the southern shelf edge (Fig. 27B). These were followed in abundance by herbivores (Fig. 27A) and omnivores (Fig. 27F). The other three trophic guilds examined, spongivores, piscivores, and planktivores, were similar in abundance (Fig. 27C - E). In general, biomass and richness within trophic guilds followed abundance patterns except for the invertivores and piscivores, which showed spatially distinct biomass patterns.

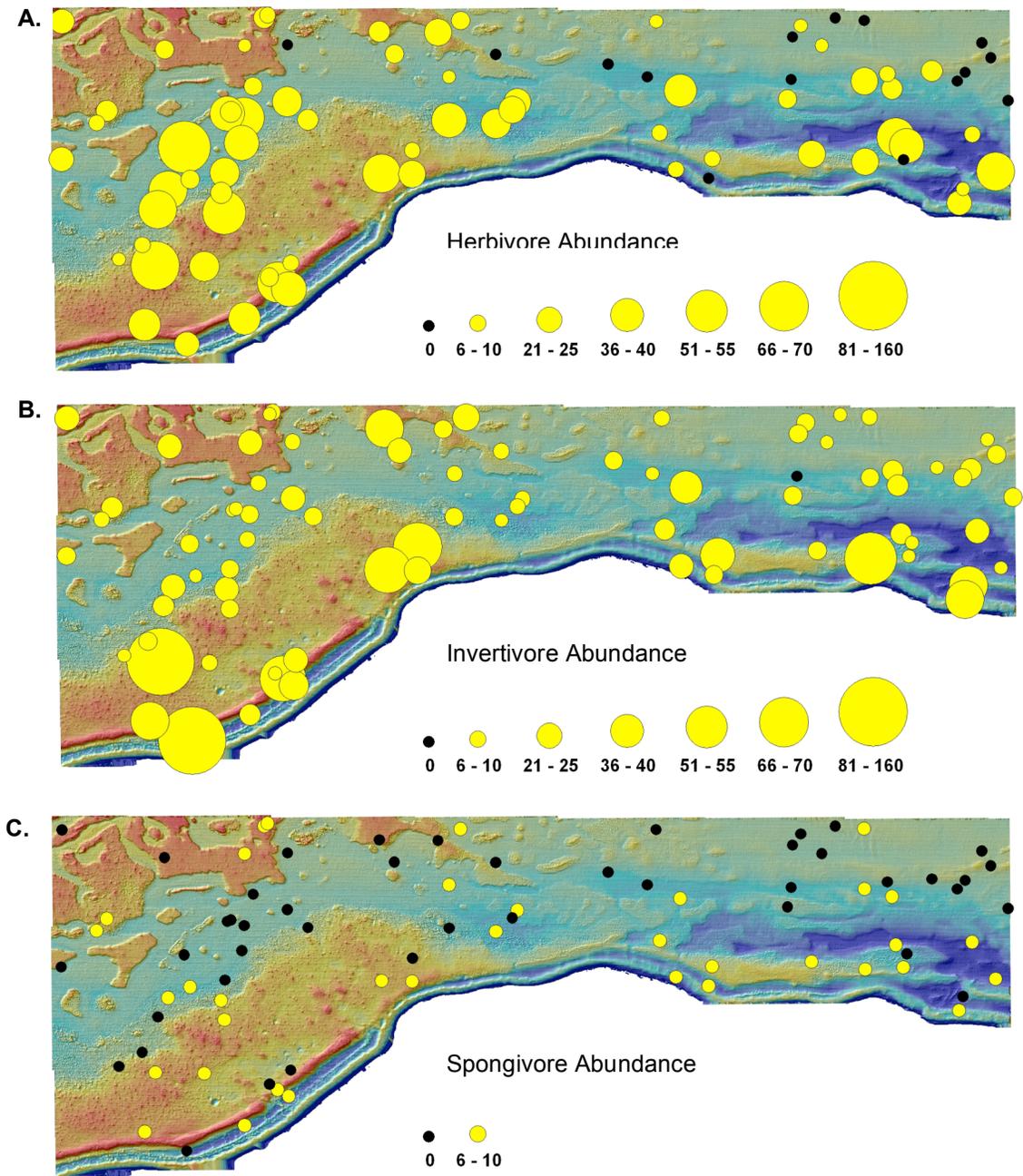


Figure 27. . Trophic level abundance (ind. /100m²) of A) herbivores, B) invertivores, C) spongivores, D) piscivores, E) planktivores, and F) omnivores at each sampling location in the Marine Conservation District.

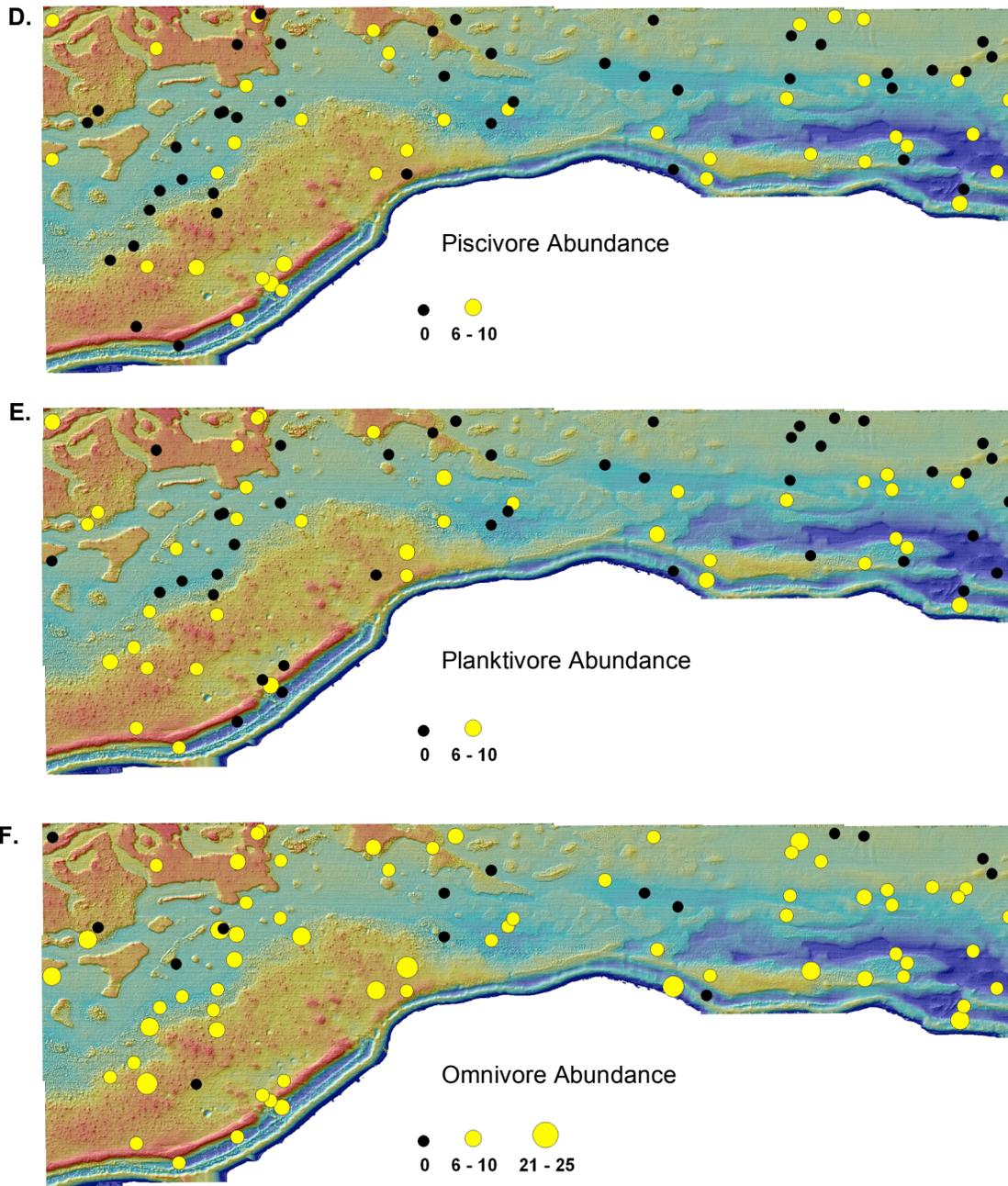


Figure 27. (continued)

Size structure of Reef Fishes

Size structure of selected commercially important species or species groups are shown in (Fig. 28). The herbivorous Acanthurids and Scarids had very similar size distributions, which ranged from 5 to 30 cm total

length and averaged 10.7 cm TL and 11.0 cm TL, respectively (Fig. 28). The two most common Scarids were *Scarus taeneopterus* and *S. iserti*, which, when combined, were approximately three times more abundant than the three common Acanthurid species. The planktivorous yellowtail snapper (*Ocyurus chrysurus*) averaged 24.3 cm FL and ranged from 5 – 50 cm. Red hind (*E. guttatus*) averaged 32.8 cm TL and the queen triggerfish (*B. vetula*) averaged 33.6 cm FL. Both species had a similar size distribution (Fig. 28). The larger and commercially important species of grouper and snapper were rarely counted along transects within the MCD (Table 5).

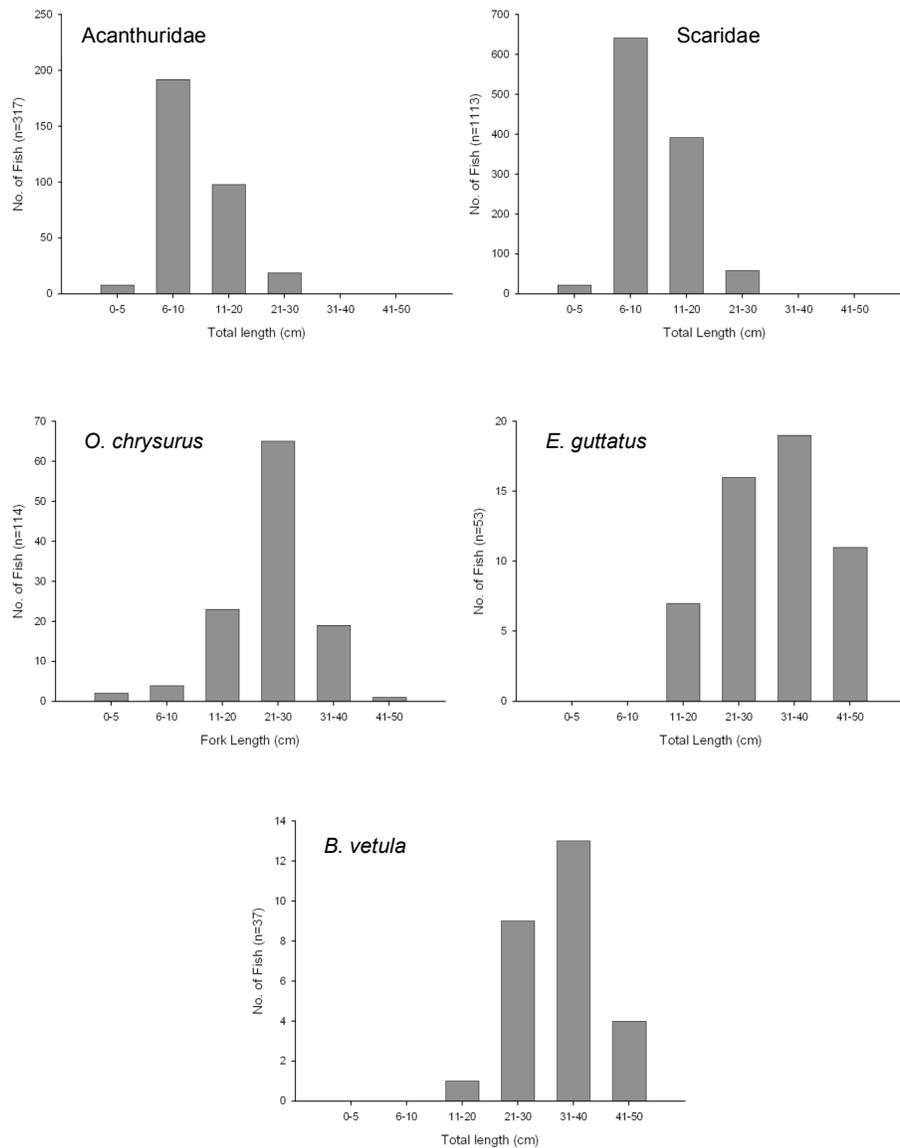


Figure 28. Length frequency histograms for selected commercially important families (Acanthuridae and Scaridae) and species (*O. chrysurus*, *E. guttatus* and *B. vetula*) in all habitat strata combined in the Marine Conservation District..

Table 5. Abundance, size and biomass of commercially important species of grouper and snappers in 80 fish transects from all habitat strata combined in the Marine Conservation District.

Species	N	Size (cm TL)					Total biomass (kg)
		0-20	20-40	40-60	60-80	80-100	
<i>Epinephelus striatus</i>	1	-	-	1	-	-	2.790
<i>Mycteroperca venenosa</i>	2	-	1	1	-	-	2.579
<i>M. tigris</i>	5	-	2	3	-	-	6.277
<i>M. interstitialis</i>	2	-	1	1	-	-	4.051
<i>Lutjanus analis</i>	12	-	7	5	-	-	12.768
<i>L. cyanopterus</i>	5	-	-	4	-	1	34.154
<i>L. jocu</i>	7	-	1	6	-	-	15.567
<i>Lachnolaimus maximus</i>	6	-	2	4	-	-	9.981

Assessment of total fish species diversity used a combination of approaches and reliance on previous fish surveys. The majority of fish diversity data was derived from the 25 x 4 m (100m²) belt transects that were conducted at each of the 80 sampling sites. An additional timed roving diver survey was conducted at one randomly selected site within each habitat strata. Because of the limited time available at each site due to depth constraints we could not conduct a timed roving survey at each site. We also included data that has been collected over several years during annual monitoring at the Hind Bank and College Shoal East study sites, fish collected during research sampling (i.e. fish traps and hook and line) on the red hind spawning aggregation site, and incidental observations during numerous dives within the MCD. The compilation of these data produced a fish list of 122 species within 35 families (Table 6). Many of these species are rare, transient pelagic fishes² that form an important component of the trophic food web.

Due to the depth and complexity of the various habitat strata it is recommended that future surveys include a larger roving diver survey component with a minimum of 60 min search time per site. A portion of these surveys should also include dives during the changeover period during sunrise or sunset to observe the many cryptic and nocturnal species that exist within the complex interstitial spaces of these deep mesophotic reefs.

² This included an approximately 160 kg blue marlin (*Makaira nigricans*) that repeatedly charged one diver pair at a 5 m decompression stop over site Sand 166 on October 8, 2007.

Table 6. Species observed during roving dives in the MCD at fixed sites (sampled 2005-2007) and on four designated habitat types sampled during the MCD survey. Fixed sites, RHB and CSE, are both coral reef habitat. Designated habitat types on the MCD survey include algae (A), coral (C), pavement (P) and sand (S). Abundance categories are 0 (no fish), 1(one fish), 2 (2-10 fish), 3 (11-100 fish) and 4 (101-1000 fish).

Family Species	Common Name	Fixed Sites		MCD Survey			
		RHB	CSE	A	C	P	S
Acanthuridae							
<i>Acanthurus bahianus</i>	ocean surgeonfish	3	3	.	2	3	1
<i>Acanthurus chirurgus</i>	doctorfish	.	2	2	.	2	2
<i>Acanthurus coeruleus</i>	blue tang	2	2	.	2	3	.
Aulostomidae							
<i>Aulostomus maculatus</i>	trumpetfish	.	1
Balistidae							
<i>Balistes vetula</i>	queen triggerfish	1	.	2	.	2	1
<i>Canthidermis sufflamen</i>	ocean triggerfish	2	3
<i>Melichthys niger</i>	black durgeon	2	3
<i>Xanthichthys ringens</i>	sargassum triggerfish	3
Carangidae							
<i>Caranx crysos</i>	blue runner	.	1
<i>Caranx latus</i>	horseeye jack	2	2
<i>Caranx lugubris</i>	black jack	2	3
<i>Caranx ruber</i>	bar jack	1	1	.	2	2	.
<i>Elagatis bipinnulata</i>	rainbow runner	.	1
<i>Seriola dumerili</i>	greater amberjack	1
Carcharhinida							
<i>Negaprion brevirostris</i>	lemon shark	2	2
<i>Galeocerdo cuvier</i>	tiger shark	1	1
<i>Carcharhinus leucas</i>	bull shark	1
<i>Carcharhinus perezii</i>	reef shark	1
Chaetodontidae							
<i>Chaetodon aculeatus</i>	longsnout butterflyfish	2	.	.	2	2	2
<i>Chaetodon capistratus</i>	four-eye butterflyfish	2	3	.	2	2	.
<i>Chaetodon sedentarius</i>	reef butterflyfish	2	1	.	2	2	.
<i>Chaetodon striatus</i>	banded butterflyfish	2	2	.	2	2	.
Coryphaenidae							
<i>Coryphaena hippurus</i>	dolphinfish	1
Echeneidae							
<i>Echeneis naucrates</i>	sharksucker	1

Table 6. (continued)

Family Species	Common Name	Fixed Sites		MCD Survey			
		RHB	CSE	A	C	P	S
Ephippidae							
<i>Chaetodipterus faber</i>	spadefish	.	2
Exocoetidae							
<i>Cheilopogon melanurus</i>	Atlantic flyingfish	3	3	3	3	3	3
Gobiidae							
<i>Coryphopterus dicrus</i>	colon goby	1
<i>Coryphopterus glaucofraenum</i>	bridled goby	.	.	2	2	.	2
Grammatidae							
<i>Gramma loreto</i>	fairy basslet	3	2	1	1	.	.
Haemulidae							
<i>Anisotremus surinamensis</i>	black margate	1	.
<i>Anisotremus virginicus</i>	porkfish	1	2	.	.	2	.
<i>Haemulon album</i>	white grunt	.	.	.	2	.	.
<i>Haemulon aurolineatum</i>	tomtate	.	.	.	3	3	.
<i>Haemulon carbonarium</i>	caesar grunt	2	.	.	.	1	.
<i>Haemulon flavolineatum</i>	French grunt	2	1	.	2	2	.
<i>Haemulon macrostomum</i>	Spanish grunt	0	1	.	.	1	.
<i>Haemulon parra</i>	sailors choice	.	.	.	1	1	.
<i>Haemulon plumieri</i>	white grunt	1	2	.	1	.	.
<i>Haemulon sciurus</i>	bluestriped grunt	2	1	.	1	2	.
Holocentridae							
<i>Holocentrus adscensionis</i>	squirrelfish	.	2	.	2	.	.
<i>Holocentrus coruscum</i>	reef squirrelfish	.	.	.	1	.	.
<i>Holocentrus marianus</i>	longjaw squirrelfish	.	.	.	1	.	.
<i>Holocentrus rufus</i>	longspine squirrelfish	2	.	2	.	2	.
<i>Myripristis jacobus</i>	blackbar soldierfish	2	1	2	3	.	.
Inermiidae							
<i>Inermia vittata</i>	boga	.	2
Istiophoridae							
<i>Makaira nigricans</i>	blue marlin	1
Kyphosidae							
<i>Kyphosus spp.</i>	chub	1	2

Table 6. (continued)

Family Species	Common Name	Fixed Sites		MCD Survey			
		RHB	CSE	A	C	P	S
Labridae							
<i>Bodianus rufus</i>	Spanish hogfish	1	2	.	2	2	.
<i>Clepticus parrae</i>	creole wrasse	4	4	.	.	4	.
<i>Halichoeres bivittatus</i>	slippery dick	1	1	.	.	.	2
<i>Halichoeres garnoti</i>	yellowhead wrasse	3	3	3	2	.	2
<i>Lachnolaimus maximus</i>	hogfish	1	1	.	1	.	.
<i>Thalassoma bifasciatum</i>	bluehead wrasse	3	3	.	2	.	2
Lutjanidae							
<i>Lutjanus analis</i>	mutton snapper	2	.	.	2	.	1
<i>Lutjanus apodus</i>	schoolmaster	4	2	.	3	1	.
<i>Lutjanus buccanella</i>	blackfin snapper	3
<i>Lutjanus cyanopterus</i>	cupera snapper	.	2
<i>Lutjanus griseus</i>	gray snapper	2
<i>Lutjanus jocu</i>	dog snapper	.	2	.	1	.	.
<i>Ocyurus chrysurus</i>	yellowtail snapper	3	2	.	2	2	.
Malacanthidae							
<i>Malacanthus plumieri</i>	sand tilefish	.	.	2	.	.	1
Mobulidae							
<i>Manta birostris</i>	manta ray	1
Mullidae							
<i>Mulloidichthys martinicus</i>	yellow goatfish	2	1	.	2	.	.
<i>Pseudupeneus maculatus</i>	striped goatfish	2	1	3	1	2	.
Muraenidae							
<i>Gymnothorax funebris</i>	green moray	2
<i>Gymnothorax moringa</i>	spotted moray	.	.	1	.	.	.
Myliobatidae							
<i>Aetobatus narinari</i>	spotted eagle ray	1
<i>Lactophrys quadricornis</i>	scrawled cowfish	1	1
<i>Lactophrys triqueter</i>	smooth trunkfish	2	1	.	.	2	.

Table 6. (continued)

Family Species	Common Name	Fixed Sites		MCD Survey			
		RHB	CSE	A	C	P	S
Pomacanthidae							
<i>Centropyge argi</i>	cherub fish	.	.	2	.	.	1
<i>Holacanthus ciliaris</i>	queen angel	2	1	.	2	2	.
<i>Holacanthus tricolor</i>	rock beauty	2	1	.	2	1	.
<i>Pomacanthus arcuatus</i>	gray angelfish	1	.	.	.	2	.
<i>Pomacanthus paru</i>	French angelfish	.	1	.	2	2	.
Pomacentridae							
<i>Chromis cyanea</i>	blue chromis	4	4	.	3	.	.
<i>Chromis multilineata</i>	brown chromis	1	4	.	2	.	.
<i>Stegastes partitus</i>	bicolor damselfish	3	3	2	3	3	3
<i>Stegastes planifrons</i>	threespot damselfish	2	.
<i>Stegastes variabilis</i>	cocoa damselfish	.	.	.	1	.	.
Priacanthidae							
<i>Priacanthus cruentatus</i>	glasseye snapper	1
Rhincodontidae							
<i>Ginglymostoma cirratum</i>	nurse shark	2	2
Scaridae							
<i>Scarus guacamaia</i>	rainbow parrotfish	1
<i>Scarus iserti</i>	striped parrotfish	2	1	.	3	3	.
<i>Scarus taeniopterus</i>	princess parrotfish	2	3	.	2	3	1
<i>Scarus vetula</i>	queen parrotfish	3	2	.	.	1	.
<i>Sparisoma aurofrenatum</i>	redband parrotfish	2	1	2	2	3	2
<i>Sparisoma chrysopteron</i>	redtail parrotfish	.	2
<i>Sparisoma rubripinne</i>	redfin parrotfish	2
<i>Sparisoma viride</i>	stoplight parrotfish	2	3	.	2	3	.
Scombridae							
<i>Scomberomorus regalis</i>	cero mackerel	1	2
<i>Scomberomorus cavalla</i>	king mackerel	1
<i>Acanthocybium solandri</i>	wahoo	1

Table 6. (continued)

Family Species	Common Name	Fixed Sites		MCD Survey			
		RHB	CSE	A	C	P	S
Serranidae							
<i>Epinephelus striatus</i>	Nassau grouper	.	1
<i>Epinephelus cruentatus</i>	graysby	2	1	.	2	1	.
<i>Epinephelus fulvus</i>	coney	1	.	2	2	2	2
<i>Epinephelus guttatus</i>	red hind	2	.	.	1	2	2
<i>Hypoplectrus chlorurus</i>	yellowtail hamlet	2	.	.	2	2	.
<i>Hypoplectrus nigricans</i>	black hamlet	1	.	.	1	.	.
<i>Hypoplectrus puella</i>	barred hamlet	2	.	.	1	.	.
<i>Hypoplectrus unicolor</i>	butter hamlet	1	1	.	.	0	.
<i>Mycteroperca interstitialis</i>	yellowmouth grouper	1
<i>Mycteroperca tigris</i>	tiger grouper	2	1
<i>Mycteroperca venenosa</i>	yellowfin grouper	2	1
<i>Liopropoma rubre</i>	peppermint basslet	1
<i>Paranthias furcifer</i>	creole-fish	3	1
<i>Serranus baldwini</i>	lantern bass	.	.	3	.	.	4
<i>Serranus tabacarius</i>	tobaccofish	1	.	2	.	.	2
<i>Serranus tigrinus</i>	harlequin bass	1	1	2	.	.	2
<i>Serranus tortugarum</i>	chalkfish	.	.	2	.	.	.
Sparidae							
<i>Calamus spp.</i>	porgy	1	.	.	2	.	1
Sphyraenidae							
<i>Sphyraena barracuda</i>	great barracuda	1	1	2	1	.	.
Tetrodontidae							
<i>Canthigaster rostrata</i>	sharpnose puffer	1	2	.	2	.	1

CONCLUSIONS AND RECOMMENDATIONS

Improved Evaluation of EFH within the MCD

- Benthic habitat classification from sonar imagery was capable of distinguishing major blocks of habitat, but was less able to correctly allocate those blocks to the habitat they represented. This may be a problem of novel habitat types, such as low relief coral reef habitat types composed of plating corals, which are not represented in benthic habitat algorithms developed for shallow-water communities. A formal re-assessment of the of the benthic maps using the recently acquired in situ surveys should be a priority and will improve the next generation of habitat classification models for Caribbean mesophotic systems.
- Based on the limited sampling within the 39.5 km² of the MCD shallower than 50 m, we found a large diversity of reef fishes (112 species total) from all trophic groups. We anticipate that the number of species will continue to increase as sampling efforts expand to include more sites. The relationship between fish species richness and habitat type was only distinct between algae and all other habitats. The three other habitats (coral, pavement and sand), as classified in the original benthic mapping, had considerable overlap in habitat features. Specifically the sand habitat included up to 60% variable hard bottom, which contributed to a high degree of species richness. The influence of edges was also apparent. Many of the large piscivores were found along the transition between coral reef and pavement, sand and algal habitats. It is recommended that these transition zones between habitats be included as a separate classification in future biodiversity surveys.
- Large areas of coral reefs within the MCD are dominated by extensive interstitial space created by holes, tunnels, channels and ledges. These areas likely contain a high diversity of invertebrates and cryptic fishes, may contain commercially important species, such as the Caribbean spiny lobster (*Panularis argus*), and could serve as essential fish habitat. Effective surveys of these habitats could be accomplished with stationary visual censuses.
- While most species represented were adults, juveniles and sub-adults of many species were also present. This was especially apparent in the herbivorous Scaridae as well as the omnivorous Labridae and the planktivorous Pomacentridae. These species may well form the base of the food web for the larger piscivores (groupers, snappers), which in turn provide food for the top predators (sharks). In the absence of fishing, we anticipate that this large deepwater ecosystem will begin to show significant changes in the trophic structure of reef fishes and subsequent resilience to natural perturbations. The protection of the red hind spawning aggregation has already resulted significant improvements in the red hind population, and has improved the fishery as well (Nemeth 2005). Potential spill-over of commercially important reef fishes from the MCD to the surrounding

areas is anticipated but needs to be studied to better quantify the benefits of this closure to the local fishery (see below).

- Considerable habitat within the MCD lies on the outer shelf edge at depths greater than 50m. Dives to 70 m along this margin revealed a unique foliose agariciid coral reef community that may extend linearly along the shelf edge in suitable depths. This area may be particularly important as a movement corridor and as habitat for commercially important species, such as Nassau and yellowfin grouper, and blackfin snapper (Authors, unpub. obs.). Surveys by scientific divers in these deeper habitats are feasible with recent improvements in the reliability of technical diving systems (closed circuit rebreathers).
- Video methods offer minimum estimates of species richness for epifauna, with the exception of scleractinian corals. Better estimates of species richness and diversity could be obtained by in situ identification along belt transects or within quadrats.
- Little is known about the seasonality of the benthos, and motile fish and invertebrate communities in the MCD, nor the processes (e.g., oceanographic forcing) that drive seasonality. More intensive longitudinal studies could resolve these patterns and processes

Expansion of the Evaluation of EFH Outside the MCD

- Evaluation of the effectiveness of the MCD in promoting and sustaining regional fisheries via spillover requires comparative surveys outside the MCD. This could help to assure stakeholders that the reserve is functioning to improve local economic and ecological condition, at the expense of access to traditional fishing grounds.
- Comparative surveys outside the MCD would also serve to describe the extent of mesophotic reefs and associated species. These reef systems are probably widespread on the Puerto Rican Shelf and are likely critical fisheries areas.
- Other federal marine protected areas in the U.S. Virgin Islands would benefit from EFH assessment. These include the Grammanik Bank, Lang Bank, and the Mutton Snapper closed areas. Comparative studies could be conducted between these three CFMC management areas, and might provide valuable insights into factors affecting successful management strategies and avenues for improved management.
- In addition to closed areas, large areas of the Puerto Rican Shelf and the St. Croix shelf have recently been surveyed with high-resolution multibeam sonar by NOAA. In situ surveys in these areas could be useful, not only for expanding knowledge of mesophotic and mid-depth systems (see above), but also for validating improved benthic habitat classification and resource prediction models,

- Comparative surveys outside the MCD would also help to determine the processes controlling the formation and degradation of mesophotic reefs. The capture of a severe cryptic disease/mortality event outside annual monitoring locations, and recent work by Menza et al. (2007), underscores the lack of knowledge of processes that shape these deeper coral reef environments. These assessments will be crucial to understanding the potential of mesophotic reef systems to serve as refugia during a period of increasing sea surface temperatures and increasing frequency of coral bleaching events.

ACKNOWLEDGEMENTS

We would like to thank the following people and organizations. Additional funding to support fieldwork was provided by the National Oceanographic and Atmospheric Administration, National Ocean Sciences Biogeography Program through the USVI Division of Coastal Zone Management (Department of Planning and Natural Resources), and the US National Science Foundation through the USVI Experimental Program to Stimulate Competitive Research (USVI NSF-EPSCoR) and the Lana Vento Charitable Trust. Excellent administrative support was given by C. Joseph. We are also indebted to the field assistance and wisdom of K. Turbe, R. Garcia, M. Carlos, J. Sabater, and K. Brown. All views expressed are those of the authors and do not necessarily reflect the views of the granting agencies and support personnel. This is contribution #102 from the Center for Marine and Environmental Studies at the University of the Virgin Islands.

LITERATURE CITED

- Armstrong RA, Singh H, Torres J, Nemeth RS, Can A, Roman C, Eustice R, Riggs L, Garcia-Moliner G (2006) Characterizing the deep insular shelf coral reef habitat of the Hind Bank marine conservation district (US Virgin Islands) using the Seabed autonomous underwater vehicle. *Continental Shelf Research* 26:194-205
- Aronson RB, Edmunds PJ, Precht WF, Swanson DW, Levitan DR (1994) Large-scale, long-term monitoring of Caribbean coral reefs: simple, quick, inexpensive techniques. *Atoll Research Bulletin* 421:1-19
- Beets J, Friedlander A (1999) Evaluation of a conservation strategy: a spawning aggregation closure for red hind, *Epinephelus guttatus*, in the U.S. Virgin Islands. *Environ Biol Fishes* 55:91-98
- Bruckner AW (2007) Field Guide to Western Atlantic Coral Diseases and Other Causes of Coral Mortality. NOAA, UNEP-WCMC, PADI
- Carleton JH, Done TJ (1995) Quantitative video sampling of coral reef benthos: Large-scale application. *Coral Reefs* 14:35-46
- Donner S, Knutson T, Oppenheimer M (2007) Model-based assessment of the role of human-induced climate change in the 2005 Caribbean coral bleaching event. *Proceedings of the National Academy of Science* 104:5483-5488
- Gardner TA, Côté IM, Gill JA, Grant A, Watkinson AR (2003) Long-term region-wide declines in Caribbean corals. *Science* 301:958-960
- Herzlieb S, Kadison E, Blondeau J, Nemeth RS (2006) Comparative assessment of coral reef systems located along the insular platform south of St. Thomas, US Virgin Islands and the relative effects of natural and human impacts. *Proc 10th Int Coral Reef Symp* 4-2:1144-1151
- Kramer P, Lang J, Marks K, Garza-Perez R, Ginsburg R (2005) AGRRA Methodology, version 4.0, June 2005. University of Miami, Miami
- Menza C, Ault J, Beets J, Bohnsack J, Caldow C, Christensen J, Friedlander A, Jeffrey C, Kendall M, Luo J, Monaco M, Smith S, Woody K (2006) A Guide to Monitoring Reef Fish in the National Park Service's South Florida/Caribbean Network. NOAA Technical Memorandum. NOS NCCOS 39 166pp.
- Menza C, Kendall M, Rogers C, Miller J (2007) A deep reef in deep trouble. *Continental Shelf Research* 27:2224-2230
- Moody G (2003) Marine Habitat Mapping Offshore St. Thomas & St. Croix, U.S. Virgin Islands EEZ. Géophysique GPR International Inc., Montreal 30pp.
- Nemeth RS (2005) Population characteristics of a recovering US Virgin Islands red hind spawning aggregation following protection. *Marine Ecology Progress Series* 286:81-97

- Nemeth RS, Quandt A (2005) Differences in fish assemblage structure following the establishment of the Marine Conservation District, St. Thomas U. S. Virgin Islands, Vol 56. Proceedings of the 56th Gulf and Caribbean Fisheries Institute. Tortola, BVI.:367-381
- Nemeth RS, Smith TB, Taylor M, Herzlieb S, Kadison E, Blondeau J, Carr L, Allen-Requa L (2006a) Coral Reef Monitoring in St. Croix and St. Thomas, United States Virgin Islands. Year Five Final Report. University of the Virgin Islands, St. Thomas 142pp.
- Nemeth RS, Kadison E, Herzlieb S, Blondeau J, Whiteman E (2006b) Status of a yellowfin grouper (*Mycteroperca venenosa*) spawning aggregation in the US Virgin Islands with notes on other species. Proc. 57th Gulf Carib Fish Inst. 57:543-558
- NOAA-NOS (2001) Benthic Habitats of Puerto Rico and the U.S. Virgin Islands. National Oceanic and Atmospheric Administration
- Olsen D, LaPlace J (1978) A study of Virgin Islands grouper fishery based on a breeding aggregation. Proc 31st Gulf Caribb Fish Inst 31:130-144
- Prada M (2003) Delineation of Benthic Habitats from Side Scan Sonar mosaics from the Marine Conservation District, St. Thomas, Lang Bank and Mutton Snapper Closed Area in St. Croix, US Virgin Islands. Géophysique GPR International, Inc, Montreal 22pp.
- Rivera J, Prada M, Arsenault J-L, Moody G, Benoit N (2006) Detecting Fish Aggregations from Reef Habitats Mapped with High Resolution Side Scan Sonar Imagery. NOAA Professional Paper NMFS 5: 88-104
- Rogers C, Miller J (2001) Coral bleaching, hurricane damage, and benthic cover on coral reefs in St. John, U.S. Virgin Islands: a comparison of surveys with the chain transect method and videography. Bulletin of Marine Science 69:459-470
- Rogers CS, Garrison G, Grober R, Hillis Z, Franke M (2001) Coral Reef Monitoring Manual for the Caribbean and Western Atlantic. US National Park Service, St. John
- Smith TB, Nemeth RS, Calnan JM, Taylor M, Kadison ES, Blondeau J, Tyner E (2007) Coral Reef Monitoring in St. Croix and St. Thomas, United States Virgin Islands. Year Six Final Report. University of the Virgin Islands, St. Thomas 52pp.
- Smith T, Calnan J, Nemeth R, Carr L, Requa-Allen L, Kadison E, Blondeau J, Taylor M, Rothenberger J (in prep) Impacts of the 2005 Caribbean mass bleaching event in the U.S. Virgin Islands
- Smith T, Nemeth R, Blondeau J, Calnan J, Kadison E, Herzlieb S (in review) Assessing coral reef health across onshore to offshore stress gradients in the US Virgin Islands
- Whiteman EA, Jennings CA, Nemeth RS (2005) Sex structure and potential female fecundity in a red hind (*Epinephelus guttatus*) spawning aggregation: applying ultrasonic imaging. J Fish Biol 66:983-995

APPENDICES**Appendix I** Sampling locations in the Marine Conservation District and their general characteristics.

Strata	Location	Lat	Long	Depth (m)	Date Sampled
Algae	MCD A 1	18.21919744	-65.05030908	42	10/4/07
Algae	MCD A 10	18.21244379	-65.009961	43	12/5/07
Algae	MCD A 11	18.20610503	-64.98787572	49.7	12/7/07
Algae	MCD A 12	18.21506709	-64.98559179	42	12/5/07
Algae	MCD A 14	18.21329212	-64.9887041	43	10/29/07
Algae	MCD A 15	18.20710693	-65.02136999	48	11/28/07
Algae	MCD A 16	18.21925007	-65.04407546	42	10/4/07
Algae	MCD A 17	18.21949375	-64.98597395	43	11/29/07
Algae	MCD A 18	18.21494988	-65.01856664	45	12/5/07
Algae	MCD A 19	18.21925069	-65.04407546	43	10/29/07
Algae	MCD A 2	18.21679587	-64.98672594	42	10/24/07
Algae	MCD A 20	18.20824479	-65.01694758	45.2	12/17/07
Algae	MCD A 21	18.21248121	-65.02591478	43	12/7/07
Algae	MCD A 3	18.21329212	-64.9887041	42	12/6/07
Algae	MCD A 4	18.21309074	-64.99823373	43.3	11/29/07
Algae	MCD A 5	18.21702446	-65.01869904	40.6	10/25/07
Algae	MCD A 6	18.21644204	-65.00624863	41.8	11/28/07
Algae	MCD A 7	18.21230098	-65.00099196	44	10/25/07
Algae	MCD A 8	18.21870191	-65.00884722	41.5	11/28/07
Algae	MCD A 9	18.21268362	-65.02757053	45	10/29/07
Coral	MCD C 51	18.19190106	-65.08136049	38	11/6/07
Coral	MCD C 52	18.19680569	-65.07908419	38.5	12/4/07
Coral	MCD C 53	18.20295433	-65.09902033	40	12/4/07
Coral	MCD C 54	18.21140366	-64.99764917	43	10/25/07
Coral	MCD C 55	18.18369391	-65.088798	36	11/12/07
Coral	MCD C 56	18.20718602	-65.09472463	43	10/12/07
Coral	MCD C 57	18.21783608	-65.05311238	44	11/28/07
Coral	MCD C 58	18.2062122	-65.02601558	41	10/9/07
Coral	MCD C 59	18.2040901	-65.05618887	38	10/4/07

Strata	Location	Lat	Long	Depth (m)	Date Sampled
Coral	MCD C 60	18.20317584	-65.01958624	39	11/28/07
Coral	MCD C 61	18.20133302	-65.05618714	35	12/5/07
Coral	MCD C 62	18.21579034	-65.08645536	34	12/5/07
Coral	MCD C 63	18.21983358	-65.07391212	34	12/5/07
Coral	MCD C 64	18.20857843	-65.09350606	42	10/26/07
Coral	MCD C 65	18.21953265	-65.07429684	36	11/7/07
Coral	MCD C 66	18.20143394	-65.05989331	35	10/4/07
Coral	MCD C 67	18.21345617	-64.99281415	42	10/24/07
Coral	MCD C 68	18.18867811	-65.07260203	34	12/4/07
Coral	MCD C 69	18.21625361	-65.0766646	37	11/7/07
Coral	MCD C 70	18.20476208	-64.99586234	45	10/15/07
Pavement	MCD P 101	18.21115147	-65.02349829	44	10/24/07
Pavement	MCD P 102	18.19062453	-65.08751514	38.5	12/4/07
Pavement	MCD P 103	18.21899776	-65.0990227	36.4	10/8/07
Pavement	MCD P 104	18.19062453	-65.08751514	36	10/5/07
Pavement	MCD P 105	18.21020925	-65.01042807	46	10/24/07
Pavement	MCD P 106	18.2085	-65.07838436	43	11/7/07
Pavement	MCD P 107	18.19091406	-65.0710052	41	10/15/07
Pavement	MCD P 108	18.2072325	-65.04598339	45	11/27/07
Pavement	MCD P 109	18.19091406	-65.0710052	41	10/4/08
Pavement	MCD P 110	18.21267149	-65.05171893	43	12/5/07
Pavement	MCD P 111	18.20883751	-65.04400212	46	10/21/07
Pavement	MCD P 112	18.20491757	-65.07700044	43	11/30/07
Pavement	MCD P 113	18.18449445	-65.07658781	39	11/6/07
Pavement	MCD P 114	18.18929168	-65.07357517	39	11/6/07
Pavement	MCD P 115	18.2076195	-65.05173277	40	11/27/07
Pavement	MCD P 116	18.19190106	-65.08136049	38	11/7/07
Pavement	MCD P 117	18.20974776	-65.07146111	43	10/30/07
Pavement	MCD P 118	18.20971644	-65.04337465	41	10/29/07
Pavement	MCD P 119	18.20781846	-65.07670851	43	10/9/07
Pavement	MCD P 120	18.20442269	-65.084038	44	11/7/07
Sand	MCD S 151	18.20310948	-64.99622517	45	10/15/07
Sand	MCD S 152	18.20289399	-65.0009099	43	10/15/07

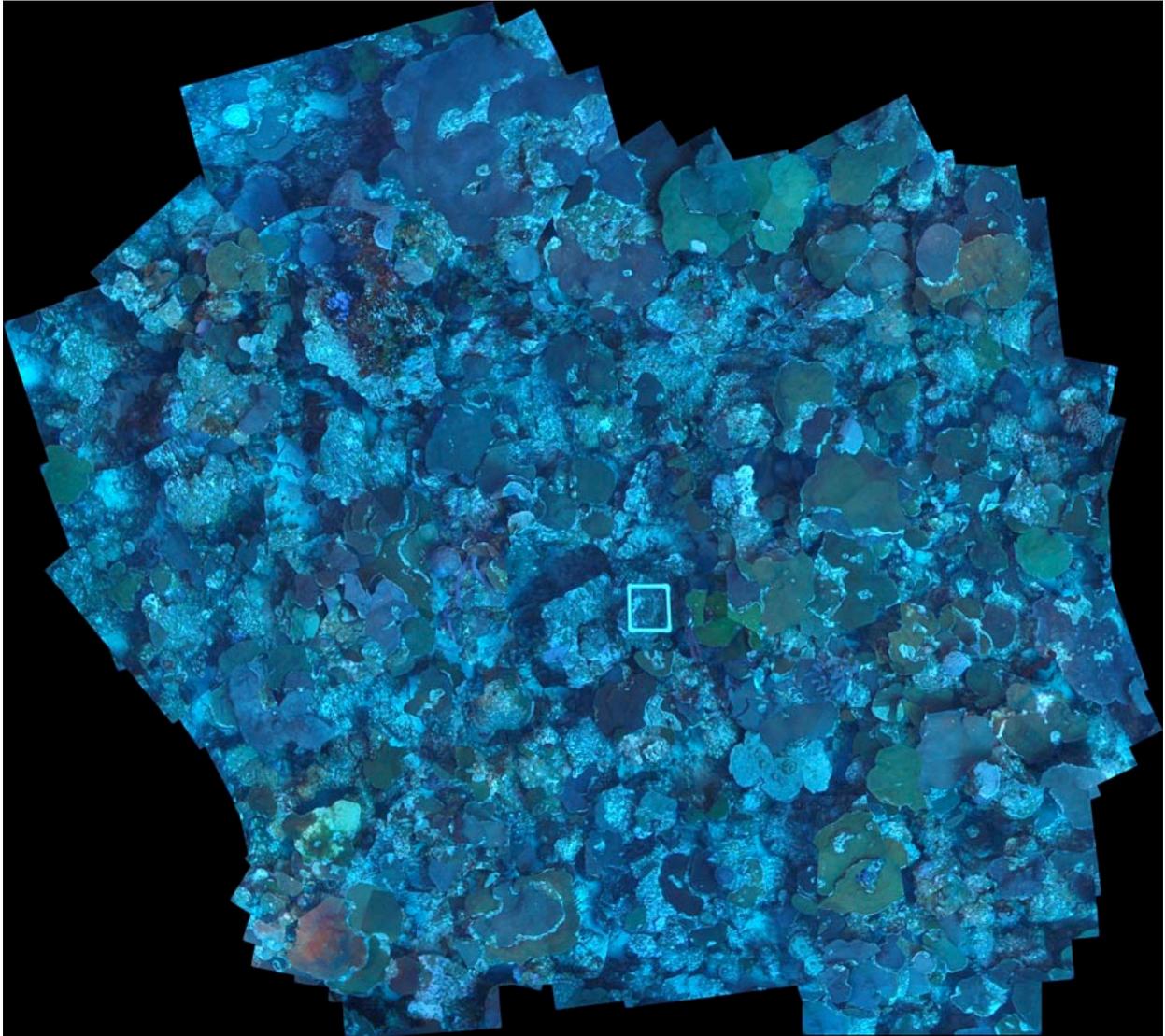
Strata	Location	Lat	Long	Depth (m)	Date Sampled
Sand	MCD S 153	18.19811209	-64.98942559	44	10/25/07
Sand	MCD S 154	18.2057528	-64.99717632	47	11/29/07
Sand	MCD S 155	18.21150359	-65.07562486	43	10/30/07
Sand	MCD S 157	18.1930068	-65.08909274	36	10/26/07
Sand	MCD S 158	18.19974027	-64.98898819	50.9	12/6/07
Sand	MCD S 159	18.20193636	-65.02400739	40	10/9/08
Sand	MCD S 160	18.18786959	-65.07122747	41	11/12/07
Sand	MCD S 161	18.19710077	-65.08721552	43	11/19/07
Sand	MCD S 163	18.19908495	-65.07949955	37	1/30/08
Sand	MCD S 164	18.2100459	-64.98351194	43	11/29/07
Sand	MCD S 165	18.19132967	-65.09198426	39	12/4/07
Sand	MCD S 166A	18.19939785	-65.08599282	42	10/8/07
Sand	MCD S 167	18.18146399	-65.08364693	44	11/12/07
Sand	MCD S 168	18.20763223	-65.0689303	38	10/9/07
Sand	MCD S 169	18.20093822	-65.02001809	50	12/6/07
Sand	MCD S 170	18.20065975	-65.08328427	44	1/11/08
Sand	MCD S 172	18.20143232	-65.07907089	44.2	12/17/07
Sand	MCD S 173	18.21532859	-65.05840878	42.4	12/17/07

Appendix II Electronic supplement (DVD). Doted images used in benthic composition assessments.

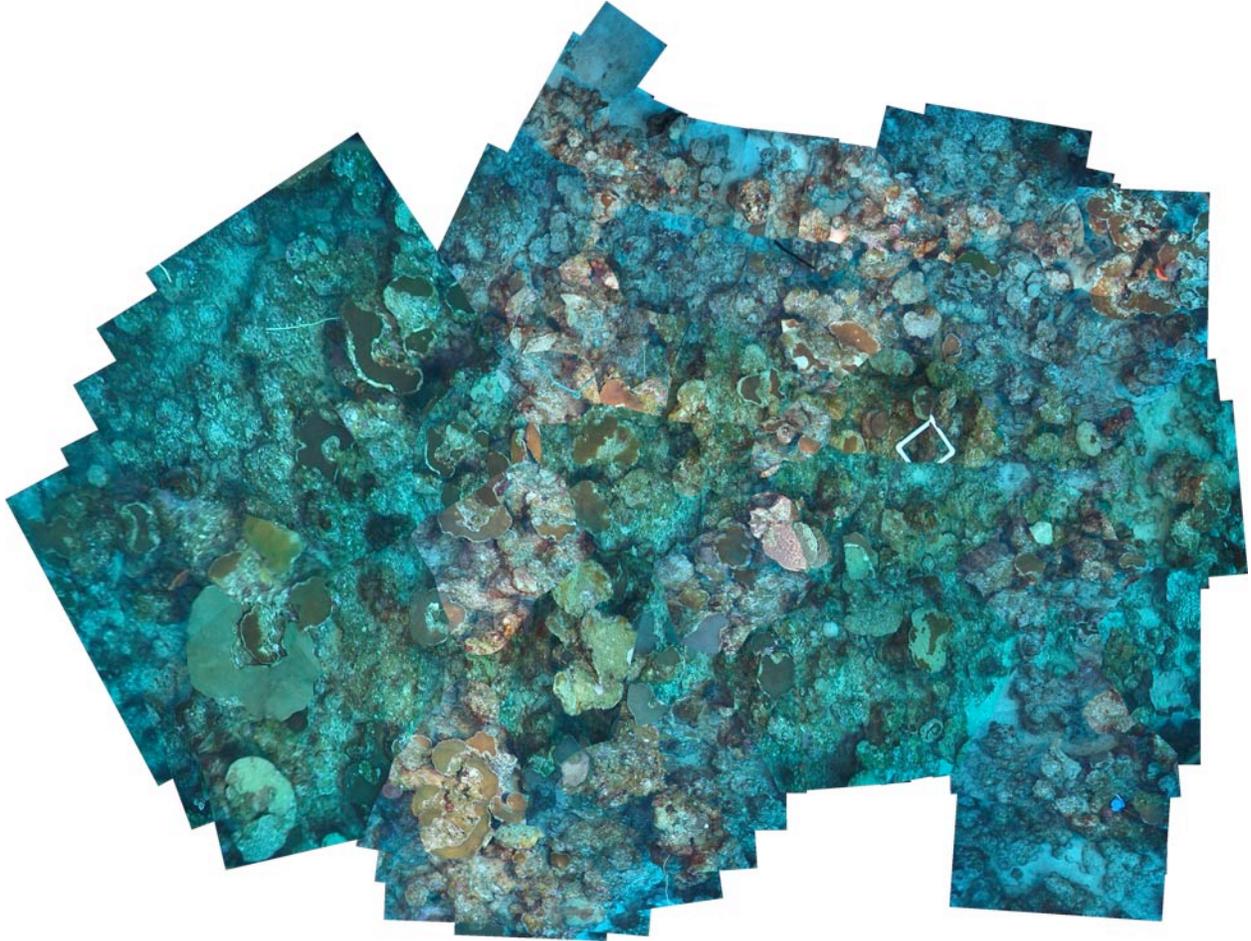
Appendix III Electronic supplement (DVD). Video captures of each sampling location.

Appendix IV Video mosaic images of representative coral reef locations from the strata coral (Coral 52) and sand (Sand 166). Scaling quadrat is 25 X 25 cm. Video mosaic analysis courtesy of A. Gleason and P. Reid (RSMAS-University of Miami).

Coral 52. January 16, 2008



S166. January 11, 2008



Appendix V Electronic supplement (DVD). Benthic cover data for each location sampled in the Marine Conservation District.

Appendix VI Electronic supplement (DVD). Coral health data for coral harboring locations sampled in the Marine Conservation District.

Appendix VII Electronic supplement (DVD). Fish data for each location sampled in the Marine Conservation District.

Appendix VIII Literature Review: literature pertinent to the Marine Conservation District

Marine Conservation District Habitat Project Literature Review

Aponte NE, Ballantine DL (2001) Depth distribution of algal species on the deep insular fore reef at Lee Stocking Island, Bahamas. *Deep Sea Research Part I* 48:2185-2194

Appeldoorn R (1985) Support for the assessment of deepwater resources and evaluation of passive gears around Puerto Rico and the Virgin Islands. *Caribbean Fisheries Management Council* 24

Armstrong RA (2007) Deep zooxanthellate coral reefs of the Puerto Rico: US Virgin Islands insular platform. *Coral Reefs* 26:945

Armstrong RA, Singh H, Torres J, Nemeth RS, Can A, Roman C, Eustice R, Riggs L, Garcia-Moliner G (2006) Characterizing the deep insular shelf coral reef habitat of the Hind Bank marine conservation district (US Virgin Islands) using the Seabed autonomous underwater vehicle. *Continental Shelf Research* 26:194-205

Armstrong R, Singh H, Torres J (2002) Benthic survey of insular slope coral reefs using the Seabed AUV. *Backscatter* 13:22-25

Asch RG, Turgeon DD (2003) Detection of gaps in the spatial coverage of coral reef monitoring projects in the US Caribbean and Gulf of Mexico. *Revista de Biología Tropical* 51:127-140

- Bak RPM, Nieuwland G (1995) Long-term changes in coral communities along depth gradients over leeward reefs in the Netherlands Antilles. *Bulletin of Marine Science* 56:609-619
- Bak R, Nieuwland G, Meesters E (2005) Coral reef crisis in deep and shallow reefs: 30 years of constancy and change in reefs of Curacao and Bonaire. *Coral Reefs* 24:475-479
- Ballantine D, Aponte N (1995) Deep-water algal distribution in the Bahamas, West Atlantic. *Journal of Phycology* 31:10
- Beets J, Friedlander AM (1997) Evaluation of the spawning aggregation closure for red hind (*Epinephelus guttatus*), St. Thomas, US Virgin Islands. Caribbean Fishery Management Council, San Juan, P.R. 17
- Beets J, Friedlander A (1999) Evaluation of a conservation strategy: a spawning aggregation closure for red hind, *Epinephelus guttatus*, in the U.S. Virgin Islands. *Environ Biol Fishes* 55:91-98
- Beets J, Rogers C (2000) Changes in fishery resources and reef fish assemblages in a Marine Protected Area in the US Virgin Islands: the need for a no take marine reserve. *Proceeding of the 9th International Coral Reefs Symposium* 1:449-454.
- Burton M, Brennen K, Munoz R, Parker Jr. R (2005) Preliminary evidence of increased spawning aggregations of mutton snapper (*Lutjanus analis*) at Riley's Hump two years after the establishment of the Tortugas South Ecological Reserve. *Fishery Bulletin* 103:404-410
- Busby RF, Bright CV, Pruna A (1966) Ocean bottom reconnaissance off the east coast of Andros Island, Bahamas U.S. Navy Oceanographic Office Technical Reports 20390. U.S. Naval Oceanographic Office, Washington, D.C.
- Calnan JM, Smith TB, Nemeth RS, Kadison E, Blondeau J (2008) Coral disease prevalence and host susceptibility on mid-depth and deep reefs in the United States Virgin Islands. *Revista de Biologia Tropical* (in press)
- Colin PL (1974) Observations and collection of deep-reef fishes off the coasts of Jamaica and British Honduras (Belize) *Marine Biology* 24:29-38

- CFMC (1996) Regulatory amendments to the fishery management of the reef fishery of PR and the USVI concerning red hind spawning aggregation, including regulatory management review and environmental assessment. Federal Register, Vol. 61 No. 235, December 5, 1996. p. 64485-64486
- Collazo JA (1980) Monitoring and assessment of commercial deepwater fishes at three locations near Puerto Rico. CODREMAR, Mayaguez, P.R. p. 37
- Cote IM, Mosqueira I, Reynolds JD (2001) Effects of marine reserve characteristics on the protection of fish populations: a meta-analysis. Journal of Fish Biology 59:178-189
- Culter JK, Ritchie KB, Earle SA, Guggenheim DE, Halley RB, Ciembronowicz KT, Hine AC, Jarrett BD, Locker SD, Jaap WC (2006) Pulley reef: a deep photosynthetic coral reef on the West Florida Shelf, USA. Coral Reefs 25:228-228
- Dammann A (1969) Study of the fisheries potential of the Virgin Islands. Special Report. Virgin Islands Ecological Research Station, University of the Virgin Islands, Caribbean Research Institute 197
- Davies GE (1982) A century of natural change in coral distribution at the Dry Tortugas: a comparison of reef maps from 1881 to 1976. Bulletin of Marine Science 32:608-623
- Dayton PK, Sala E, Tegner MJ, Thrush S (2000) Marine reserves: Parks, baselines, and fishery enhancement. Bulletin of Marine Science 66:617-634
- Dennis G, Hensley D, Colin PL, Kimmel J (2004) New records of marine fishes from the Puerto Rican plateau. Caribbean Journal of Marine Science 40:70-87
- Evans R, Russ G, Kritzer J (2008) Batch fecundity of *Lutjanus carponotatus* (Lutjanidae) and implications of no-take marine reserves on the Great Barrier Reef, Australia. Coral Reefs 27:179-189
- Franklin EC, Ault JS, Smith SG, Luo J, Meester GA, Diaz JM (2003) Benthic habitat mapping in the Tortugas region, Florida. Marine Geodesy 26:19-34
- Fricke H, Meischner D (1985) Depth limits of Bermudan scleractinian corals: A submersible survey. Marine Biology. Berlin, Heidelberg 88:175-187
- Garcia-Sais J, Castro R, Sabater J (2001) Coral reef communities from Natural Reserves in Puerto Rico: a baseline quantitative assessment for prospective monitoring programs. Vol. 1 - Cordillera de

Fajardo, Guanica, Bahia de Mayaguez, Caja de Muertos. U.S. Coral Reef Initiative Program, NOAA-DNER p. 232

Garcia-Sais J, Castro R, Sabater J, Carlo M (2001) Coral reef communities from Natural Reserves in Puerto Rico: a baseline quantitative assessment for prospective monitoring programs. Vol.2 - La Parguera, Boqueron, Isla de Mona, Isla Desecheo. U.S. Coral Reef Initiative Program, NOAA-DNER p. 193

Garcia-Sais J, Castro R, Sabater J, Carlo M (2001) Coral reef communities from Natural Reserves in Puerto Rico: a baseline quantitative assessment for prospective monitoring programs. Vol 3 - GuaGuayanilla, Ponce, Guayama, Arroyo. U.S. Coral Reef National Monitoring Program, NOAA-DNER p.108

Garcia-Sais J, Castro R, Sabater J, Carlo M (2001) Baseline characterization of coral reef and seagrass communities from Isla de Vieques, Puerto Rico U.S. Coral Reef National Monitoring Program. NOAA-DNER p.108

Garcia-Sais J, Appeldoorn R, Bruckner AW, Caldow C, Christensen JD, Lilyestrom M, Monaco ME, Sabater J, Williams E, Diaz E (2005) The State of Coral Reef Ecosystems of the Commonwealth of Puerto Rico. In: Waddell J (ed) The State of Coral Reef Ecosystems of the United States and Pacific Freely Associated States: NOAA Technical Memorandum NOS NCCOS 11. NOAA/NCCOS Center for Coastal Monitoring and Assessment's Biogeography Team. Silver Springs, MD.

Garcia-Sais J, Castro R, Sabater J, Carlo M (2005) Inventory and atlas of corals and coral reefs from the U.S. Caribbean EEZ (Puerto Rico and the United States Virgin Islands). CFMC/NOAA p. 215

Gardner J, Mayer L, Clarke J, Keiner A (1998) High-resolution multibeam bathymetry of East and West Flower Gardens and Stetson Banks, Gulf of Mexico. Gulf of Mexico Science XVI:128

Gilnack ML (1987) Deep water coral reef algal turfs: Analysis of community structure and biomass. Masters thesis, Southeastern Massachusetts University, p 65

Ginsburg RN, Harris PM, Eberli GP, Swart PK (1991) The growth potential of a bypass margin, Great Bahama Bank. Journal of Sedimentary Petrology 61:976-987

- Glynn PW (1973) Aspects of ecology of coral reefs in the Western Atlantic region. In: Jones OA, Endean R (eds) *Biology and Geology of Coral Reefs*. Volume II: Biology 1. Academic Press, New York, pp 480
- Graham NAJ, Evans RD, Russ GR (2003) The effects of marine reserve protection on the trophic relationships of reef fishes on the Great Barrier Reef. *Environmental Conservation* 30:200-208
- Grober-Dunsmore R, Frazer TK, Beets J, Lindberg WJ, Zwick P, Funniceilli N (2008) Influence of landscape structure on reef fish assemblages. *Landscape Ecology* 23:37-53
- Grober-Dunsmore R, Frazer TK, Lindberg WJ, Beets J (2007) Reef fish and habitat relationships in a Caribbean seascape: the importance of reef context. *Coral Reefs* 26:201-216
- Halpern BS (2003) The impact of marine reserves: Do reserves work and does reserve size matter? *Ecological Applications* 13:S117-S137
- Hamilton SL, Regetz J, Warner RR (2008) Postsettlement survival linked to larval life in a marine fish. *Proceedings of the National Academy of Sciences*. 105 (5): 1561-1566
- Herzlieb S, Kadison E, Blondeau J, Nemeth RS (2005) Comparative assessment of coral reef systems located along the insular platform south of St. Thomas, US Virgin Islands and the relative effects of natural and human impacts. *Proc 10th Int Coral Reef Symp* 4-2:1144-1151.
- Hubbard DK, Scaturo D (1985) Growth rates of seven species of scleractinian corals from Cane Bay and Salt River, St. Croix, USVI. *Bulletin of Marine Science* 36:325-338
- Johnson KG, Jackson JBC, Budd AF (2008) Caribbean reef development was independent of coral diversity over 28 million years. *Science* 319:1521-1523
- Jones GP, McCormick MI, Srinivasan M, Eagle JV (2004) Coral decline threatens fish biodiversity in marine reserves. *Proceedings of the National Academy of Sciences* 101:8251-8253
- Jordan-Dahlgren E, Maldonado MA, Rodriguez-Martinez RE (2005) Diseases and partial mortality in *Montastraea annularis* species complex in reefs with differing environmental conditions (NW Caribbean and Gulf of Mexico). *Diseases of Aquatic Organisms* 63:3-12

- Kadison E, Nemeth R, Herzlieb S, Blondeau J (2006) Temporal and spatial dynamics of *Lutjanus cyanopterus* and *L. jocu* (Pisces: *Lutjanidae*) spawning aggregations on a multi-species spawning site in the USVI. *Revista de Biologia Tropical* 54 (suppl. 3):69-78
- Kadison E, Nemeth RS, Blondeau J (In Revision) Investigation of a red hind (*Epinephelus guttatus*) spawning aggregation on Saba Bank in Netherland Antilles. *Bulletin of Marine Science*
- Kendall MS, Eschelbach K (2006) Spatial-analysis of the benthic habitats within the limited-use zones around Vieques, Puerto Rico. *Bulletin of Marine Science* 79:389-400
- Kiraly S, Moore J, Jasinski P (2003) Deepwater and other sharks of the U.S. Atlantic Ocean Exclusive Economic Zone. *Marine Fisheries Review*. Fall 2003.
- Koenig C, Coleman F, Grimes C, Fitzhugh G, Scanlon K, Gledhill C, Grace M (2000) Protecting of fish-spawning habitat for the conservation of warm-temperate reef-fish fisheries on the shelf edge reefs of Florida. *Bulletin of Marine Science* 66:593-616
- Koenig C, Shepard A, Reed J, Coleman F, Brooke S, Brusher J, Scanlon K (2005) Habitat and fish populations in the deep-sea *Oculina* coral ecosystem of the western Atlantic. *American Fisheries Society Symposium* 41
- Kumpf HE, Randall HA (1961) Charting the marine environments of St. John, U.S. Virgin Islands. *Bulletin of Marine Science of the Gulf and Caribbean* 11:543-551
- Lang J (1974) Biological zonation at the base of a reef. *American Scientist* 62:272-281
- Lang J, Lasker HR, Gladfelter EH, Hallock P, Jaap WC, Losada FJ, Muller RG (1992) Spatial and Temporal Variability during Periods of "Recovery" after Mass Bleaching on Western Atlantic Coral Reefs. *American Zoologist* 32:696-706
- Lang JC, Hartman WC, Land LS (1975) Sclerosponges: primary framework constructors on the Jamaican deep fore-reef. *Journal of Marine Research* 33:223-231
- Leichter JJ, Genovese SJ (2006) Intermittent upwelling and subsidized growth of the scleractinian coral *Madracis mirabilis* on the deep fore-reef of Discovery Bay, Jamaica. *Marine Ecology Progress Series* 316:95-103

- Lewis SM (1986) The role of herbivorous fishes in the organization of a Caribbean reef community. *Ecological Monographs* 56:183-200
- Manzello DP, Brandt M, Smith TB, Lirman D, Hendee JC, Nemeth RS (2007) Hurricane-associated cooling benefits bleached corals. *Proceedings of the National Academy of Science* 104:12035-12039
- Matos C, Garcia-Sais J, Diaz E (2000) Puerto Rico's Coral Reefs - Status and Trends Report. Department of Natural and Environmental Resources. Commonwealth of Puerto Rico 41
- McClanahan T (2008) Response of the coral reef benthos and herbivory to fishery closure management and the 1998 ENSO disturbance. *Oecologia* 155:169-177
- McClanahan TR, Graham NAJ, Calnan JM, MacNeil MA (2007) Toward pristine biomass: Reef fish recovery in coral reef marine protected areas in Kenya. *Ecological Applications* 17:1055-1067
- Melvin G, Cochrane N, Li Y (2003) Extraction and comparison of acoustic backscatter from a calibrated multi- and single-beam sonar. *ICES Journal of Marine Science* 60:669-677
- Menza C, Kendall M, Rogers C, Miller J (2007) A deep reef in deep trouble. *Continental Shelf Research* 27:2224-2230
- Micheli F, Halpern BS, Botsford LW, Warner RR (2004) Trajectories and correlates of community change in no-take marine reserves. *Ecological Applications* 14:1709-1723
- Miller SL, Chiappone M, Swanson DW, Ault JS, Smith SG, Meester GA, Luo J, Franklin EC, Bohnsack JA, Harper DE, McClellan DB (2001) An extensive deep reef terrace on the Tortugas Bank, Florida Keys National Marine Sanctuary. *Coral Reefs* 20:299-300
- Mosqueira I, Cote IM, Jennings S, Reynolds JD (2000) Conservation benefits of marine reserves for fish populations. *Animal Conservation* 3:321-332
- Mumby PJ, Skirving W, Strong AE, Hardy JT, LeDrew EF, Hochberg EJ, Stumpf RP, David LT (2004) Remote sensing of coral reefs and their physical environment. *Marine pollution bulletin* 48:219-228
- Nagelkerken I, van der Velde G, Cocheret de la Morinière E (2001) Fish feeding guilds along a gradient of bay biotopes and coral reef depth zones. *Aquatic Ecology* 35:73-86

- Nemeth R, Quandt A, Requa L, Rothenberger J, Taylor M (2003) A rapid assessment of coral reefs in the Virgin Islands (Part I: stony corals and algae). *Atoll Research Bulletin* 496:544-565
- Nemeth RS, Whaylen LD, Pattengill-Semmens C (2003) A rapid assessment of coral reefs in the Virgin Islands (Part 2: fishes). *Atoll Research Bulletin* 496:566-589
- Nemeth RS (2005) Population characteristics of a recovering US Virgin Islands red hind spawning aggregation following protection. *Marine Ecology Progress Series* 286:81-97
- Nemeth RS, Herzlieb S, Blondeau J (2006) Comparison of two seasonal closures for protecting red hind spawning aggregations in the US Virgin Islands. *Proc. 10th International Coral Reef Conference* 4:1306-1313.
- Nemeth RS, Kadison E, Herzlieb S, Blondeau J, Whiteman E (2006) Status of a yellowfin grouper (*Mycteroperca venenosa*) spawning aggregation in the US Virgin Islands with notes on other species. *Proc. 57th Gulf Carib Fish Inst.* 57:543-558.
- Nemeth RS, Blondeau J, Herzlieb S, Kadison E (2007) Spatial and temporal patterns of movement and migration at spawning aggregations of red hind, *Epinephelus guttatus*, in the US Virgin Islands. *Environmental Biology of Fishes* 78:365-381
- Nemeth RS, Kadison E, Blondeau J, Idrisi N, Watlington R, Brown K, Smith T, Carr L (2008) Regional coupling of red hind spawning aggregations to oceanographic processes in the Eastern Caribbean. NOAA Special Publications. In press
- Nemeth RS (In Review) Dynamics of reef fish and decapod crustacean spawning aggregations: underlying mechanisms, habitat linkages and trophic interactions. In: Negelkerken I (ed) *Ecological interactions among tropical coastal ecosystems*. Springer
- Newman M, Paredes G, Sala E, JBC J (2006) Structure of Caribbean coral reef communities across a large gradient of fish biomass. *Ecology Letters* 9:1216-1227
- Nichols J (1929) The Fishes of Puerto Rico and the Virgin Islands: Branchiostomidae to Sciaenidae. *Scientific Survey of Puerto Rico and the Virgin Islands*. New York Acad. Sci. 10:161-295
- NOAA (2002) Tortugas Ecological Reserve: Final Supplemental Environmental Impact Statement/Final Supplemental Management Plan. NOAA, Silver Spring, MD

- NRC (2001) Marine protected areas: tools for sustaining ocean ecosystems. National Academy Press, Washington, D.C.
- Olsen D, LaPlace J (1978) A study of Virgin Islands grouper fishery based on a breeding aggregation. Proc 31st Gulf Carib Fish Inst 31:130-144.
- Pantos O, Cooney RP, Le Tissier MDA, Barer MR, O'Donnell AG, Bythell JC (2003) The bacterial ecology of a plague-like disease affecting the Caribbean coral *Montastrea annularis*. Environmental Microbiology 5:370-382
- Parnell PE, Dayton PK, Lennert-Cody CE, Rassmussen LL, Leichter JJ (2006) Marine Reserve Design: Optimal size, habitats, species affinities, diversity, and ocean microclimate. Ecological Applications 16:945-962
- Pittman S, Christensen JD, Caldow C, Menza C, Monaco ME (2007) Predictive mapping of fish species richness across shallow-water seascapes in the Caribbean. Ecological Modeling 204:9-21
- Polunin NVC, Roberts CM (1993) Greater biomass and value of target coral-reef fishes in 2 small Caribbean marine reserves. Marine Ecology-Progress Series 100:167-176
- Quinn N, Kojis B (1999) Subsurface seawater temperature variation and the recovery of corals from the 1993 coral bleaching event in waters off St. Thomas, U. S. Virgin Islands. Bulletin of Marine Science 65:210-214
- Rakitin A, Kramer DL (1996) Effect of a marine reserve on the distribution of coral reef fishes in Barbados. Marine Ecology-Progress Series 131:97-113
- Randall JE (1982) Tropical marine sanctuaries and their significance in reef fisheries research. In: Huntsman GR, Nicholson WR, Fix WWJ (eds) Workshop on Biological Bases for Reef Fishery Management, St. Thomas (USVI), 7 Oct 1980; the biological bases for reef fishery management. proceedings of a workshop held October 7-10, 1980 at St. Thomas, Virgin Islands of the United States. 167-178
- Reed J (1980) Distribution and structure of deep-water *Oculina vericosa* coral reefs off central eastern Florida. Bulletin of Marine Science 30:667-677

- Riegl B, Piller WE (2003) Possible refugia for reefs in times of environmental stress. *International Journal of Earth Sciences* 92:520-531
- Rivera JA, Prada MC, Arsenault JL, Moody G, Benoit N (2005) Detecting fish aggregations from reef habitats mapped with high resolution side scan sonar imagery. National Marine Fisheries Service Professional Paper 5:88-104
- Roberts C, Bohnsack J, Gell F, Hawkins J, Goodridge T (2001) Effects of marine reserves on adjacent fisheries. *Science* 294:1920-1923
- Roberts CM (1995) Rapid buildup of fish biomass in a Caribbean marine reserve. *Conservation Biology* 9:815-826
- Rodwell LD, Barbier EB, Roberts CM, McClanahan TR (2003) The importance of habitat quality for marine reserve - fishery linkages. *Canadian Journal of Fisheries and Aquatic Sciences* 60:171-181
- Rogers A (1999) The biology of *Lophelia pertusa* (Linnaeus, 1758) and other deep-water reef-forming corals and impacts from human activities. *International Review of Hydrobiology* 84:315-406
- Rogers C, Miller J, Muller E, Edmunds PJ, Nemeth RS, Beets J, Friedlander AM, Smith T, Boulon R, Jeffery C, Menza C, Caldow C, Idrisi N, Kojis B, Monaco ME, Spitzack T, Gladfelter B, Ogden J, Hillis-Starr Z, Lundgren I, Bane Schill W, Kuffner IB, Richardson L, Devine B, Voss JD (2008) Coral Reefs of the US Virgin Islands. In: Dodge R, Reigl B (eds) *Coral Reefs of the United States*.
- Rosario A (1986) Survey of commercially exploited fish species and exploratory fishing of underutilized resources around Puerto Rico. National Marine Fishery Service, NOAA 128
- Rosen BR (2000) Platy coral assemblages: 200 million years of functional stability in response to the limiting effects of light and turbidity. *Proc 9th Int Coral Reef Symp* 1:255-264.
- Russ GR, Alcalá AC (1996) Marine reserves: Rates and patterns of recovery and decline of large predatory fish. *Ecological Applications* 6:947-961
- Singh H (2003) New imaging vehicle maps of coral reefs to determine health of reef and fisheries. Woods Hole Oceanographic Institute

- Ruyter van Steveninck E, Breeman A (1987) Deep water vegetations of *Lobophora variegata* (Phaeophyceae) in the coral reef of Curaçao: population dynamics in relation to mass mortality of the sea urchin *Diadema antillarum*. Marine Ecology Progress Series 36:81-90
- Sale PF (ed.) (1991) The Ecology of Fishes on Coral Reefs. Academic Press Inc. New York
- Salm RV, Coles SL (2001) Coral bleaching and marine protected areas. Proceedings of the workshop on mitigating coral bleaching impact through MPA design. The Nature Conservancy, Honolulu.
<http://www.conserveonline.org>
- Sanchez Lizaso JL, Goni R, Renones O, Garcia Charton JA, Galzin R, Bayle JT, Sanchez Jerez P, Perez Ruzafa A, Ramos AA (2000) Density dependence in marine protected populations: a review. Environmental Conservation 27:144-158
- Santodomingo R, Gracia N, Borrero G, Mejia-Ladino M, Bermudez A, Benavides M (2003) Biodiversity survey of south Caribbean deep sea communities. INVEMAR Abstract, ISDSC Poster Presentation
- Shinn E (1980) Geological reconnaissance dive using the Johnson Sea-Link submersible in Key Largo Coral Reef Marine Sanctuary. NOAA
- Shulman MJ, Bermingham E (1995) Early life histories, ocean currents, and the population genetics of Caribbean reef fishes. Evolution 49:897-910
- Silvester J, Dammann A (1974) Some observations on the deepwater fishery resources of the Virgin Islands. Caribbean Journal of Marine Science 14 163-165
- Singh H (2003) New imaging vehicle maps of coral reefs to determine health of reef and fisheries. Woods Hole Oceanographic Institute
- Singh H, Armstrong R, Gibles F, Eustice R, Roman CT, Pizarro O, Torres J (2004) Imaging coral I: Imaging coral habitats with the Seabed AUV. Subsurface Sensing Technologies and Applications 5:25-42
- Smith TB, Nemeth RS, Calnan JM, Taylor M, Kadison ES, Blondeau J, Tyner E (2007) Coral reef monitoring in St. Croix and St. Thomas, United States Virgin Islands. Year six final report

submitted to Department of Planning and Natural Resources. University of the Virgin Islands, St. Thomas 52

St Mary CM, Osenberg CW, Frazer TK, Lindberg WJ (2000) Stage structure, density dependence and the efficacy of marine reserves. *Bulletin of Marine Science* 66:675-690

van den Hoek C, Breeman AM, Bak RPM, van Buurt G (1978) The distribution of algae, corals and gorgonians in relation to depth, light attenuation, water movement and grazing pressure in the fringing coral reef of Curacao, Netherlands Antilles. *Aquatic Botany* 5: 1-46

Vaughan T (1900) The stony corals of Porto Rican waters. Investigations of the Aquatic Resources and Fisheries of Porto Rico by the U.S. Fish Commission Steamer Fish Hawk in 1899. *Bulletin of U.S. Fish Commission* 20:289-320

Vermeij GJ, Fogarty ND, Miller MW (2006) Pelagic conditions affect larval behavior, survival, and settlement patterns in the Caribbean coral *Montastraea faveolata*. *Marine Ecology Progress Series* 310:119-128

Vermeij MJA (2005) Substrate composition and adult distribution determine recruitment patterns in a Caribbean brooding coral. *Marine Ecology Progress Series* 295:123-133

Vermeij M, Bak RPM (2003) Species-specific population structure of closely related coral morphospecies along a depth gradient (5-60m) over a Caribbean reef slope. *Bulletin of Marine Science* 73:725-744

Villinski JT (2003) Depth-independent reproductive characteristics for the Caribbean reef-building coral *Montastraea faveolata*. *Marine Biology* 142:1043-1053

Vize P (2006) Deepwater broadcast spawning by *Montastraea cavernosa*, *Montastraea franksi*, and *Diploria strigosa* at the Flower Garden Banks, Gulf of Mexico. *Coral Reefs* 25:169-171

Weil E, Knowlton N (1994) A multi-character analysis of the Caribbean coral species *Montastraea annularis* (Ellis and Solander, 1786), and its two sibling species, *M. faveolata* (Ellis and Solander, 1786) and *M. franksi* (Gregory, 1895). *Bulletin of Marine Science* 55:151-175

Wellington GM (1982) Depth zonation of corals in the Gulf of Panama: control and facilitation by resident reef fishes. *Ecological Monographs* 52:223-241